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<u>COTK 14/445</u>





THE PATENT COOPERATION TREATY (PCT)

ernati nal Publication Number:

WO 96/40766

C07K 14/445, C12N 15/30, A61K 39/015

(43) Internati nal Publication Date:

19 December 1996 (19.12.96)

(21) International Application Number:

PCT/US96/09508

(22) International Filing Date:

7 June 1996 (07.06.96)

(30) Priority Data:

08/487,826

7 June 1995 (07.06.95)

US

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- (81) Designated States: AL, AM, AT, AU, AZ, BB, BG, BR, BY, CA, CH, CN, CZ, DE, DK, EE, ES, FI, GB, GE, HU, IL, IS, JP, KE, KG, KP, KR, KZ, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, TJ, TM, TR, TT, UA, UG, UZ, VN, ARIPO patent (KE, LS, MW, SD, SZ, UG), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG).

Published

Without international search report and to be republished upon receipt of that report.

(54) Title: BINDING DOMAINS FROM PLASMODIUM VIVAX AND PLASMODIUM FALCIPARUM ERYTHROCYTE BINDING **PROTEINS**

(57) Abstract

The present invention provides isolated polypeptides useful in the treatment and prevention of malaria caused by Plasmodium falciparum or P. vivax. In particular, the polypeptides are derived from the binding domains of the proteins in the DBL family as well as the sialic acid binding protein (SABP) on P. falciparum merozoites. The polypeptides may also be derived from the Duffy antigen binding protein (DABP) on P. vivax merozoites.

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BINDING DOMAINS FROM PLASMODIUM VIVAX AND PLASMODIUM FALCIPARUM ERYTHROCYTE BINDING PROTEINS

BACKGROUND OF THE INVENTION

Malaria infects 200 - 400 million people each year causing 1-2 million deaths, thus remaining one of the most important infectious diseases in the world. Approximately 25 percent of all deaths of children in rural Africa between the ages of one and four years are caused by malaria. Due to the importance of the disease as a worldwide health problem, considerable effort is being expended to identify and develop malaria vaccines.

Malaria in humans is caused by four species of the parasite *Plasmodium: P. falciparum, P. vivax, P. knowlesi* and *P. malariae*. The major cause of malaria in humans is *P. falciparum* which infects 200 million to 400 million people every year, killing 1 to 4 million.

Duffy Antigen Binding Protein (DABP) and Sialic Acid Binding Protein (SABP) are soluble proteins that appear in the culture supernatant after infected erythrocytes release merozoites. Immunochemical data indicate that DABP and SABP which are the respective ligands for the *P. vivax* and *P. falciparum* Duffy and sialic acid receptors on erythrocytes, possess specificities of binding which are identical either in soluble or membrane bound form.

DABP is a 135 kDa protein which binds specifically to Duffy blood group determinants (Wertheimer et al., Exp. Parasitol. 69: 340-350 (1989); Barnwell, et al., J. Exp. Med. 169: 1795-1802 (1989)). Thus, binding of DABP is specific to human Duffy positive erythrocytes. There are four major Duffy phenotypes for human erythrocytes: Fy(a), Fy(b), Fy(ab) and Fy(negative), as defined by the anti-Fy^a and anti-Fy^b sera (Hadley et al., In Red Cell Antigens and Antibodies, G. Garratty, ed. (Arlington, Va.:American Association of Blood Banks) pp. 17-33 (1986)). DABP binds equally to both Fy(a) and Fy(b) erythrocytes which are equally susceptible to invasion by P. vivax; but not to Fy(negative) erythrocytes.

In the case of SABP, a 175kDa protein, binding is specific to the glycophorin sialic acid residues on erythrocytes (Camus and Hadley, *Science* 230:553-556 (1985); Orlandi, et al., J. Cell Biol. 116:901-909 (1992)). Thus, neuraminidase treatment (which cleaves off sialic acid residues) render erythrocytes immune to *P. falciparui. invasion*.

The specificities of binding and correlation to invasion by the parasite thus indicate that DABP and SABP are the proteins of *P. vivax* and *P. falciparum* which interact with sialic acids and the Duffy antigen on the erythrocyte. The genes encoding both proteins have been cloned and the DNA and predicted protein sequences have been determined (B. Kim Lee Sim, *et al.*, *J. Cell Biol.* 111: 1877-1884 (1990); Fang, X., *et al.*, *Mol. Biochem Parasitol.* 44: 125-132 (1991)).

Despite considerable research ettorts worldwide, because of the complexity of the *Piasmodium* parasite and its interaction with its host, it has not been possible to discover a satisfactory solution for prevention or abatement of the blood stage of malaria. Because malaria is a such a large worldwide health problem, there is a need for methods that abate the impact of this disease. The present invention provides effective preventive and therapeutic measures against *Plasmodium* invasion.

SUMMARY OF THE INVENTION

The present invention provides compositions comprising an isolated DABP binding domain polypeptides and/or isolated SABP binding domain polypeptides. The DABP binding domain polypeptides preferably comprise between about 200 and about 300 amino acid residues while the SABP binding domain polypeptides preferably comprises between about 200 and about 600 amino acid residues. A preferred DABP binding domain polypeptide has about 325 residues of the amino acid sequence found in SEQ ID NO:2. A preferred SABP binding domain polypeptide has about 616 residues of the amino acid sequence of SEQ ID NO:4, encoded by the DNA sequence of SEQ ID NO: 3. The preferred DABP binding domain and SABP binding domain include the cysteine-rich portions of the proteins shown in Figure 1.

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The present invention also includes pharmaceutical compositions comprising a pharmaceutically acceptable carrier and an isolated DABP binding domain polypeptide in an amount sufficient to induce a protective immune response to *Plasmodium vivax* merozoites in an organism. In addition, isolated SABP binding domain polypeptide in an amount sufficient to induce a protective immune response to *Plasmodium falciparum* may be added to the pharmaceutical composition.

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Also provided are pharmaceutical compositions comprising a pharmaceutically acceptable carrier and an isolated SABP binding domain polypeptide in an amount sufficient to induce a protective immune response to *Plasmodium falciparum* merozoites in an organism. In addition, isolated DABP binding domain polypeptide in an amount sufficient to induce a protective immune response to *Plasmodium vivax* may be added to the pharmaceutical composition.

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Isolated polynucleotides which encode a DABP binding domain polypeptides or SABP binding domain polypeptides are also disclosed. In addition, the present invention includes a recombinant cell comprising the polynucleotide encoding the DABP binding domain polypeptide.

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The current invention further includes methods of inducing a protective immune response to Plasmodium merozoites in a patient. The methods comprise administering to the patient an immunologically effective amount of a pharmaceutical composition comprising a pharmaceutically acceptable carrier and an isolated DABP binding domain polypeptide, an SABP binding domain polypeptide or a combination thereof.

The present disclosure also provides DNA sequences from additional *P. falciparum* genes in the Duffy-binding like (*DBL*) family that have regions conserved with the *P. falciparum* 175 kD and *P. vivax* 135 kD binding proteins.

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DEFINITIONS

As used herein a "DABP binding domain polypeptide" or a "SABP binding domain polypeptide" are polypeptides substantially identical (as defined below) to a sequence from the cysteine-rich, amino-terminal region of the Duffy antigen binding protein (DABP) or sialic acid binding protein (SABP), respectively. Such polypeptides are capable of binding either the Duffy antigen or sialic acid residues on glycophorin. In particular, DABP binding domain polypeptides consist of amino acid residues substantially similar to a sequence of SABP within a binding domain

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containing the cysteine-rich sequence shown in Figure 1. SABP binding domain polypeptides consist of residues substantially similar to a sequence of DABP within a binding domain containing the cysteine-rich sequence shown in Figure 1.

The binding domain polypeptides encoded by the genes of the *DBL* family consist of those residues substantially identical to the sequence of the binding domains of DABP and SABP as defined above. The DBL family comprises sequences with substantial similarity to the conserved regions of the DABP and SABP. These include those sequences reported here as *ebl-1* (SEQ ID NO:5 and SEQ ID NO:6), E31a (SEQ ID NO:7 and SEQ ID NO:8), *var-7* (SEQ. ID. NO:13 and SEQ. ID. NO:14, GenBank Accession No. L42636) and *var-1* (SEQ. ID. NO:15 and SEQ ID NO:16, GenBank Accession No. L40608). The sequence *ebl-2*, (SEQ ID NO:9 and SEQ ID NO:10) represents the binding domains of *var-7*, and Proj3 (SEQ ID NO:11 and SEQ ID NO:12) is the binding domain of *var-1*. The DBL family also includes two other members *var-2* and *var-3* (GenBank Accession No. L40609).

The polypeptides of the invention can consist of the full length binding domain or a fragment thereof. Typically DABP binding domain polypeptides will consist of from about 50 to about 325 residues, preferably between about 75 and 300, more preferably between about 100 and about 250 residues. SABP binding domain polypeptides will consist of from about 50 to about 616 residues, preferably between about 75 and 300, more preferably between about 100 and about 250 residues.

Particularly preferred polypeptides of the invention are those within the binding domain that are conserved between SABP and the *DBL* family. Residues within these conserved domains are shown in Figure 1, below.

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Two polynucleotides or polypeptides are said to be "identical" if the sequence of nucleotides or amino acid residues in the two sequences is the same when aligned for maximum correspondence. Optimal alignment of sequences for comparison may be conducted by the local homology algorithm of Smith and Waterman Adv. Appl. Math. 2: 482 (1981), by the homology alignment algorithm of Needleman and Wunsch J. Mol. Biol. 48:443 (1970), by the search for similarity method of Pearson and Lipman Proc. Natl. Acad. Sci. (U.S.A.) 85: 2444 (1988), by computerized implementations of these algorithms (GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group, 575 Science Dr., Madison, WI), or by inspection. The term "substantial identity" means that a polypeptide comprises a sequence that has at least 80% sequence identity, preferably 90%, more preferably 95% or more, compared to a reference sequence over a comparison window of about 20 residues to about 600 residues- typically about 50 to about 500 residues usually about 250 to 300 residues. The values of percent identity are determined using the programs above. Particularly preferred peptides of the present invention comprise a sequence in which at least 70% of the cysteine residues conserved in DARP and SABP are present. Additionally, the peptide will comprise a sequence in which at least 50% of the tryptophan residues conserved in DABP and SABP are present. The term substantial similarity is also specifically defined here with respect to those amino acid residues found to be conserved between DABP, SABP and the sequences of the DBL family. These conserved amino acids consist prominently of tryptophan and cysteine residues conserved among all sequences reported here. In addition the conserved amino acid residues include phenylalanine residues which may

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be substituted with tyrosine. These amino acid residues may be determined to be conserved after the sequences have been aligned using methods outlined above by someone skilled in the art.

Another indication that polypeptide sequences are substantially identical is if one protein is immunologically reactive with antibodies raised against the other protein. Thus, the polypeptides of the invention include polypeptides immunologically reactive with antibodies raised against the SABP binding domain, the DABP binding domain or raised against the conserved regions of the DBL family.

Another indication that nucleotide sequences are substantially identical is if two molecules hybridize to each other under stringent conditions. Stringent conditions are sequence dependent and will be different in different circumstances. Generally, stringent conditions are selected to be about 5° C lower than the thermal melting point (Tm) for the specific sequence at a defined ionic strength and pH. The Tm is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. Typically, stringent conditions will be those in which the salt concentration is about 0.02 molar at pH 7 and the temperature is at least about 60°C.

Nucleotide sequences are also substaintially identical for purposes of this application when the polypeptides which they encode are substantially identical. Thus, where one nucleic acid sequence encodes essentially the same polypeptide as a second nucleic acid sequence, the two nucleic acid sequences are substantially identical, even if they would not hybridize under stringent conditions due to silent substitutions permitted by the genetic code (see, Darnell et al. (1990) Molecular Cell Biology, Second Edition Scientific American Books, W.H. Freeman and Company, New York, NY, for an explanation of codon degeneracy and the genetic code).

The phrases "isolated" or "biologically pure" refer to material which is substantially or essentially free from components which normally accompany it as found in its native state. Thus, the binding domain polypeptides of this invention do not contain materials normally associated with their *in situ* environment, e.g., other proteins from a merozoite membrane. Typically, isolated proteins of the invention are at least about 80% pure, usually at least about 90%, and preferrably at least about 95% as measured by band intensity on a silver stained gel.

Protein purity or homogeneity may be indicated by a number of means well known in the art, such as polyacrylamide gel electrophoresis of a protein sample, followed by visualization upon staining. For certain purposes high resolution will be needed and HPLC or a similar means for purification utilized.

The term "residue" refers to an amino acid (D or L) or amino acid mimetic incorporated in a oligopeptide by an amide bond or amide bond mimetic. An amide bond mimetic of the invention includes peptide backbone modifications well known to those skilled in the art.

BRIEF DESCRIPTION OF THE DRAWINGS

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Figure 1 represents an alignment of the predicted amino acid sequences of the DABP binding domain (Vivax) (SEQ ID NO:25), the two homologous SABP domains (SABP F1 (SEQ ID NO:26) and SABP F2 (SEQ ID NO:27)) and the sequenced members of the *DBL* gene family (ebl-1 (SEQ ID NO:28), E31a (SEQ ID NO:39), EBL-2 (SEQ ID NO:30)) and the three homologous Proj3 domains (F1 (SEQ ID NO:31), F2 (SEQ ID NO:32) and F3 (SEQ ID NO:33)).

Figure 2 represents a schematic of the pRE4 cloning vector.

Figure 3 shows primers useful for isolating sequences encoding the conserved motifs of the invention. Primers UNIEBP5 (SEQ ID NO:35) and UNIEBP5A (SEQ ID NO:36) encode the amino acid sequence of SEQ ID NO:34; primers UNIEBP5B (SEQ ID NO:38) and UNIEBP5C (SEQ ID NO:39) encode the amino acid sequence of SEQ ID NO:37; primers UNIEBP3 (SEQ ID NO:41) and UNIEBP3A (SEQ ID NO:42) encode the amino acid sequence of SEQ ID NO:40; and primers UNIEBP3B (SEQ ID NO:44) and UNIEBP3C (SEQ ID NO:45) encode the amino acid sequence of SEQ ID NO:43.

Figure 4 shows the relative position of the E31a ORF on chromosome 7.

Figure 5 shows a map of a *var* gene cluster on chromosome 7. Relative positions of four YACs (PfYEF2, PfYFE6, PfYKF8, PfYED9) are indicated under the chromosome 7 line at the top of the figure. YACs PfYFE6 and PfYKF8 lie entirely within a segment linked to COR in a genetic cross, whereas YACs PfYED9 and PfYEF2 extend beyond sites (identified by pE53a and pH270.5) that are dissociated from the chloroquine response. The *var* cluster extends over a region of 100-150 kb in PfYED9. Exons and introns of the *var-1*, *var-2* and *var-3* genes within the sequenced 40 kb segment are represented by solid and dotted lines, respectively; arrows show the coding direction. Two more *var* elements outside of the sequenced region, identified by conserved restriction sites and cross-hybridization, are indicated by dashed-lines (*var-2c* and *var-3c*). Bold letters mark repeated restriction sites that suggest a duplication in the *var-2/var-3* and *var-2c/var-3c* segments. Enzyme recognition sites: A, *Apal*; B, *BgA*; C, *Clal*; D, *Hind*III; E, *Hae*III; H, *Bss*HII; K, *Kpn*I; M, *Bam*HI; P, *Hpal*; S, *Smal*. *Hind*III and *Hae*III sites outside of the sequenced region were not mapped. Positions and sizes of inserts from the Dd2 subsegment library are indicated: a, pE280b; b, pB20.3; c, pB600; d, pE21b; e, pB20.24; f, pE32b; h, pE241a; i, pE240a/51d; j, pE33a; k, pB20.23; l, *A*L17BA6; m, pB20.26; n, pB20SU.27; o, p15J2J3. Inserts from the PfYED9 34 kb *Apal-Smal* fragment library: r, pB3; s, p3G11; t, pJVs; u, p2E10; v, pIG3; w, p2E3; x, p2B6; y, PE10; z, pJYr; α, pC5; β, p1A3; γ, p1F6; δ, p3C3; ε, pA2; ζ, p2A9; η, p3C4; θ, pJZn; κ, p3D8.

DESCRIPTION OF THE PREFERRED EMBODIMENT

The binding of merozoites and schizonts to erythrocytes is mediated by specific binding proteins on the surface of the merozoite or schizont and is necessary for erythrocyte invasion. In the case of *P. felciparum*, this binding involves specific interaction between sialic acid glycophorin residues on the erythrocyte and the sialic acid binding protein (SABP) on the surface of the merozoite or schizont. The ability of purified SABP to bind erythrocytes with chemically or enzymatically altered sialic acid residues paralleled the ability of *P. falciparum* to invade these erythrocytes. Furthermore, sialic acid deficient erythrocytes neither bind SABP nor support invasion by *P. falciparum*. The DNA encoding SABP from *P. falciparum* has also been cloned and sequenced.

In *P. vivax*, specific binding to the erythrocytes involves interaction between the Duffy blood group antigen on the erythrocyte and the Duffy antigen binding protein (DABP) on the merozoite. Duffy binding proteins were defined biologically as those soluble proteins that appear in the culture supernatant after the infected erythrocytes release merozoites which bind to human Duffy positive, but not to human Duffy negative erythrocytes. It has been shown that binding of the *P. vivax* DABP protein to Duffy positive erythrocytes is blocked by antisera to the Duffy blood group determinants. Purified Duffy blood group antigens also block the binding to erythrocytes. DABP has also been shown to bind Duffy blood group determinants on Western blots.

Duffy positive blood group determinants on human erythrocytes are essential for invasion of human erythrocytes by *Plasmodium vivax*. Both attachment and reorientation of *P. vivax* merozoites occur equally well on Duffy positive and negative erythrocytes. A junction then forms between the apical end of the merozoite and the Duffy-positive erythrocyte, followed by vacuole formation and entry of the merozoite into the vacuole. Junction formation and merozoite entry into the erythrocyte do not occur on Duffy negative cells, suggesting that the receptor specific for the Duffy determinant is involved in apical junction formation but not initial attachment. The DNA sequences encoding the DABP from *P. vivax* and *P. knowlesi* have been cloned and sequenced.

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P. vivax red cell invasion has an absolute requirement for the Duffy blood group antigen. Isolates of P. falciparum, however, vary in their dependency on sialic acid for invasion. Certain P. falciparum clones have been developed which invade sialic acid deficient erythrocytes at normal rates. This suggests that certain strains of P. falciparum can interact with other ligands on the erythrocyte and so may possess multiple erythrocyte binding proteins with differing specificities.

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A basis for the present invention is the discovery of the binding domains in both DABP and SABP. Comparison of the predicted protein sequences of DABP and SABP reveals an amino-terminal, cysteine-rich region in both proteins with a high degree of similarity between the two proteins. The amino-terminal, cysteine-rich region of DABP contains about 325 amino acids, whereas the amino-terminal, cysteine-rich region of SABP contains about 616 amino acids. This is due to an apparent duplication of the amino-terminal, cysteine-rich region in the SABP protein. The cysteine residues are conserved between the two regions of SABP and DABP, as are the amino acids surrounding the cysteine residues and a number of aromatic amino acid residues in this region. The amino-terminal cysteine rich region and another cysteine-rich region near the carboxyl-terminus show the most similarity between the DABP and SABP proteins. The region of the amino acid sequence between these two cysteine-rich regions show only limited similarity between DABP and SABP.

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Other *P. falciparum* open reading frames and genes with regions that have substantial identity to binding domains of SABP and DABP have been identified. Multiple copies of these sequences exist in the parasite genome, indicating their important activity in host-parasite interactions. A family of these sequences (the *DBL* family) have been cloned from chromosome 7 subsegment libraries that were constructed during genetic studies of the chloroquine resistance locus (Wellems *et. al.*, *PNAS* 88: 3382-3386 (1991)). Certain of these transcripts are known to be from the *var* family of genes that modulate cytoadherence and antigenic variation of *P. falciparum*-infected erythrocytes (*see*, Example 3, below).

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Genes of the *P. falciparum* var family encode 200-350 kD variant surface molecules that determine antigenic and adhesive properties of parasitized erythrocytes. The large repertoire of var genes (50-150 copies, having sufficient DNA to account for 2-6% of the haploid genome), the dramatic sequence variation among-the gene copies, their variable expression in different parasite lines, the ready detection of DNA rearrangements, and the receptor binding features of the encoded extracellular domains all implicate var genes as the major determinants of antigenic variation and cytoadherence in *P. falciparum* malaria.

A second class of *DBL*-encoding transcripts includes single-copy genes such as *ebl-1*. Genetic linkage studies have placed this gene within a region of chromosome 13 that affects invasion of malarial parasites in human red blood cells (Wellems *et al.*, *Cell* 49:633-642 (1987)). Both SABP and *ebl-1* show restriction patterns that are well conserved among different parasite isolates. This conservation of gene structure and the sequence relationships between the *ebl-1* and SABP domains suggest that *ebl-1* encodes a novel erythrocyte binding molecule having receptor properties distinct from those of SABP.

Southern hybridization experiments using probes from these open reading frames have indicated that additional copies of these conserved sequences are located elsewhere in the genome. The largest of the open reading frames on chromosome 7 is 8 kilobases and contains four tandem repeats homologous to the N-terminal, cysteine-rich unit of SABP and DABP.

Figure 1 represents an alignment of the DBL family with the DABP binding domain and two homologous regions of SABP (F_1 and F_2). The DBL family is divided into two sub-families to achieve optimal alignment. Conserved cysteine residues are shown in bold face and conserved aromatic residues are underlined.

The polypeptides of the invention can be used to raise monoclonal antibodies specific for the binding domains of SABP, DABP or the conserved regions in the *DBL* gene family. The antibodies can be used for diagnosis of malarial infection or as therapeutic agents to inhibit binding of merozoites to erythrocytes. The production of monoclonal antibodies against a desired antigen is well known to those of skill in the art and is not reviewed in detail here.

The multitude of techniques available to those skilled in the art for production and manipulation of various immunoglobulin molecules can thus be readily applied to inhibit binding. As used herein, the terms "immunoglobulin" and "antibody" refer to a protein consisting of one or more polypeptides substantially encoded by immunoglobulin genes. Immunoglobulins may exist in a variety of forms besides antibodies, including for example, Fv, Fab, and F(ab)₂, as well as in single chains. For a general review of immunoglobulin structure and function see, Fundamental Immunology, 2d Ed., W.E. Paul ed., Ravens Press, N.Y., (1989).

Antibodies which bind polypeptides of the invention may be produced by a variety of means. The production of non-human monoclonal antibodies, e.g., murine, lagomorpha, equine, etc., is well known and may be accomplished by, for example, immunizing the animal with a preparation containing the polypeptide. Antibody-producing cells obtained from the immunized animals are immortalized and screened, or screened first for the production of antibody which inhibits binding between and meroxoites and erythrocytes and then immortalized.

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For a discussion of general procedures of monoclonal antibody production see Harlow and Lane, *Antibodies, A Laboratory Manual* Cold Spring Harbor Publications, N.Y. (1988).

Thus, the present invention allows targeting of protective immune responses or monoclonal antibodies to sequences in the binding domains that are conserved between SABP, DABP and encoded regions of the DBL family. Identification of the binding regions of these proteins facilitates vaccine development because it allows for a focus of effort upon the functional elements of the large molecules. The particular sequences within the binding regions refine the target to critical regions that have been conserved during evolution, and are thus preferred for use as vaccines against the parasite.

The genes of the *DBL* family (which have not previously been sequenced) can be used as markers to detect the presence of the *P. falciparum* parasite in patients. This can be accomplished by means well known to practitioners in the art using tissue or blood from symptomatic patients in PCR reactions with oligonucleotides complementary to portions of the genes of the *DBL* family. Furthermore, sequencing the *DBL* family provides a means for skilled practitioners to generate defined probes to be used as genetic markers in a variety of applications.

Additionally, the present invention defines a conserved motif present in, but not restricted to other members of the subphylum Apicomplexa which participates in host parasite interaction. This motif can be identified in Plasmodium species and other parasitic protozoa by the polymerase chain reaction using the synthetic oligonucleotide primers shown in Figure 3. PCR methods are described in detail below. These primers are designed from regions in the conserved motif showing the highest degree of conservation among DABP, SABP and the DBL family. Figure 3 shows these regions and the consensus amino acid sequences derived from them.

A. <u>General Methods</u>

Much of the nomenclature and general laboratory procedures required in this application can be found in Sambrook, et al., Molecular Cloning A Laboratory Manual, 2nd Ed., Vol. 1-3, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1989. The manual is hereinafter referred to as "Sambrook, et al., 1989."

The practice of this invention involves the construction of recombinant nucleic acids and the expression of genes in transfected cells. Molecular cloning techniques to achieve these ends are known in the art. A wide variety of cloning and *in vitro* amplification methods suitable for the construction of recombinant nucleic acids are well-known to persons of skill. Examples of these techniques and instructions sufficient to direct persons of skill through many cloning exercises are found in Berger and Kimmel, *Guide to Molecular Cloning Techniques, Methods in Enzymology* volume 152 Academic Press, Inc., San Diego, CA (Berger); and *Current Protocols in Molecular Biology*, F.M. Ausubel *et al.*, eds., Current Protocols, a joint venture between Greene Publishing Associates, Inc. and John Wiley & Sons, Inc., (1994 Supplement) (Ausubel).

Examples of techniques sufficient to direct persons of skill through *in vitro* amplification methods, including the polymerase chain reaction (PCR) the ligase chain reaction (LCR), Q\$\beta\$-replicase amplification and other RNA polymerase mediated techniques are found in Berger, Sambrook et al., 1989, and Ausubel, as well as Mullis et al., (1987) U.S. Patent No. 4,683,202; PCR Protocols A Guide to Methods and Applications (Innis et al. eds), Academic Press Inc., San Diego, CA, 1990) ("Innis"); Arnheim & Levinson (October 1, 1990) C&EN 36-47; The

Journal Of NIH Research (1991) 3, 81-94; Kwoh et al. (1989) Proc. Natl. Acad. Sci. USA 86, 1173; Guatelli et al. (1990) Proc. Natl. Acad. Sci. USA 87, 1874; Lomell et al. (1989) J. Clin. Chem 35, 1826; Landegren et al., (1988) Science 241, 1077-1080; Van Brunt (1990) Biotechnology 8, 291-294; Wu and Wallace, (1989) Gene 4, 560; and Barringer et al. (1990) Gene 89, 117. Improved methods of cloning in vitro amplified nucleic acids are described in Wallace et al., U.S. Pat. No. 5,426,039.

The culture of cells used in the present invention, including cell lines and cultured cells from tissue or blood samples is well known in the art. Freshney (*Culture of Animal Cells, a Manual of Basic Technique, third ed.*, Wiley-Liss, New York, NY (1994)) and the references cited therein provides a general guide to the culture of cells.

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Methods of producing polyclonal and monoclonal antibodies are known to those of skill in the art. See, e.g., Coligan (1991) Current Protocols in Immunology Wiley/Greene, NY; and Harlow and Lane (1989) Antibodies: A Laboratory Manual Cold Spring Harbor Press, NY; Stites et al. (eds.) Basic and Clinical Immunology (4th ed.) Lange Medical Publications, Los Altos, CA, and references cited therein; Goding (1986) Monoclonal Antibodies: Principles and Practice (2d ed.) Academic Press, New York, NY; and Kohler and Milstein (1975) Nature 256: 495-497. Other suitable techniques for antibody preparation include selection of libraries of recombinant antibodies in phage or similar vectors. See, Huse et al. (1989) Science 246: 1275-1281; and Ward, et al. (1989) Nature 341: 544-546. Specific Monoclonal and polyclonal antibodies will usually bind with a KD of at least about .1 mM, more usually at least about 1 μ M, and most preferably at least about .1 μ M or better.

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B. Methods for isolating DNA encoding SABP, DABP and DBL binding regions

The nucleic acid compositions of this invention, whether RNA, cDNA, genomic DNA, or a hybrid of the various combinations, may be isolated from natural sources or may be synthesized in vitro. The nucleic acids claimed may be present in transformed or transfected whole cells, in a transformed or transfected cell lysate, or in a partially purified or substantially pure form.

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Techniques for nucleic acid manipulation of genes encoding the binding domains of the invention, such as subcloning nucleic acid sequences encoding polypeptides into expression vectors, labelling probes, DNA hybridization, and the like are described generally in Sambrook *et al.*, 1989.

Recombinant DNA techniques can be used to produce the binding domain polypeptides. In general, the DNA encoding the SABP and DABP binding domains are first cloned or isolated in a form suitable for ligation into an expression vector. After ligation, the vectors containing the DNA fragments or inserts are introduced into a suitable host cell for expression of the recombinant binding domains. The polypeptides are then isolated from the host cells.

There are various methods of isolating the DNA sequences encoding the SABP, DABP and DBL binding domains. Typically, the DNA is isolated from a genomic or cDNA library using labelled oligonucleotide probes specific for sequences in the DNA. Restriction endonuclease digestion of genomic DNA or cDNA containing the appropriate genes can be used to isolate the DNA encoding the binding domains of these proteins. Since the DNA

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sequences of the SABP and DABP genes are known, a panel of restriction endonucleases can be constructed to give cleavage of the DNA in the desired regions. After restriction endonuclease digestion, DNA encoding SABP binding domain or DABP binding domain is identified by its ability to hybridize with nucleic acid probes, for example on Southern blots, and these DNA regions are isolated by standard methods familiar to those of skill in the art. See Sambrook, et al., 1989.

The polymerase chain reaction can also be used to prepare DABP, SABP DBL binding domain DNA. Polymerase chain reaction technology (PCR) is used to amplify nucleic acid sequences of the DABP and SABP binding domains directly from mRNA, from cDNA, and from genomic libraries or cDNA libraries. The primers shown in Figure 3 are particularly preferred for this process.

Appropriate primers and probes for amplifying the SABP and DABP binding region DNA's are generated from analysis of the DNA sequences. In brief, oligonucleotide primers complementary to the two 3' borders of the DNA region to be amplified are synthesized. The polymerase chain reaction is then carried out using the two primers. See *PCR Protocols: A Guide to Methods and Applications*. (Innis, M, Gelfand, D., Sninsky, J. and White, T., (eds.), Academic Press, San Diego, CA (1990). Primers can be selected to amplify the entire DABP regions or to amplify smaller segments of the DABP and SABP binding domains, as desired.

Oligonucleotides for use as probes are chemically synthesized according to the solid phase phosphoramidite triester method first described by Beaucage, S.L. and Caruthers, M.H., 1981, Tetrahedron Letts., 22(20):1859-1862 using an automated synthesizer, as described in Needham-VanDevanter, D.R., et al. 1984, Nucleic Acids Res., 12:6159-6168. Purification of oligonucleotides is by either native acrylamide gel electrophoresis or by anion-exchange HPLC as described in Pearson, J.D. and Regnier, F.E., 1983, J. Chrom., 255:137-149.

The sequence of the synthetic oligonucleotides can be verified using the chemical degradation method of Maxam, A.M. and Gilbert, 1980, in W., Grossman, L. and Moldave, D., eds. Academic Press, New York, NY, *Methods in Enzymology* 65:499-560.

Other methods known to those of skill in the art may also be used to isolate DNA encoding all or part of the SABP or DABP binding domains. See Sambrook, et al., 1989.

C. Expression of DABP, SABP and DBL Binding Domain Polypeptides

Once binding domain DNAs are isolated and cloned, one may express the desired polypeptides in a recombinantly engineered cell such as bacteria, yeast, insect (especially employing baculoviral vectors), and mammalian cells. It is expected that those of skill in the art are knowledgeable in the numerous expression systems available for expression of the DNA encoding the DABP and SABP binding domains. No attempt to describe in detail the various methods known for the expression of proteins in prokaryotes or eukaryotes will be made.

In brief summary, the expression of natural or synthetic nucleic acids encoding binding domains will typically be achieved by operably linking the DNA or cDNA to a promoter (which is either constitutive or inducible), followed by incorporation into an expression vector. The vectors can be suitable for replication and integration in either prokaryotes or eukaryotes. Typical expression vectors contain transcription and translation terminators, initiation sequences, and promoters useful for regulation of the expression of the DNA encoding the

binding domains. To obtain high level expression of a cloned gene, it is desirable to construct expression plasmids which contain, at the minimum, a strong promoter to direct transcription, a ribosome binding site for translational initiation, and a transcription/translation terminator.

1. <u>Expression in Prokaryotes</u>

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Examples of regulatory regions suitable for this purpose in *E. coli* are the promoter and operator region of the *E. coli* tryptophan biosynthetic pathway as described by Yanofsky, C., 1984, J. Bacteriol., 158:1018-1024 and the leftward promoter of phage lambda (P_L) as described by Herskowitz, I. and Hagen, D., 1980, Ann. Rev. Genet., 14:399-445. The inclusion of selection markers in DNA vectors transformed in *E. coli* is also useful. Examples of such markers include genes specifying resistance to ampicillin, tetracycline, or chloramphenicol. See Sambrook *et al.*, 1989, for details concerning selection markers for use in *E. coli*.

The vector is selected to allow introduction into the appropriate host cell. Bacterial vectors are typically of plasmid or phage origin. Appropriate bacterial cells are infected with phage vector particles or transfected with naked phage vector DNA. If a plasmid vector is used, the bacterial cells are transfected with the plasmid vector DNA.

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Expression systems for expressing the DABP and SABP binding domains are available using *E. coli, Bacillus* sp. (Palva, I *et al.*, 1983, Gene 22:229-235; Mosbach, K. *et al.* Nature, 302:543-545 and *Salmonella*. *E. coli* systems are preferred.

The binding domain polypeptides produced by prokaryote cells may not necessarily fold properly. During purification from *E. coli*, the expressed polypeptides may first be denatured and then renatured. This can be accomplished by solubilizing the bacterially produced proteins in a chaotropic agent such as guanidine HCI and reducing all the cysteine residues with a reducing agent such as beta-mercaptoethanol. The polypeptides are then renatured, either by slow dialysis or by gel filtration. U.S. Patent No. 4,511,503.

Detection of the expressed antigen is achieved by methods known in the art as radioimmunoassays, Western blotting techniques or immunoprecipitation. Purification from *E. coli* can be achieved following procedures described in U.S. Patent No. 4,511,503.

2. Synthesis of SABP, DABP and DBL Binding Domains in Eukaryotes

A variety of eukaryotic expression systems such as yeast, insect cell lines and mammalian cells, are known to those of skill in the art. As explained briefly below, the DABP and SABP binding domains may also be expressed in these eukaryotic systems.

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a. <u>Expression in Yeast</u>

Synthesis of heterologous proteins in yeast is well known and described. *Methods in Yeast Genetics*, Sherman, F., et al., Cold Spring Harbor Laboratory, (1982) is a well recognized work describing the various methods available to produce the binding domains in yeast.

Examples of promoters for use in yeast include GAL1,10 (Johnson, M., and Davies, R.W., 1984, Mol. and Cell. Biol., 4:1440-1448) ADH2 (Russell, D., et al. 1983, J. Biol. Chem., 258:2674-2682), PH05 (EMBO J. 6:675-680, 1982), and MF α l (Herskowitz, I. and Oshima, Y., 1982, in The Molecular Biology of the Yeast

Saccharomyces, (eds. Strathern, J.N. Jones, E.W., and Broach, J.R., Cold Spring Harbor Lab., Cold Spring Harbor, N.Y., pp. 181-209. A multicopy plasmid with a selective marker such as Leu-2, URA-3, Trp-1, and His-3 is also desirable.

A number of yeast expression plasmids like YEp6, YEp13, YEp4 can be used as vectors. A gene of interest can be fused to any of the promoters in various yeast vectors. The above-mentioned plasmids have been fully described in the literature (Botstein, et al., 1979, Gene, 8:17-24; Broach, et al., 1979, Gene, 8:121-133).

Two procedures are used in transforming yeast cells. In one case, yeast cells are first converted into protoplasts using zymolyase, lyticase or glusulase, followed by addition of DNA and polyethylene glycol (PEG). The PEG-treated protoplasts are then regenerated in a 3% agar medium under selective conditions. Details of this procedure are given in the papers by J.D. Beggs, 1978, Nature (London), 275:104-109; and Hinnen, A., et al., 1978, Proc. Natl. Acad. Sci. USA, 75:1929-1933. The second procedure does not involve removal of the cell wall. Instead the cells are treated with lithium chloride or acetate and PEG and put on selective plates (Ito, H., et al., 1983, J. Bact., 153:163-168).

The binding domains can be isolated from yeast by lysing the cells and applying standard protein isolation techniques to the lysates. The monitoring of the purification process can be accomplished by using Western blot techniques or radioimmunoassays of other standard immunoassay techniques.

b. <u>Expression in Mammalian and Insect Cell Cultures</u>

Illustrative of cell cultures useful for the production of the binding domains are cells of insect or mammalian origin. Mammalian cell systems often will be in the form of monolayers of cells although mammalian cell suspensions may also be used. Illustrative examples of mammalian cell lines include VERO and HeLa cells, Chinese hamster ovary (CHO) cell lines, W138, BHK, Cos-7 or MDCK cell lines.

As indicated above, the vector, e. g., a plasmid, which is used to transform the host cell, preferably contains DNA sequences to initiate transcription and sequences to control the translation of the antigen gene sequence. These sequences are referred to as expression control sequences. When the host cell is of insect or mammalian origin illustrative expression control sequences are obtained from the SV-40 promoter (Science, 222:524-527, 1983), the CMV I.E. Promoter (Proc. Natl. Acad. Sci. 81:659-663, 1984) or the metallothionein promoter (Nature 296:39-42, 1982). The cloning vector containing the expression control sequences is cleaved using restriction enzymes and adjusted in size as necessary or desirable and ligated with DNA coding for the SABP or DABP polypeptides by means well known in the art.

As with yeast, when higher animal host cells are employed, polyadenlyation or transcription terminator sequences from known mammalian genes need to be incorporated into the vector. An example of a terminator sequence is the polyadenlyation sequence from the bovine growth hormone gene. Sequences for accurate splicing of the transcript may also be included. An example of a splicing sequence is the VPI intron from SV40 (Sprague, J. et al., 1983, J. Virol. 45: 773-781).

Additionally, gene sequences to control replication in the host cell may be incorporated into the vector such as those found in bovine papilloma virus type-vectors. Saveria-Campo, M., 1985, "Bovine Papilloma virus

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DNA a Eukaryotic Cloning Vector" in DNA Cloning Vol. II a Practical Approach Ed. D.M. Glover, IRL Press, Arlington, Virginia pp. 213-238.

The host cells are competent or rendered competent for transformation by various means. -There are several well-known methods of introducing DNA into animal cells. These include: calcium phosphate precipitation, fusion of the recipient cells with bacterial protoplasts containing the DNA, treatment of the recipient cells with liposomes containing the DNA, DEAE dextran, electroporation and micro-injection of the DNA directly into the cells.

The transformed cells are cultured by means well known in the art. <u>Biochemical Methods in Cell Culture and Virology</u>, Kuchler, R.J., Dowden, Hutchinson and Ross, Inc., (1977). The expressed DABP and SABP binding domain polypeptides are isolated from cells grown as suspensions or as monolayers. The latter are recovered by well known mechanical, chemical or enzymatic means.

c. Expression in recombinant vaccinia virus- or adenovirus-infected cells

In addition to use in recombinant expression systems, the isolated binding domain DNA sequences can also be used to transform viruses that transfect host cells in the patient. Live attenuated viruses, such as vaccinia or adenovirus, are convenient alternatives for vaccines because they are inexpensive to produce and are easily transported and administered. Vaccinia vectors and methods useful in immunization protocols are described, for example, in U.S. Patent No. 4,722,848.

Suitable viruses for use in the present invention include, but are not limited to, pox viruses, such as canarypox and cowpox viruses, and vaccinia viruses, alpha viruses, adenoviruses, and other animal viruses. The recombinant viruses can be produced by methods well known in the art, for example, using homologous recombination or ligating two plasmids. A recombinant canarypox or cowpox virus can be made, for example, by inserting the DNA's encoding the DABP and SABP binding domain-polypeptides into plasmids so that they are flanked by viral sequences on both sides. The DNA's encoding the binding domains are then inserted into the virus genome through homologous recombination.

A recombinant adenovirus can be produced, for example, by ligating together two plasmids each containing about 50% of the viral sequence and the DNA sequence encoding erythrocyte binding domain polypeptide. Recombinant RNA viruses such as the alpha virus can be made via a cDNA intermediate using methods known in the art.

In the case of vaccinia virus (for example, strain WR), the DNA sequence encoding the binding domains can be inserted in the genome by a number of methods including homologous recombination using a transfer vector, pTKgpt-OFIS as described in Kaslow, et al., Science 252:1310-1313 (1991).

Alternately the DNA encoding the SABP and DABP binding domains may be inserted into another plasmid designed for producing recombinant vaccinia, such as pGS62, Langford, C.L., et al., 1986, Mol. Cell. Biol. 6:3191-3199. This plasmid consists of a cloning site for insertion of foreign genes, the P7.5 promoter of vaccinia to direct synthesis of the inserted gene, and the vaccinia TK gene flanking both ends of the foreign gene.

Confirmation of production of recombinant virus can be achieved by DNA hybridization using cDNA encoding the DABP and SABP binding domain polypeptides and by immunodetection techniques using antibodies

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specific for the expressed binding domain polypeptides. Virus stocks may be prepared by infection of cells such as HELA S3 spinner cells and harvesting of virus progeny.

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The recombinant virus of the present invention can be used to induce anti-SABP and anti-DABP binding domain antibodies in mammals, such as mice or humans. In addition, the recombinant virus can be used to produce the SABP and DABP binding domains by infecting host cells in vitro, which in turn express the polypeptide (see section on expression of SABP and DABP binding domains in eukaryotic cells, above).

The present invention also relates to host cells infected with the recombinant virus. The host cells of the present invention are preferably mammalian, such as BSC-1 cells. Host cells infected with the recombinant virus express the DABP and SABP binding domains on their cell surfaces. In addition, membrane extracts of the infected cells induce protective antibodies when used to inoculate or boost previously inoculated mammals.

D. <u>Purification of the SABP, DABP and DBL Binding Domain Polypeptides</u>

The binding domain polypeptides produced by recombinant DNA technology may be purified by standard techniques well known to those of skill in the art. Recombinantly produced binding domain polypeptides can be directly expressed or expressed as a fusion protein. The protein is then purified by a combination of cell lysis (e. g., sonication) and affinity chromatography. For fusion products, subsequent digestion of the fusion protein with an appropriate proteolytic enzyme release the desired SABP and DABP binding domains.

The polypeptides of this invention may be purified to substantial purity by standard techniques well known in the art, including selective precipitation with such substances as ammonium sulfate, column chromatography, immunopurification methods, and others. See, for instance, R. Scopes, *Protein Purification: Principles and Practice,* Springer-Verlag, New York, NY (1982).

E. <u>Production of Binding Domains by protein chemistry techniques</u>

The polypeptides of the invention can be synthetically prepared in a wide variety of ways. For instance polypeptides of relatively short size, can be synthesized in solution or on a solid support in accordance with conventional techniques. Various automatic synthesizers are commercially available and can be used in accordance with known protocols. See, for example, Stewart and Young, *Solid Phase Peptide Synthesis*, 2d. ed., Pierce Chemical Co. (1984).

Alternatively, purified and isolated SABP, DABP or DBL family proteins may be treated with proteolytic enzymes in order to produce the binding domain polypeptides. For example, recombinant DABP and SABP proteins may be used for this purpose. The DABP and SABP protein sequence may then be analyzed to select proteolytic enzymes to be used to generate polypeptides containing desired regions of the DABP and SABP binding domain. The desired polypeptides are then purified by using standard techniques for protein and peptide purification. For a review of standard techniques see, *Methods in Enzymology*, "Guide to Protein Purification", M. Deutscher, ed. Vol. 182 (1990), pages 619-626.

F. <u>Modification of nucleic acid and polypeptide sequences</u>

The nucleotide sequences used to transfect the host cells used for production of recombinant binding domain polypeptides can be modified according to standard techniques to yield binding domain polypeptides,

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with a variety of desired properties. The binding domain polypeptides of the present invention can be readily designed and manufactured utilizing various recombinant DNA techniques well known to those skilled in the art. For example, the binding domain polypeptides can vary from the naturally-occurring sequence at the primary structure level by amino acid insertions, substitutions, deletions, and the like. These modifications can be used in a number of combinations to produce the final modified protein chain.

The amino acid sequence variants can be prepared with various objectives in mind, including facilitating purification and preparation of the recombinant polypeptides. The modified polypeptides are also useful for modifying plasma half-life, improving therapeutic efficacy, and lessening the severity or occurrence of side effects during therapeutic use. The amino acid sequence variants are usually predetermined variants not found in nature but exhibit the same immunogenic activity as naturally occurring polypeptides. For instance, polypeptide fragments comprising only a portion (usually at least about 60-80%, typically 90-95%) of the primary structure may be produced. For use as vaccines, polypeptide fragments are typically preferred so long as at least one epitope capable of eliciting production of blocking antibodies remains.

In general, modifications of the sequences encoding the binding domain polypeptides may be readily accomplished by a variety of well-known techniques, such as site-directed mutagenesis (see, Giliman and Smith, *Gene* 8:81-97 (1979) and Roberts, S. *et al.*, *Nature* 328:731-734 (1987)). One of ordinary skill will appreciate that the effect of many mutations is difficult to predict. Thus, most modifications are evaluated by routine screening in a suitable assay for the desired characteristic. For instance, changes in the immunological character of the polypeptide can be detected by an appropriate competitive binding assay. Modifications of other properties such as redox or thermal stability, hydrophobicity, susceptibility to proteolysis, or the tendency to aggregate are all assayed according to standard techniques.

G. <u>Diagnostic and Screening Assays</u>

The polypeptides and nucelic acids of the invention can be used in diagnostic applications for the detection of merozoites or nucleic acids in a biological sample. The presence of parasites can be detected using several well recognized specific binding assays based on immunological results. (See U.S. Patents 4,366,241; 4,376,110; 4,517,288; and 4,837,168). For instance, labeled monoclonal antibodies to polypeptides of the invention can be used to detect merozoites in a biological sample. Alternatively, labelled polypeptides of the invention can be used to detect the presence of antibodies to SABP or DABP in a biological sample. For a review of the general procedures in diagnostic immunoassays, see also *Basic and Clinical Immunology* 7th Edition (D. Stites and A. Terr ed.) 1991.

In addition, modified polypeptides, antibodies or other compounds capable of inhibiting the interaction between SABP or DABP and erythrocytes can be assayed for biological activity. For instance, polypeptides can be recombinantly expressed on the surface of cells and the ability of the cells to bind erythrocytes can be measured as described below. Alternatively, peptides or antibodies can tested for the ability to inhibit binding between erythrocytes and merozoites or SABP and DABP.

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Cell-free assays can also be used to measure binding of DABP or SABP polypeptides to isolated Duffy antigen or glycophorin polypeptides. For instance, the erythrocyte proteins can be immobilized on a solid surface and binding of labelled SABP or DABP polypeptides can be measured.

Many assay formats employ labelled assay components. The labelling systems can be in a variety of forms. The label may be coupled directly or indirectly to the desired component of the assay according to methods well known in the art. A wide variety of labels may be used. The component may be labelled by any one of several methods. The most common method of detection is the use of autoradiography with 3H , ^{125}I , ^{35}S , ^{14}C , or ^{32}P labelled compounds or the like. Non-radioactive labels include ligands which bind to labelled antibodies, fluorophores, chemiluminescent agents, enzymes, and antibodies which can serve as specific binding pair members for a labelled ligand. The choice of label depends on sensitivity required, ease of conjugation with the compound, stability requirements, and available instrumentation.

In addition, the polypeptides of the invention can be assayed using animal models, well known to those of skill in the art. For P falciparum the in vivo models include Actus sp. monkeys or chimpanzees; for P. vivax the in vivo models include Saimiri monkeys.

In the case of the use nucleic acids for diagnostic purposes, standard nucleic hybridization techniques can be used to detect the presence of the genes identified here (e.g., members of the DBL family). If desired, nucleic acids in the sample may first be amplified using standard procedures such as PCR. Diagnostic kits comprising the appropriate primers and probes can also be prepared.

H. **DBL** Targeted Therepeutics

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DBL polypeptides are expressed on the surface of Plasmodium-infected erythrocytes. As such, they present ideal targets for therepeutics which target infected erythrocytes. In one preferred embodiement of the present invention, cytotoxic antibodies or antibody fusion proteins with cytotoxic agents are targeted against DBL proteins, killing infected erythrocytes and inhibiting the reproduciton of *Plasmodium* in an infected host.

The procedure for attaching a cytotoxic agent to an antibody will vary according to the chemical structure of the agent. Antibodies and cytotoxic agents are typically bound together chemically or, where the antibody and cytotoxic agents are both polypeptides, are optionally synthesized recombinantly as a fusion protein. Polypeptides typically contain variety of functional groups; e.g., carboxylic acid (COOH) or free amine (-NH₂) groups, which are available for reaction with a suitable functional group on either the antibody or the cytotoxic agent.

Alternatively, antibodies or cytotoxic agents are derivitized to attach additional reactive functional groups. The derivatization optionally involves attachment of linker molecules such as those available from Pierce Chemical Company, Rockford Illinois. A "linker", as used herein, is a molecule that is used to join the nucleic acid binding molecule to the receptor ligand. The linker is capable of forming covalent bonds to both the antibody and the cytotoxic agent. Suitable linkers are well known to those of skill in the art and include, but are not limited to, straight or branched-chain carbon linkers, heterocyclic carbon linkers, or peptide linkers. Where the antibody and the cytotoxic agent are polypeptides, the linkers are joined to the constituent amino acids through their side groups (e.g., through a disulfide linkage to cysteine) or to the alpha carbon amino and carboxyl groups of the terminal amino acids.

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A bifunctional linker having one functional group reactive with a group on a particular ligand, and another group reactive with a nucleic acid binding molecule, can be used to form the desired conjugate. Alternatively, derivatization can proceed through chemical treatment of the ligand or nucleic acid binding molecule, e.g., glycol cleavage of the sugar moiety of a glycoprotein with periodate to generate free aldehyde groups. The free aldehyde groups on the glycoprotein may be reacted with free amine or hydrazine groups on an agent to bind the agent thereto (See, e.g., U.S. Patent No. 4,671,958). Procedures for generation of free sulfhydryl groups on polypeptides, are known (See, e.g., U.S. Pat. No. 4,659,839).

Many procedures and linker molecules for attachment of various compounds to proteins are known. See, for example, European Patent Application No. 188,256; U.S. Patent Nos. 4,671,958, 4,659,839, 4,414,148, 4,699,784; 4,680,338; 4,569,789; and 4,589,071; and Borlinghaus *et al. Cancer Res.* 47: 4071-4075 (1987). In particular, production of various antibody conjugates is well-known within the art and can be found, for example in Thorpe *et al., Monoclonal Antibodies in Clinical Medicine,* Academic Press, pp. 168-190 (1982), Waldmann, *Science*, 252: 1657 (1991), and U.S. Patent Nos. 4,545,985 and 4,894,443.

A number of antibodies which bind cell surface receptors have been converted to form suitable for incorporation into fusion proteins, and similar strategies are used to create fusion-protein antibodies which bind DBR polypeptides. see Batra et al., Mol. Cell. Biol., 11: 2200-2205 (1991); Batra et al., Proc. Natl. Acad. Sci. USA, 89: 5867-5871 (1992); Brinkmann, et al. Proc. Natl. Acad. Sci. USA, 88: 8616-8620 (1991); Brinkmann et al., Proc. Natl. Acad. Sci. USA, 87: 1066-1070 (1990); Friedman et al., Cancer Res. 53: 334-339 (1993); Kreitman et al., J. Immunol., 149: 2810-2815 (1992); Nicholls et al., J. Biol. Chem., 268: 5302-5308 (1993); and Wells, et al., Cancer Res., 52: 6310-6317 (1992), respectively).

B. Production of Fusion Proteins

Where the antibody fragment and/or the cytotoxic agents are relatively short polypeptides (i.e., less than about 50 amino acids) they are often synthesized using standard chemical peptide synthesis techniques. Where both molecules are relatively short, a chimeric molecule is optionally synthesized as a single contiguous polypeptide. Alternatively, the ligand and the nucleic acid binding molecule can be synthesized separately and then fused chemically.

Solid phase synthesis in which the C-terminal amino acid of the sequence is attached to an insoluble support followed by sequential addition of the remaining amino acids in the sequence is a preferred method for the chemical synthesis of the ligands of this invention. Techniques for solid phase synthesis are described by Barany and Merrifield, Solid-Phase Peptide Synthesis; pp. 3-284 in The Peptides: Analysis, Synthesis, Biology. Vol. 2: Special Methods in Peptide Synthesis, Part A., Merrifield, et al., J. Am. Chem. Soc., 95: 2149-2156 (1963), and Stewart et al., Solid Phase Peptide Synthesis, 2nd ed. Pierce Chem. Co., Rockford, Ill. (1984).

In a preferred embodiment, the fusion molecules of the invention are synthesized using recombinant nucleic acid methodology. Generally this involves creating a nucleic acid sequence that encodes the receptor-targeted fusion molecule, placing the nucleic acid in an expression cassette under the control of a particular promoter, expressing the protein in a host, isolating the expressed protein and, if required, renaturing the protein. Techniques

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sufficient to guide one of skill through such procedures are found in, e.g., Berger, Sambrook, Ausubel, Innis, and Freshney (all supra).

While the two molecules are often joined directly together, one of skill will appreciate that the molecules may be separated by a peptide spacer consisting of one or more amino acids. Generally the spacer will have no specific biological activity other than to join the proteins or to preserve some minimum distance or other spatial relationship between them. However, the constituent amino acids of the spacer may be selected to influence some property of the molecule such as the folding, net charge, or hydrophobicity.

Once expressed, recombinant fusion proteins can be purified according to standard procedures, including ammonium sulfate precipitation, affinity columns, column chromatography, gel electrophoresis and the like (see, generally, R. Scopes, *Protein Purification*, Springer-Verlag, N.Y. (1982), Deutscher, *Methods in Enzymology Vol.* 182: Guide to Protein Purification., Academic Press, Inc. N.Y. (1990)). Substantially pure compositions of about 50 to 95% homogeneity are preferred, and 80 to 95% or greater homogeneity are most preferred for use as therepeutic agents.

One of skill in the art will recognize that after chemical synthesis, biological expression, or purification, the fusion molecule may possess a conformation substantially different than the native conformations of the constituent polypeptides. In this case, it is often necessary to denature and reduce the polypeptide and then to cause the polypeptide to re-fold into the preferred conformation. Methods of reducing and denaturing proteins and inducing re-folding are well known to those of skill in the art (See, Debinski *et al. J. Biol. Chem.*, 268: 14065-14070 (1993); Kreitman and Pastan, *Bioconjug. Chem.*, 4: 581-585 (1993); and Buchner, *et al., Anal. Biochem.*, 205: 263-270 (1992).

I. Pharmaceutical compositions comprising binding domain polypeptides

The polypeptides of the invention are useful in therapeutic and prophylactic applications for the treatment of malaria. Pharmaceutical compositions of the invention are suitable for use in a variety of drug delivery systems. Suitable formulations for use in the present invention are found in *Remington's Pharmaceutical Sciences*, Mack Publishing Company, Philadelphia, PA, 17th ed. (1985). For a brief review of methods for drug delivery, see, Langer, *Science* 249:1 527-1533 (1990).

The polypeptides of the present invention can be used in pharmaceutical and vaccine compositions that are useful for administration to mammals, particularly humans. The polypeptides can be administered together in certain circumstances, e.g. where infection by both P. falciparum and P. vivax is likely. Thus, a single pharmaceutical composition can be used for the treatment or prophylaxis of malaria caused by both parasites.

The compositions are suitable for single administrations or a series of administrations. When given as a series, inoculations subsequent to the initial administration are given to boost the immune response and are typically referred to as booster inoculations.

The pharmaceutical compositions of the invention are intended for parenteral, topical, oral or local administration. Preferably, the pharmaceutical compositions are administered parenterally, e.g., intravenously, subcutaneously, intradermally, or intramuscularly. Thus, the invention provides compositions for parenteral

administration that comprise a solution of the agents described above dissolved or suspended in an acceptable carrier, preferably an aqueous carrier. A variety of aqueous carriers may be used, e.g., water, buffered water, 0.4% saline, 0.3% glycine, hyaluronic acid and the like. These compositions may be sterilized by conventional, well known sterilization techniques, or may be sterile filtered. The resulting aqueous solutions may be packaged for use as is, or lyophilized, the lyophilized preparation being combined with a sterile solution prior to administration. The compositions may contain pharmaceutically acceptable auxiliary substances as required to approximate physiological conditions, such as pH adjusting and buffering agents, tonicity adjusting agents, wetting agents and the like, for example, sodium acetate, sodium lactate, sodium chloride, potassium chloride, calcium chloride, sorbitan monolaurate, triethanolamine oleate, etc.

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For solid compositions, conventional nontoxic solid carriers may be used which include, for example, pharmaceutical grades of mannitol, lactose, starch, magnesium stearate, sodium saccharin, talcum, cellulose, glucose, sucrose, magnesium carbonate, and the like. For oral administration, a pharmaceutically acceptable nontoxic composition is formed by incorporating any of the normally employed excipients, such as those carriers previously listed, and generally 10-95% of active ingredient and more preferably at a concentration of 25%-75%.

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For aerosol administration, the polypeptides are preferably supplied in finely divided form along with a surfactant and propellant. The surfactant must, of course, be nontoxic, and preferably soluble in the propellant. Representative of such agents are the esters or partial esters of fatty acids containing from 6 to 22 carbon atoms, such as caproic, octanoic, lauric, palmitic, stearic, linoleic, linolenic, olesteric and oleic acids with an aliphatic polyhydric alcohol or its cyclic anhydride. Mixed esters, such as mixed or natural glycerides may be employed. A carrier can also be included, as desired, as with, e.g., lecithin for intranasal delivery.

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In certain embodiments patients with malaria may be treated with SABP or DABP polypeptides or other specific blocking agents (e.g. monoclonal antibodies) that prevent binding of *Plasmodium* merozoites and schizonts to the erythrocyte surface.

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The amount administered to the patient will vary depending upon what is being administered, the state of the patient and the manner of administration. In therapeutic applications, compositions are administered to a patient already suffering from malaria in an amount sufficient to inhibit spread of the parasite through erythrocytes and thus cure or at least partially arrest the symptoms of the disease and its complications. An amount adequate to accomplish this is defined as "therapeutically effective dose." Amounts effective for this use will depend on the severity of the disease, the particular composition, and the weight and general state of the patient. Generally, the dose will be in the range of about 1mg to about 5gm per day, preferably about 100 mg per day, for a 70 kg patient.

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Alternatively, the polypeptides of the invention can be used prophylactically as vaccines. The vaccines of the invention contain as an active ingredient an immunogenically effective amount of the binding domain polypeptide or of a recombinant virus as described herein. The immune response may include the generation of antibodies; activation of cytotoxic T lymphocytes (CTL) against cells presenting peptides derived from the peptides encoded by the SABP, DABP or DBL sequences of the present invention, or other mechanisms well known in the art.

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See e.g. Paul Fundamental Immunology, Second Edition (Raven Press, New York, NY) for a description of immune response. Useful carriers are well known in the art, and include, for example, thyroglobulin, albumins such as human serum albumin, tetanus toxoid, polyamino acids such as poly(D-lysine:D-glutamic acid), influenza, hepatitis B virus core protein, hepatitis B virus recombinant vaccine. The vaccines can also contain a physiologically tolerable (acceptable) diluent such as water, phosphate buffered saline, or saline, and further typically include an adjuvant. Adjuvants such as incomplete Freund's adjuvant, aluminum phosphate, aluminum hydroxide, or alum are materials well known in the art.

The DNA or RNA encoding the SABP or DABP binding domains and the DBL gene family motifs may be introduced into patients to obtain an immune response to the polypeptides which the nucleic acid encodes. Wolff et. al., *Science* 247: 1465-1468 (1990) which is describes the use of nucleic acids to produce expression of the genes which the nucleic acids encode.

Vaccine compositions containing the polypeptides, nucleic acids or viruses of the invention are administered to a patient to elicit a protective immune response against the polypeptide. A "protective immune response" is one which prevents or inhibits the spread of the parasite through erythrocytes and thus at least partially prevent the symptoms of the disease and its complications. An amount sufficient to accomplish this is defined as an "immunogenically effective dose." Amounts effective for this use will depend on the composition, the manner of administration, the weight and general state of health of the patient, and the judgment of the prescribing physician. For peptide compositions, the general range for the initial immunization (that is for therapeutic or prophylactic administration) is from about 100 μ g to about 1 gm of peptide for a 70 kg patient, followed by boosting dosages of from about 100 μ g to about 1 gm of the polypeptide pursuant to a boosting regimen over weeks to months depending upon the patient's response and condition e.g. by measuring levels of parasite in the patient's blood. For nucleic acids, typically 30-1000ug of nucleic acid is injected into a 70kg patient, more typically about 50-150ug of nucleic acid is injected into a 70kg patient followed by boosting doses as appropriate.

The following examples illustrate preferred embodiments of the invention.

EXAMPLE 1: <u>Identification of the amino-terminal, cysteine-rich region of SABP and DABP as binding</u> <u>domains for erythrocytes</u>

1. Expression of the SABP binding domain polypeptide on the surface of Cos cells.

To demonstrate that the amino-terminal, cysteine-rich region of the SABP protein is the sialic acid binding region, this region of the protein was expressed on the surface of mammalian Cos cells *in vitro*. This DNA sequence is from position 1 to position 1848 of the SABP DNA sequence (SEQ ID No 3). Polymerase chain reaction technology (PCR) was used to amplify this region of the SABP DNA directly from the cloned gene.

Sequences corresponding to restriction endonuclease sites for Pvull or Apal were incorporated into the oligonucleotide sequence of the probes used in PCR amplification in order to facilitate insertion of the PCR-amplified regions into the pRE4 vector (see below). The specific oligonucleotides, 5'-ATCGATCAGCTGGGAAGAAATACTTCATCT-3'(SEQID NO:17) and 5'-ATCGATGGGCCCCGAAGTTTGTTCATTATT-3'

(SEQ ID NO:18) were synthesized. These oligonucleotides were used as primers to PCR-amplify the region of the DNA sequence encoding the cysteine-rich amino terminal region of the SABP protein.

PCR conditions were based on the standard described in Saiki, et al., Science 239: 487-491 (1988). Template DNA was provided from cloned fragments of the gene encoding SABP which had been spliced and re-cloned as a single open-reading frame piece.

The vector, pRE4, used for expression in Cos cells is shown in Figure 2. The vector has an SV40 origin of replication, an ampicillin resistance marker and the Herpes simplex virus glycoprotein D gene (HSV glyd) cloned downstream of the Rous sarcoma virus long terminal repeats (RSV LTR). Part of the extracellular domain of the HSV glyd gene was excised using the Pvull and Apal sites in HSV glyd.

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As described above, the PCR oligonucleotide primers contained the Pvull or Apal restriction sites. The PCR-amplified DNA fragments obtained above were digested with the restriction enzymes Pvull and Apal and cloned into the Pvull and Apal sites of the vector pRE4. These constructs were designed to express regions of the SABP protein as chimeric proteins with the signal sequence of HSV glyd at the N-terminal end and the transmembrane and cytoplasmic domain of HSV glyd at the C-terminal end. The signal sequence of HSV glyd targets these chimeric proteins to the surface of Cos cells and the transmembrane segment of HSV glyd anchors these chimeric proteins to the Cos cell surface.

Mammalian Cos cells were transfected with the pRE4 constructs containing the PCR-amplified SABP DNA regions, by calcium phosphate precipitation according to standard techniques.

2. Expression of the DABP binding domain polypeptide on the surface of Cos cells.

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To demonstrate that the amino-terminal, cysteine-rich region of the DABP protein is the binding domain, this region was expressed on the surface of Cos cells. This region of the DNA sequence from position 1-975 was first PCR-amplified (SEQ ID No 1).

Sequences corresponding to restriction endonuclease sites for Pvull or Apal were incorporated into the oligonucleotide probes used for PCR amplification in order to facilitate subsequent insertion of the amplified DNA into the pRE4 vector, as described above. The oligonucleotides, 5'-TCTCGTCAGCTGACGATCTCTAGTGCTATT-3' (SEQ ID NO:19) and 5'-ACGAGTGGGCCCTGTCACAACTTCCTGAGT-3' (SEQ ID NO:20) were synthesized. These oligonucleotides were used as primers to amplify the region of the DABP DNA sequence encoding the cysteine-rich, amino-terminal region of the DABP protein directly from the cloned DABP gene, using the same conditions described above.

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The same pRE4 vector described above in the section on expression of SABP regions in Cos cells was also used as a vector for the DABP DNA regions.

3. Binding studies with erythrocytes.

To demonstrate their ability to bind human erythrocytes, the transfected Cos cells expressing binding domains from DABP and SABP were incubated with erythrocytes for two hours at 37°C in culture media (DMEM/10% FBS). The non-adherent erythrocytes were removed with five washes of phosphate-buffered saline and the bound erythrocytes were observed by light microscopy. Cos cells expressing the amino terminal, cysteine-rich

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SABP polypeptides on their surface bound untreated human erythrocytes, but did not bind neuraminidase treated erythrocytes, that is, erythrocytes which lack sialic acid residues on their surface. Cos cells expressing other regions of the SABP protein on their surface did not bind human erythrocytes. These results identified the amino-terminal, cysteine-rich region of SABP as the erythrocyte binding domain and-indicated that the binding of Cos cells expressing these regions to human erythrocytes is specific. Furthermore, the binding of the expressed region to erythrocytes is identical to the binding pattern seen for the authentic SABP- 175 molecule upon binding to erythrocytes.

Similarly, Cos cells expressing the amino-terminal cysteine-rich region of DABP on their surface bound Duffy-positive human erythrocytes, but did not bind Duffy-negative human erythrocytes, that is erythrocytes which lack the Duffy blood group antigen. Cos cells expressing other regions of the DABP protein on their surface did not bind human erythrocytes. These results identified the amino-terminal cysteine rich region of DABP as the erythrocyte binding domain and indicated that the binding of the Cos cells was specific.

EXAMPLE 2: Isolation of polynucleotide sequences in the DBL family

P.falciparum clones and cell line used include the following. P. falciparum clones 3D7, D10, LF4/1, Camp/A1, SL/D6, HB3, 7G8, V1/S, T2/C6, KMWII, ItG2F6, FCR3/A2 and Dd2 have been previously tabulated (Dolan, et al. (1993), Mol. Biochem. Parasitol. 61, 137-142). Line Dd2/NM1 was selected from clone Dd2 for invasion via a sialic acid-independent pathway (Dolan, et al. (1990), J. Clin. Invest. 86, 618-624). All parasites were maintained in vitro by standard methods (Trager, et al. (1976), Science 193, 673-675).

DNA and RNA Isolation and Analysis. DNA was extracted as described (Peterson, et al. (1990), Proc. Natl. Acad. Sci. USA 87, 3018-3022). Endonuclease digestion, agarose gel electrophoresis, and filter hybridizations were performed by standard methods (Sambrook, et al., 1989). All hybridizations were at 56°C (Sambrook, et al., 1989). Blots were washed for 2 min. at room temperature in 2x standard saline/phosphate/EDTA (SSPE) with 0.5% SDS, followed by two higher stringency washes at 50°C in 0.3xSSPE with 0.5% SDS. Parasite chromosomes were embedded in agarose blocks and separated by pulsed field gel electrophoresis (Dolan, et al. (1993), Methods. Mol. Biol. 21, 319-332). RNA was isolated from cultured parasites by LiC1 extraction of Catrimox-14-precipitated RNA (Dahle, et al. (1993), BioTechniques 15, 1102-1105). Agarose gel electrophoresis of total RNA and filter hybridizations were performed by standard methods (Sambrook, et al., (1989).

Oligonucleotide Primers and PCR. Primers specific for E31a used in a RT-PCR to test for expression of this sequence were E31aT2 (5'-AGA-CCT-CAA-TTT-CTA-AG-3') (SEQ ID NO:21) and E31aRev1 (5'-AAT-CGC-GAG-CAT-CAT-CTG-3') (SEQ ID NO:22).

Two primers were used to amplify additional sequences from genes encoding *DBL* domains. These were designed from conserved amino acids encoded in the *DBL* domain of the eba-175 and E31a sequences. After adaptation to incorporate the most frequently-used *P. falciparum* codons, forward primer UNIEBP5' [5'-CC(A/G)-AG(G/A)-AG(G/A)-CAA-(G/A)AA-(C/T)TA-TG-3'] (SEQ ID NO:23), based upon the amino acid sequence PRRQKLC, and reverse primer UNIEBP3' [5'-CCA-(A/T)C(T/G)-(T/G)A(A/G)-(A/G)AA-TTG-(A/T)GG-3'] (SEQ ID NO:24), based upon the amino acid sequence PQFLRW, were synthesized.

RT-PCR amplifications were performed as described (Kawasaki, et al. (1990), PCR Protocols, A Guide to Methods and Applications, eds. Innis, M.A., Gelfand, D.H., Sninsku, J.J. & White, T.J. (Academic, San Diego), pp. 21-27). In brief, 0.5 to 1 mg of total RNA was treated with RQ1 DNAse (Promega), phenol/chloroform extracted, and ethanol precipitated. The RNA was then annealed with random oligonucleotide primers and extended with Superscript reverse transcriptase (GIBCO/BRL). PCR cycling conditions were 94°C for 10 sec, 45°C for 15 sec, and 72°C for 45 sec, for 30 cycles. All PCRs were performed in an Idaho Technology air thermal cycler using buffer containing 2 mM Mg2+.

PCR amplification products were separated by use of PCR Purity Plus gels and protocols (AT Biochem, Malvern, PA).

DNA Clones and Hybridization Probes. Clone pE31a was isolated from a genomic library prepared from the region of chromosome 7 linked to chloroquine resistance Walker-Jonah, et al. (1992), Mol. Biochem. Parasitol. 51, 313-320. Clone pS31H (GenBank accession no. L38454), containing an insert encompassing that of pE31a, was cloned from a size-selected Hind III restriction digest of Dd2 genomic DNA.

Clone pEBLe1 was cloned from a RT-PCR of Dd2 cDNA after amplification with primers UNIEBP5' (SEQ ID NO:23) and UNIEBP3' (SEQ ID NO:24). Clone pEBP1.2 (GenBank accession no. L38450), containing an insert encompassing that of pEBLe1, was isolated from a Dd2 cDNA library probed with pEBLe1. *DBL*-encoding sequences of *dbl-nm1-4* (GenBank accession no. L38455) and *dbl-nm1-5* (GenBank accession no. L38453) were amplified by RT-PCR from first strand cDNA of line Dd2/NM using primers UNIEBP5' and UNIEBP3'. Sequencing was performed on double stranded DNA templates by standard protocols for the dideoxynucleotide method. (Sequenase; U.S. Biochemicals).

Sequences related to the E31a sequence were detected with the 3005 bp insert of clone pS31H. The eba-175 gene was detected with a PCR amplified probe consisting of the first 1825 bp of the coding sequence. ebl-1 sequences were detected with the 2098 bp insert of clone pEBP1.2. All probes were comparable in organization, each containing a region encoding at least one DBL domain and varying amounts of flanking sequence.

Homology searches and alignments. Homology searches were performed with BLAST and the Genetics Computer Group program FASTA (Altschul, et al. (1990), J. Mol. Biol. 215, 403-410; Devereux, et al. (1984), Nucleic Acids. Res. 12(1 Pt 1, 387-395). Optimized alignments were produced with MACAW sequence alignment software (Schuler, et al. (1991), Proteins. 9, 180-190).

Multiple P. falciparum sequences encode DBL domains. Positional cloning experiments directed to P. falciparum chromosome 7 identified an ORF (E31a) encoding a DBL domain that is homologous to the domains found in the P. vivax and P. knowlesi DABPs and the P. falciparum SABP. Figure 4 shows the realtive position of the E31a ORF on chromosome 7.

The homology between the *DBL* domains of E31a and the erythrocyte-binding proteins is due to the presence of short motifs of highly conserved amino acids. These well-conserved stretches are separated by non-homologous sequences and by deletions and insertions that vary the size of the domain by greater than 60 aa. The typical *DBL* domain contains 12 or more cysteine residues and has 7 conserved tryptophan residues. Additional

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well conserved amino acids include 4 arginines, 3 aspartates, 9 positions with aliphatic residues (alanine, isoleucine, leucine, or valine) and 4 with aromatic amino acids (tryptophan, phenylalanine, or tyrosine).

Probes spanning the sequence that encodes the E31a *DBL* domain hybridized to multiple fragments within a single restriction digest and yielded bands that varied among parasite lines. The numerous distinct bands from a selection of different parasite DNAs indicated a large number of diverse but related elements. These multiple bands varied among different *P. falciparum* clones, in contrast to the well-conserved, single-copy signal obtained with the *eba-175* probe.

Because of the numerous cross-hybridizing sequences, it seemed likely that many of these related sequences would be on different chromosomes of the parasite. PFG electrophoresis of *P. falciparum* Dd2 chromosomes and hybridization with the E31a probe identified a number of cross-hybridizing sequences on multiple chromosomes. A control hybridization with the *eba-175* probe under identical conditions yielded a single band of hybridization from chromosome 7.

RNA Analysis of *DBL* Elements. Sequences from E31a (pS31H insert) were used to probe RNA blots for corresponding transcripts. No hybridization was detected. Because it was still possible that a message of low abundance was not being detected on the RNA blot, RT-PCR was used as a means of more sensitive detection. For this purpose, cDNA was generated by RT from random primers annealed to DNAse-treated total RNA. E31a-specific oligonucleotides were then used to test for amplification from the cDNA. No amplification of the E31a sequence was obtained, while genomic DNA controls and amplification from cDNA by dihydrofolate reductase/thymidylate synthetase-specific primers yielded the expected bands. A screen of a cDNA library with E31a specific probes also failed to detect any clones hybridizing with the ORF. These results indicate that E31a is either a pseudogene, or is expressed in parasite strains or stages not examined in this work.

A PCR Method to Isolate Sequences Encoding DBL Domains. The identification of short conserved motifs in DBL domains that otherwise have extreme diversity led to a PCR strategy using degenerate oligonucleotide primers designed from conserved amino acid sequences in the DBL domains. Sequences PRROKLC and PQFLRW were judged most suitable for minimizing degeneracy while allowing amplification of expressed DBL sequences. After these considerations and adjustment for P. falciparum codon usage, primers UNIEBP5' and UNIEBP3' were synthesized.

While some *P. falciparum* lines yielded similar patterns of amplified bands *(e. g.* Dd2 and MCamp; FCR3/A2 and K-1), no two separate isolates showed identical patterns, reflecting the diversity of the *DBL* domains in the parasite lines. A few bands of the same apparent size were present in many isolates. These included a consistent 490 bp product that was determined to be the *eba-175* gene by its expected size and hybridization to a gene-specific probe. The number of discernible bands probably underestimates the number of amplifiable sequences because of overlapping products of the same size and possible preferential amplification of some sequences over others. Nevertheless, the parasite-specific patterns in the amplified bands may provide a means to quickly type isolates and serves as a measure of parasite diversity in field samples.

To identify *DBL*-encoding sequences in RNA transcripts, the UNIEBP primers were used to amplify first-strand cDNAs generated from DNAse-treated RNA preparations. Amplified products from Dd2, 3D7, HB3 and MCAMP cDNAs had diverse sizes ranging from 400 bp to nearly 1 kb. These included a band at 480-500 bp that was determined to be *eba-175* from its expected size and cross-hybridization to an *eba-175*-specific probe. Other bands were from amplification of different transcripts encoding *DBL* domains. Dd2-NM1 RNA, for example, yielded bands above the *eba-175* product that included two related sequences (*dbl-nm1-4,dbl-nm1-5*). These bands were found to be isolate-specific and to have features consistent with the *var* genes described in Example 3, below. Probes that detect *dbl-nm1-4* and *dbl-nm1-5* hybridized to multiple chromosomes and aligned more closely with E31a than with EBA-175 or DABP.

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The RT-PCR amplifications also yielded a consistent band that encoded a novel *DBL* domain distinct from *eba-175*. A cDNA clone corresponding to this product was isolated by screening a *Agt10 Dd2 cDNA* library with a radiolabeled *ebl-1* probe. Sequence from this and additional overlapping cDNA clones confirmed the conserved motifs of the *DBL* domain. The alignment of the predicted amino acid sequences showed that the *DBL* domain of *ebl-1* is more similar to *eba-175* than to the multicopy genes. There was, however, extensive divergence from *eba-175* and other known genes outside of the amplified region.

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In contrast to the multicopy hybridization patterns of dbl-nm1-4 and dbl-nm1-5, the ebl-1 sequence, like that of eba-175, was found to have hybridization patterns consistent with a conserved single-copy gene. Probes specific for ebl-1 hybridized only to chromosome 13, and restriction analysis with the enzymes Cla I, EcoRI, HindIII, Hinf I, Nsi I, Rsa I, and Spe I, all yielded bands expected from a single copy sequence. RNA blots probed with ebl-1-specific sequences showed several bands of hybridization, however, corresponding to 8-9.5 kb transcripts in mRNA from the Dd2 and 3D7 parasites. The transcripts of different size may result from alternative start and termination points or from incompletely processed species containing introns.

EXAMPLE 3: Isolation of var genes

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Parasite clones, DNA analysis and Chromosome Mapping. Parasite clones were cultivated by the methods of (Trager, et al. (1976), Science 193, 673-675). DNA was extracted from parasite cultures as described (Peterson, et al. (1988), Proc. Natl. Acad. Sci. USA 85, 9114-9118) except that the DNA was as recoverd by ethanol precipitation rather than spooling. Fingerprint analysis with the pC4.H32 probe was used to confirm DNA preparations (Dolan, et al. (1993), Mol. Biochem. Parasitol. 61, 137-142). Southern blotting to Nytran membranes was recommended by the manufacturer (Schleicher & Schuell, Keene, NH). PFG separation of the 14 P. falciparum chromosomes and chromosome mapping were performed as described (Wellems, et al. (1987), Cell 49, 633-642; Sinnis, et al. (1986); Genomics 3, 237-295).

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RNA isolation. Parasites from 200 ml mixed stage cultures (5-10% parasitemia) were released by saponin lysis as for DNA preparations except that the procedures were performed with ice-cold solutions. RNA was immediately isolated from the parasite pellet by guanidine thiocyanate/phenol-chloroform methods, recovered and treated with RNAase-free DNAse (Creedon, et al. (1994), J. Biol. Chem. 269, 16364-16370. RNA in H₂O was combined with 2 vol 100% ETOH, distributed into 2 ml vials and frozen as stock at -70°C. RNA was recovered by

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precipitation with 0.1 vol 3M NaOAc. RNA blots were generated and probed as described (Creedon, et al. (1994), J. Biol. Chem. 269, 16364-16370).

YAC isolation, chromosome-segment libraries and cDNA libraries. Overlapping YACs spanning the 300 kb segment of chromosome 7 that contains the CQR locus were obtained from a YAC library of a CQR FCR3 parasite line de Bruin, et al. (1992), Genomics 14, 332-339) by the procedures of Lanzer, et al. (1993), Nature 361, 654-657. Orientation of the YACs and their overlaps were identified with probes obtained from the YAC ends by inverted PCR.

Attempts to construct cosmid libraries and large insert (~ 10 kb) λ libraries from high molecular weight P. falciparum genomic DNA yielded only rearranged clones. An alternative approach was therefore taken in which chromosome-segment libraries were constructed that contained small (0.5-5 kb) inserts in plasmid vectors. Plasmid libraries containing Alul, Hinfl, Rsal and Sspl inserts in pCDNAII were constructed from Dd2 chromosome 7 restriction fragments purified by pulsed-field gel (PFG) electrophoresis (Wellems, $et\ al.\ (1991)$, $Proc.\ Natl.\ Acad.\ Sci.\ USA\ 88$, 3382-3386). A plasmid library from a 34 kb $Apal\ Smal\$ restriction fragment of YAC PfYED9 was constructed by the same methods. Inserts in the plasmid libraries were generally 0.5-4 kb.

The λ gt10 Dd2 cDNA library was prepared under contract by CloneTech Laboratories Inc. (Palo Alto, CA) from the DNAse-treated, polyA + fraction of Dd2 RNA. The cDNA was generated in two separate reactions using oligodT primers or random primers. Products of these reactions were combined, processed and cloned into the EcoRI site of λ gt10. 1.6 x 10⁶ independent recombinants were obtained and amplified.

Isolation of overlapping clones and DNA sequencing. Plasmid clones from the chromosome-segment and YAC-segment libraries were picked at random and their locations were established by restriction mapping. After sequence data from these clones were generated, overlapping clones were isolated in a process of "chromosome walking" by rescreening the libraries with oligonucleotide probes near the ends of sequenced inserts. Sufficient divergence was present among repetitive elements in the sequences to allow distinction of clones and unambiguous assignment of overlaps (generally 50-200 bp).

Sequencing reactions with single-strand M13 DNA (1 μ g) and double-strand plasmid DNA (2-5 μ g) were performed in 96-well polyvinyl chloride U-bottom microassay plates using a Sequenase protocol recommended by United States Biochemical Corp. (Cleveland, OH). Reactions were separated by 8M urea-6% polyacrylamide sequencing gels and exposed to Kodak BioMax MR film. Sequence data from some clones were also obtained by use of an ABI 373A automated DNA sequencer (Applied Biosystems Inc., Foster City, CA). Cycle sequencing reactions were performed using the ABI PRISM DyeDeoxy system.

DNA sequence editing, analyses and display were performed with MacVector software (International Biotechnologies Inc., New Haven, CT), BLAST (Altschul, et al. (1990), J. Mol. Biol. 215, 403-410), Genetics Computer Group programs (Devereux, et al. (1984), Nucleic Acids Res. 12, 387-395) and the DNADRAW package (Shapiro, et al. (1986), Nucleic Acids Res. 14, 65-73) maintained at the National Institutes of Health.

Identification of a large hypervariable region within a chromosome 7 segment linked to chloroquine resistance. Four overlapping yeast artificial chromosomes from the *P. falciparum* FCR3 line were obtained that span the 300 kb chromosome segment linked to CQR, a segment located 300-600 kb from the telomere of chromosome

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7. Figure 5 shows the positions of these YACs (PfYEF2, PfYFE6, PfYKF8, PfYED9) relative to the chromosome map. In order to define the structure of this 300 kb segment, we performed comparative hybridizations to search for polymorphisms between parasite lines. Clones were randomly picked from chromosome segment-specific plasmid libraries and their inserts were hybridized against restriction digests of the YAC and parasite DNAS. Over thirty inserts were identified that recognized PfYEF2, PfYFE6 or PfYKF8 and showed a predonderance of single copy sequences with few polymorphisms (Alul, Hinfl, Rsal and Sspl digests), consistent with prior findings that chromosome internal regions are largely conserved and contain a preponderance of single copy sequences. However, fifteen other inserts that recognized PfYED9 showed highly polymorphic sets of repetitive elements in the parasite DNAs. Southern analysis indicated that these polymorphic elements were part of a chromosome hypervariable region contained within the PfYED9 clone.

Mapping and DNA sequencing of the hypervariable region spanned by YAC PfYED9. Single copy sequences detected by pE45b and pH270.5 flank the hypervariable region spanned by PfYED9 (Figure 5). The pE45b and pH270.5 probes were therefore used to assign large restriction fragments on the PfYED9 map and establish enzyme recognition sites as reference points. A detailed restriction map of the PfYED9 hypervariable region was then developed. Fifteen overlapping clones ("a"-"f' and "h"-"o" in Figure 5) were isolated by a chromosome walking approach from Dd2 chromosome subsegment libraries (Wellems *et al.*, *supra*) The inserts yielded 19.1 kb of continuous Dd2 sequence having predicted enzyme recognition sites in perfect accord with the PfYED9 restriction map. Such agreement indicates that the Dd2 and FCR3 sequences in this part of the chromosome are very similar, despite differences elsewhere in the genome that are evident by restriction analysis.

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We also obtained genomic sequence data from the 34 kb Apal-Smal fragment of PfYED9. Purified PfYED9 DNA was cut with Smal to yield a 110 kb fragment, which was then isolated by PFG electrophoresis and digested with Apal. The resulting 34 kb Apal-Smal band was purified by PFG electrophoresis, digested in four separate reactions by Alul, Hinfl, Rsal or Sspl and incorporated into a plasmid (PCDNAII) library. Cloned inserts from the library were checked for hybridization to the PfYED9 34 kb fragment, assigned to the PfYED9 map and sequenced (Figure 5). Overlapping inserts were obtained by the chromosome walking approach except for three gaps ("t", "z", " θ " in Figure 5) which were closed by PCR amplification of PfYED9 DNA using primers from flanking sequences. The clones from PfYED9 ("r"·"z","y", " κ " and " α "+" β " in Figure 5) yielded 22.2 kb of continuous DNA sequence that overlaps the Dd2 sequence at the "f"l" β " junction and has predicted restriction sites that match the PfYED9 map perfectly. The composite sequence from the Dd2 and PfYED9 segments is 40,171 kb.

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Structure of a var gene cluster and comparative analysis of predicted amino acid sequences. The 40,171 bp sequence contains three 10-12 kb regions that have related sequences and structure. Each of these regions harbors a pair of ORFS. The first ORF in each pair begins with a consensus ATG start codon preceded by typical *P. falciparum* non-coding sequence of abundant A+T content. The ORFs of each pair are separated by an intervening AT-rich and non-coding sequence of 0.9 kb to 1.1 kb. Presence of consensus intron-exon splice junction sequences at either end of these intervening sequences and lack of a consistent translation start site in the 3' ORF indicate that the each pair of ORFs belongs to an individual gene having a two exon structure. This has been verified by

comparison of the genomic sequences to the cDNA sequence of an expressed gene (*var-7*; see subsequent section). The three 10 kb to 12 kb regions thus contain members of a variant gene family which have coding regions of 9.23kb (*var-1*), 7.99 kb (*var-2*) and 9.01 kb (*var-3*). Predicted molecular weights of the encoded proteins are 350 kD, 302 kD and 344 kD. respectively.

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The var genes are flanked by additional members of the var family in PfYED9. Restriction analysis identified two additional genes that are 12-35 kb upstream of the sequenced region and are closely related to var-2 and var-3 (var-2c and Var-3c, Figure 5). The var genes thus have a clustered arrangement in which many individual members are organized in head-to-tail fashion. Between var-1 and var-2 is a 5 kb DNA sequence that harbors a short ORF homologous to that of a repetitive element (rij) suggested to be a transposable element in P. falciparum.

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The deduced protein sequences of the var genes are highly diverse, yet all contain certain conserved motifs and common structural features. Database searches identified 2 to 4 domains within each var sequence that are homologous to cysteine-rich domains of SABP and DABP. In the var sequences, the first domain near the amino-terminus (DBL domain 1) is the most conserved of the DBL domains and has amino acid signatures that differentiate it from subsequent domains (e.g. consensus peptide sequences GAcAp[Y/F]rrL, CTxLARsfadlgdlVrgrdLYLG and VPTYFDYVpqylrwF). Between DBL domains 1 and 2 is another type of conserved domain, a cysteine-rich interdomain region (CIDR) of 300-400 amino acids. The CIDR does not have all the motifs of a DBL domain, but it does have a region at the 3'end which is homologous to the end of the FI DBL domain in SABP. The conservation evident in the sequences of DBL domain I and the CIDR suggest that these regions maintain important structures in the head of the variant molecule.

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DBL domains 2, 3 and 4 (numbering is according to *var- 1*, the first sequence completed) have less discriminating signatures than domain 1, and show features of cross-alignment and variation in number that suggest these domains can undergo shuffling and deletion.

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DBL domain 4 is followed by a segment of variable length and a hydrophobic region that is encoded at the end of the first exon (exon 1). In all var sequences this hydrophobic region fits the criteria of a transmembrane segment. The second exon (exon II) encodes a large (45-55 kD) conserved C-terminal sequence that has an acid character (predicted pl = 4.5, vs. 5.9 for the part of the protein upstream of the splice junction) and a cysteine content of < 1% (vs. > 4% upstream). The position of this C-terminal sequence downstream of a single transmembrane segment suggests that it has a cytoplasmic location.

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No consensus signal sequence was detected in the NH₂-terminal region of the predicted var ORFs. We note the presence of several motifs in the protein sequences that are known to act as ligands and receptors in the integrin family. These include RGD (var-1 codons 886-88, 1992-94) and DGEA (var-1 codons 2111-14). Not all of these motifs occur in each protein sequence and, when they do occur, their positions vary.

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Identification of var transcripts and chromosome expression sites. To identify transcribed var sequences we screened a Agt10 Dd2 cDNA library with var-containing BssHII restriction fragments that had been purified from PfYED9 and radiolabeled by random hexamer priming. This screening yielded 18 clones with inserts that hybridized back to PfYED9. By cross-hybridization studies and DNA sequence analysis the inserts fell into two groups: group

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I inserts that aligned with sequences of var exon I (AT240, AT242, AT244, AT284, AT287, AT288, AT295, AT296); and group II inserts that aligned with sequences of var exon II (AT140, AT141, AT142, AT145, AT147, AT148, AT150, AT152).

The full ORF of an expressed var gene (var-7) was determined from AT242 and overlapping cDNA clones that were obtained by a PCR-based walking strategy. The sequence showed that var-7 has a 6.6 kb ORF containing two DBL domains, a hydrophobic transmembrane sequence and carboxy-terminal region typical of var genes (predicted molecular weight 249 kD). Comparison of var-7 with the var-1 sequence demonstrated continuity of the alignments at the predicted splice junction between the ORFs of exons 1 and II. PCR amplification of Dd2 genomic DNA was also performed with primers derived from the two var-7 exons. Sequence of this var-7 PCR product confirmed consensus splice sites and a 1 kb intron typical of the var genes. Transcription of var-7 was detected as a 7.5 kb band by RNA blot analysis.

Chromosome mapping experiments with a var-7-specific probe localized the var-7 gene to a region that is 600 kb from one end of Dd2 chromosome 12 (chromosome 12 has a length of 2600 kb). No hybridization of the var-7 probe was detected to any other Dd2 chromosome nor to any chromosomes of the HB3, 3D7 or A4 parasites. Other cDNA inserts from the group I clones were also sequenced and examined for chromosome hybridization signals. The \(\lambda\tau240\) cDNA insert mapped to the \(\var-1\lnotar-2\lnotar-3\) cluster on Dd2 chromosome 7 and its sequence matched that of \(\var-3\). The \(\lambda\tau244\), \(\lambda\tau284\), \(\lambda\tau287\), \(\lambda\tau288\), \(\lambda\tau295\) and \(\lambda\tau296\) inserts all showed overlapping sequences and yielded the same hybridization patterns. Chromosome sites recognized by these inserts included regions within two \(Smal\) fragments from Dd2 chromosome 7 and another from chromosome 9. We note that loss of a cytoadherence phenotype has been correlated with a chromosome 9 deletion in certain \(P\). falciparum lines.

1.8 kb to 2.4 kb RNA transcripts related to var exon II. In addition to the 7.5 kb var-7 band, a broad 1.8 kb to 2.4 kb band was detected on RNA blots after hybridization with a probe that recognizes var exon II. Sequences of eight group II cDNA inserts homologous to exon II were therefore determined and aligned against the var genes. Comparative analysis of the insert sequences showed that all differed from one another in regions of overlap, indicating that transcription of the corresponding RNAs was from different loci. Three of the cDNA sequences (AT140, AT141 and AT148) aligned downstream of the intron/exon II splice junction. However, five other cDNA inserts (AT142, AT145, AT147, AT150 and AT152) had sequences that aligned upstream of the var intron/exon II splice site and included regions homologous to var intron sequences. In the vicinity of the splice junction, consensus splice sites occurred in three of the cDNA sequences (AT142, AT147, AT150) while a fourth sequence (AT145) showed the required AG dinucleotide but not the expected pyrimidine tract of the splice consensus. The part of the fifth sequence (AT152) that aligned with the var intron extended upstream only to the TAG of the splice sequence. All five sequences lacked a consensus start codon preceded by A+T-rich non-coding DNA that is typical of P. falciparum translation start sites.

<u>Isolate-specific var sequences and evidence for DNA recombination in cultivated parasite clones.</u> The diversity of var forms expressed by *P. falciparum* parasites reflects a tremendous repertoire in the var gene family.

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This repertoire is evident in the patterns of restriction polymorphism detected by var probes as well as in the detection of var-specific sequences that hybridize to some parasite DNAs but not to others. The var-7 gene expressed by Dd2, for example, is not present in the HB3, 3D7 or A4 genomes. Such var diversity suggests that frequent DNA rearrangements underlie the production of antigenically variant types in different parasite strains.

To test for DNA rearrangements in parasites cultivated *in vitro*, we used *var* sequences to probe restricted DNAs from Dd2 lines adapted to neuraminidase-treated erythrocytes. In one rearrangement a novel 35 kb *BgN* fragment is seen in NM1 DNA probed with the *X*T142 (group II) insert. In another rearrangement a deletion of a 20 kb *Pst*1 band is evident in NM8 DNA probed with a *var*-7 sequence. Deletion of this 20 kb band was also detected in the Dd2/R8 subclone obtained before neuraminidase selection, indicating that the DNA rearrangement was not produced by selection in neuraminidase-treated erythrocytes.

The above examples are provided to illustrate the invention and other variants of the invention encompassed by the claims will be readily apparent to one of ordinary skill in the art.

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	(1) GENERAL INFORMATION:				
5	(i) Secretar	APPLICANT: The United States, As Represented by the y, Department of Health and Human Services			
10		TITLE OF INVENTION: BINDING DOMAINS FROM PLASMODIUM VIVAX AND PLASMODIUM FALCIPARUM ERYTHROCYTE BINDING PROTEINS NUMBER OF SEQUENCES: 45			
	(111)	NOMBER OF SEQUENCES: 45			
15	(iv)	CORRESPONDENCE ADDRESS: (A) ADDRESSEE: Knobbe Martens Olson & Bear (B) STREET: 620 Newport Center Drive 16th Floor (C) CITY: Newport Beach (D) STATE: California (E) COUNTRY: US (F) ZIP: 92660			
20	· (v)	COMPUTER READABLE FORM:			
25	(,,	(A) MEDIUM TYPE: Floppy disk (B) COMPUTER: IBM PC compatible (C) OPERATING SYSTEM: PC-DOS/MS-DOS (D) SOFTWARE: PatentIn Release #1.0, Version #1.25			
30	(vi)	CURRENT APPLICATION DATA: (A) APPLICATION NUMBER: (B) FILING DATE: (C) CLASSIFICATION:			
35	(vii)	PRIOR APPLICATION DATA (A) APPLICATION NUMBER: US08/487826 (B) FILING DATE: 07-JUN-1996			
	·(viii)	ATTORNEY/AGENT INFORMATION: (A) NAME: Israelsen, Ned (B) REGISTRATION NUMBER: 29,655 (C) REFERENCE/DOCKET NUMBER: NIH121.001QPC			
40		· ·			
٠	(ix)	TELECOMMUNICATION INFORMATION: (A) TELEPHONE: (619) 235-8550 (B) TELEFAX: (619) 235-0176			
45	(2) INFOR	FORMATION FOR SEQ ID NO:1:			
50	(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 4084 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear			
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	(vi)	ORIGINAL SOURCE: (A) ORGANISM: Plasmodium vivax			
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- 5 (2) INFORMATION FOR SEQ ID NO:2:
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 - (A) LENGTH: 1115 amino acids
 - (B) TYPE: amino acid
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 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
- 15 (iii) HYPOTHETICAL: NO
- 20 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

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Leu Ile Tyr Asp Ala Ala Val Glu Gly Asp Leu Leu Lys Leu Asn

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GACCATGAAG GATCATTTCA TTGAAGCCTC TAAAAAAGAA TCTCAACTTT TGCTTAAAAA 720 AAATGATAAC AAATATAATT CTAAATTTTG TAATGATTTG AAGAATAGTT TTTTAGATTA 780



TGGACATCTT GCTATGGGAA ATGATATGGA TTTTGGAGGT TATTCAACTA AGGCAGAAAA 840 CAAAATTCAA GAAGTTTTTA AAGGGGCTCA TGGGGAAATA AGTGAACATA AAATTAAAAA 900 TTTTAGAAAA GAATGGTGGA ATGAATTTAG AGAGAAACTT TGGGAAGCTA TGTTATCTGA 960 GCATAAAAAT AATATAAATA ATTGTAAAAA TATTCCCCAA GAAGAATTAC AAATTACTCA 1020 ATGGATAAAA GAATGGCATG GAGAATTTTT GCTTGAAAGA GATAATAGAT CAAAATTGCC 1080 5 AAAAAGTAAA TGTAAAAATA ATACATTATA TGAAGCATGT GAGAAGGAAT GTATTGATCC 1140 ATGTATGAAA TATAGAGATT GGATTATTAG AAGTAAATTT GAATGGCATA CGTTATCGAA 1200 AGAATATGAA ACTCAAAAAG TTCCAAAGGA AAATGCGGAA AATTATTTAA TCAAAATTTC 1260 AGAAAACAAG AATGATGCTA AAGTAAGTTT ATTATTGAAT AATTGTGATG CTGAATATTC 1320 AAAATATTGT GATTGTAAAC ATACTACTAC TCTCGTTAAA AGCGTTTTAA ATGGTAACGA 1380 10 CAATACAATT AAGGAAAAGC GTGAACATAT TGATTTAGAT GATTTTCTA AATTTGGATG 1440 TGATAAAAAT TCCGTTGATA CAAACACAAA GGTGTGGGAA TGTAAAAACC CTTATATATT 1500 ATCCACTAAA GATGTATGTG TACCTCCGAG GAGGCAAGAA TTATGTCTTG GAAACATTGA 1560 TAGAATATAC GATAAAAACC TATTAATGAT AAAAGAGCAT ATTCTTGCTA TTGCAATATA 1620 15 TGAATCAAGA ATATTGAAAC GAAAATATAA GAATAAAGAT GATAAAGAAG TTTGTAAAAT 1680 CATAAATAAA ACTTTCGCTG ATATAAGAGA TATTATAGGA GGTACTGATT ATTGGAATGA 1740 TTTGAGCAAT AGAAAATTAG TAGGAAAAAT TAACACAAAT TCAAAATATG TTCACAGGAA 1800 TAAAAAAAAT GATAAGCTTT TTCGTGATGA GTGGTGGAAA GTTATTAAAA AAGATGTATG 1860 GAATGTGATA TCATGGGTAT TCAAGGATAA AACTGTTTGT AAAGAAGATG ATATTGAAAA 1920 TATACCACAA TTCTTCAGAT GGTTTAGTGA ATGGGGTGAT GATTATTGCC AGGATAAAAC 1980 20 AAAAATGATA GAGACTCTGA AGGTTGAATG CAAAGAAAAA CCTTGTGAAG ATGACAATTG 2040 TAAAAGTAAA TGTAATTCAT ATAAAGAATG GATATCAAAA AAAAAAGAAG AGTATAATAA 2100 ACAAGCCAAA CAATACCAAG AATATCAAAA AGGAAATAAT TACAAAATGT ATTCTGAATT 2160 TAAATCTATA AAACCAGAAG TTTATTTAAA GAAATACTCG GAAAAATGTT CTAACCTAAA 2220 25 TTTCGAAGAT GAATTTAAGG AAGAATTACA TTCAGATTAT AAAAATAAAT GTACGATGTG 2280 TCCAGAAGTA AAGGATGTAC CAATTTCTAT AATAAGAAAT AATGAACAAA CTTCGCAAGA 2340 AGCAGTTCCT GAGGAAAACA CTGAAATAGC ACACAGAACG GAAACTCCAT CTATCTCTGA 2400 AGGACCAAAA GGAAATGAAC AAAAAGAACG TGATGACGAT AGTTTGAGTA AAATAAGTGT 2460 ATCACCAGAA AATTCAAGAC CTGAAACTGA TGCTAAAGAT ACTTCTAACT TGTTAAAATT 2520 AAAAGGAGAT GTTGATATTA GTATGCCTAA AGCAGTTATT GGGAGCAGTC CTAATGATAA 2580 30 TATAAATGTT ACTGAACAAG GGGATAATAT TTCCGGGGTG AATTCTAAAC CTTTATCTGA 2640 TGATGTACGT CCAGATAAAA AGGAATTAGA AGATCAAAAT AGTGATGAAT CGGAAGAAAC 2700 TGTAGTAAAT CATATATCAA AAAGTCCATC TATAAATAAT GGAGATGATT CAGGCAGTGG 2760 AAGTGCAACA GTGAGTGAAT CTAGTAGTTC AAATACTGGA TTGTCTATTG ATGATGATAG 2820 AAATGGTGAT ACATTTGTTC GAACACAAGA TACAGCAAAT ACTGAAGATG TTATTAGAAA 2880 35 AGAAAATGCT GACAAGGATG AAGATGAAAA AGGCGCAGAT GAAGAAAGAC ATAGTACTTC 2940 TGAAAGCTTA AGTTCACCTG AAGAAAAAAT GTTAACTGAT AATGAAGGAG GAAATAGTTT 3000 AAATCATGAA GAGGTGAAAG AACATACTAG TAATTCTGAT AATGTTCAAC AGTCTGGAGG 3060 AATTGTTAAT ATGAATGTTG AGAAAGAACT AAAAGATACT TTAGAAAAATC CTTCTAGTAG 3120 40 CTTGGATGAA GGAAAAGCAC ATGAAGAATT ATCAGAACCA AATCTAAGCA GTGACCAAGA 3180 TATGTCTAAT ACACCTGGAC CTTTGGATAA CACCAGTGAA GAAACTACAG AAAGAATTAG 3240 TAATAATGAA TATAAAGTTA ACGAGAGGGA AGATGAGAGA ACGCTTACTA AGGAATATGA 3300 AGATATTGTT TTGAAAAGTC ATATGAATAG AGAATCAGAC GATGGTGAAT TATATGACGA 3360 AAATTCAGAC TTATCTACTG TAAATGATGA ATCAGAAGAC GCTGAAGCAA AAATGAAAGG 3420 45 AAATGATACA TCTGAAATGT CGCATAATAG TAGTCAACAT ATTGAGAGTG ATCAACAGAA 3480 AAACGATATG AAAACTGTTG GTGATTTGGG AACCACACAT GTACAAAACG AAATTAGTGT 3540 TCCTGTTACA GGAGAAATTG ATGAAAAATT AAGGGAAAGT AAAGAATCAA AAATTCATAA 3600 GGCTGAAGAG GAAAGATTAA GTCATACAGA TATACATAAA ATTAATCCTG AAGATAGAAA 3660 TAGTAATACA TTACATTTAA AAGATATAAG AAATGAGGAA AACGAAAGAC ACTTAACTAA 3720 TCAAAACATT AATATTAGTC AAGAAAGGGA TTTGCAAAAA CATGGATTCC ATACCATGAA 3780 50 TAATCTACAT GGAGATGGAG TTTCCGAAAG AAGTCAAATT AATCATAGTC ATCATGGAAA 3840 CAGACAAGAT CGGGGGGGAA ATTCTGGGAA TGTTTTAAAT ATGAGATCTA ATAATAATAA 3900 TTTTAATAAT ATTCCAAGTA GATATAATTT ATATGATAAA AAATTAGATT TAGATCTTTA 3960 TGAAAACAGA AATGATAGTA CAACAAAAGA ATTAATAAAG AAATTAGCAG AAATAAATAA 4020 55 ATGTGAGAAC GAAATTTCTG TAAAATATTG TGACCATATG ATTCATGAAG AAATCCCATT 4080 AAAAACATGC ACTAAAGAAA AAACAAGAAA TCTGTGTTGT GCAGTATCAG ATTACTGTAT 4140 GAGCTATTTT ACATATGATT CAGAGGAATA TTATAATTGT ACGAAAAGGG AATTTGATGA 4200 TCCATCTTAT ACATGTTTCA GAAAGGAGGC TTTTTCAAGT ATGATATTCA AATTTTTAAT 4260 AACAAATAAA ATATATTATT ATTTTTATAC TTACAAAACT GCAAAAGTAA CAATAAAAA 4320 AATTAATTTC TCATTAATTT TTTTTTTTTTT TAGGTATGCC ATATTATGCA 4380 60 GGAGCAGGTG TGTTATTTAT TATATTGGTT ATTTTAGGTG CTTCACAAGC CAAATATCAA 4440 AGGTTAGAAA AAATAAATAA AAATAAAATT GAGAAGAATG TAAATTAAAT ATAGAATTCG 4500 AGCTCGG

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(2) INFORMATION FOR SEQ ID NO:4:
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(i) SEQUENCE CHARACTERISTICS:
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- (A) LENGTH: 1435 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
 - (vi) ORIGINAL SOURCE:

(A) ORGANISM: Plasmodium falciparum

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

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Met Lys Cys Asn Ile Ser Ile Tyr Phe Phe Ala Ser Phe Phe Val Leu
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      Tyr Phe Ala Lys Ala Arg Asn Glu Tyr Asp Ile Lys Glu Asn Glu Lys
      Phe Leu Asp Val Tyr Lys Glu Lys Phe Asn Glu Leu Asp Lys Lys
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                                                       45
      Tyr Gly Asn Val Gln Lys Thr Asp Lys Lys Ile Phe Thr Phe Ile Glu
25
                              55
      Asn Lys Leu Asp Ile Leu Asn Asn Ser Lys Phe Asn Lys Arg Trp Lys
                                              75
      Ser Tyr Gly Thr Pro Asp Asn Ile Asp Lys Asn Met Ser Leu Ile Asn
                                          90
      Lys His Asn Asn Glu Glu Met Phe Asn Asn Asn Tyr Gln Ser Phe Leu
30
                  100
                                      105
                                                          110
      Ser Thr Ser Ser Leu Ile Lys Gln Asn Lys Tyr Val Pro Ile Asn Ala
              115
                                  120
                                                      125
      Val Arg Val Ser Arg Ile Leu Ser Phe Leu Asp Ser Arg Ile Asn Asn
35
                              135
     Gly Arg Asn Thr Ser Ser Asn Asn Glu Val Leu Ser Asn Cys Arg Glu
                          150
                                              155
     Lys Arg Lys Gly Met Lys Trp Asp Cys Lys Lys Asn Asp Arg Ser
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                                          170
                                                              175
      Asn Tyr Val Cys Ile Pro Asp Arg Ile Gln Leu Cys Ile Val Asn
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                                      185
                                                          190
      Leu Ser Ile Ile Lys Thr Tyr Thr Lys Glu Thr Met Lys Asp His Phe
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                                                       205
      Ile Glu Ala Ser Lys Lys Glu Ser Gln Leu Leu Lys Lys Asn Asp
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                              215
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      Asn Lys Tyr Asn Ser Lys Phe Cys Asn Asp Leu Lys Asn Ser Phe Leu
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                                              235
     Asp Tyr Gly His Leu Ala Met Gly Asn Asp Met Asp Phe Gly Gly Tyr
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                                          250
                                                              255
     Ser Thr Lys Ala Glu Asn Lys Ile Gln Glu Val Phe Lys Gly Ala His
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                  260
                                      265
                                                          270
      Gly Glu Ile Ser Glu His Lys Ile Lys Asn Phe Arg Lys Glu Trp Trp
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      Asn Glu Phe Arg Glu Lys Leu Trp Glu Ala Met Leu Ser Glu His Lys
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                              295
                                                  300
      Asn Asn Ile Asn Asn Cys Lys Asn Ile Pro Gln Glu Glu Leu Gln Ile
                                              315
     Thr Gln Trp Ile Lys Glu Trp His Gly Glu Phe Leu Leu Glu Arg Asp
                      325
                                          330
60
     Asn Arg Ser Lys Leu Pro Lys Ser Lys Cys Lys Asn Asn Thr Leu Tyr
                  340
                                      345
     Glu Ala Cys Glu Lys Glu Cys Ile Asp Pro Cys Met Lys Tyr Arg Asp
                                  360
      Trp Ile Ile Arg Ser Lys Phe Glu Trp His Thr Leu Ser Lys Glu Tyr
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Glu Thr Gln Lys Val Pro Lys Glu Asn Ala Glu Asn Tyr Leu Ile Lys Ile Ser Glu Asn Lys Asn Asp Ala Lys Val Ser Leu Leu Leu Asn Asn Cys Asp Ala Glu Tyr Ser Lys Tyr Cys Asp Cys Lys His Thr Thr Leu Val Lys Ser Val Leu Asn Gly Asn Asp Asn Thr Ile Lys Glu Lys Arg Glu His Ile Asp Leu Asp Asp Phe Ser Lys Phe Gly Cys Asp Lys Asn Ser Val Asp Thr Asn Thr Lys Val Trp Glu Cys Lys Asn Pro Tyr Ile Leu Ser Thr Lys Asp Val Cys Val Pro Pro Arg Arg Gln Glu Leu Cys Leu Gly Asn Ile Asp Arg Ile Tyr Asp Lys Asn Leu Leu Met Ile Lys Glu His Ile Leu Ala Ile Ala Ile Tyr Glu Ser Arg Ile Leu Lys Arg Lys Tyr Lys Asn Lys Asp Asp Lys Glu Val Cys Lys Ile Ile Asn Lys Thr Phe Ala Asp Ile Arg Asp Ile Ile Gly Gly Thr Asp Tyr Trp Asn Asp Leu Ser Asn Arg Lys Leu Val Gly Lys Ile Asn Thr Asn Ser Lys Tyr Val His Arg Asn Lys Lys Asn Asp Lys Leu Phe Arg Asp Glu Trp Trp Lys Val Ile Lys Lys Asp Val Trp Asn Val Ile Ser Trp Val Phe Lys Asp Lys Thr Val Cys Lys Glu Asp Asp Ile Glu Asn Ile Pro Gln Phe Phe Arg Trp Phe Ser Glu Trp Gly Asp Asp Tyr Cys Gln Asp Lys Thr Lys Met Ile Glu Thr Leu Lys Val Glu Cys Lys Glu Lys Pro Cys Glu Asp Asp Asn Cys Lys Ser Lys Cys Asn Ser Tyr Lys Glu Trp Ile Ser Lys Lys Glu Glu Tyr Asn Lys Gln Ala Lys Gln Tyr Gln Glu Tyr Gln Lys Gly Asn Asn Tyr Lys Met Tyr Ser Glu Phe Lys Ser Ile Lys Pro Glu Val Tyr Leu Lys Lys Tyr Ser Glu Lys Cys Ser Asn Leu Asn Phe Glu Asp Glu Phe Lys Glu Glu Leu His Ser Asp Tyr Lys Asn Lys Cys Thr Met Cys Pro Glu Val Lys Asp Val Pro Ile Ser Ile Ile Arg Asn Asn Glu Gln Thr Ser Gln Glu Ala Val Pro Glu Glu Asn Thr Glu Ile Ala His Arg Thr Glu Thr Pro Ser Ile Ser Glu Gly Pro Lys Gly Asn Glu Gln Lys Glu Arg Asp Asp Asp Ser Leu Ser Lys Ile Ser Val Ser Pro Glu Asn Ser Arg Pro Glu Thr Asp Ala Lys Asp Thr Ser Asn Leu Leu Lys Leu Lys Gly Asp Val Asp Ile Ser Met Pro Lys Ala Val Ile Gly Ser Ser Pro Asn Asp Asn Ile Asn Val Thr Glu Gln Gly Asp Asn Ile Ser Gly Val Asn Ser Lys Pro Leu Ser Asp Asp Val Arg Pro Asp Lys Lys Glu Leu Glu Asp Gln Asn Ser Asp Glu Ser Glu Glu Thr Val Val Asn His Ile Ser Lys Ser Pro Ser Ile Asn Asn Gly

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885
                                         890
      Asp Asp Ser Gly Ser Gly Ser Ala Thr Val Ser Glu Ser Ser Ser
                  -900
                                     905
                                                          910
      Asn Thr Gly Leu Ser Ile Asp Asp Asp Arg Asn Gly Asp Thr Phe Val
5
                                 920
      Arg Thr Gln Asp Thr Ala Asn Thr Glu Asp Val Ile Arg Lys Glu Asn
                              935
                                                  940
      Ala Asp Lys Asp Glu Asp Glu Lys Gly Ala Asp Glu Glu Arg His Ser
                          950
                                              955
      Thr Ser Glu Ser Leu Ser Ser Pro Glu Glu Lys Met Leu Thr Asp Asn
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                      965
                                          970
                                                             975
      Glu Gly Gly Asn Ser Leu Asn His Glu Glu Val Lys Glu His Thr Ser
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      Asn Ser Asp Asn Val Gln Gln Ser Gly Gly Ile Val Asn Met Asn Val
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                                1000
                                                     1005
      Glu Lys Glu Leu Lys Asp Thr Leu Glu Asn Pro Ser Ser Leu Asp
                            1015
                                                 1020
      Glu Gly Lys Ala His Glu Glu Leu Ser Glu Pro Asn Leu Ser Ser Asp
                          1030
                                             1035
      Gln Asp Met Ser Asn Thr Pro Gly Pro Leu Asp Asn Thr Ser Glu Glu
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                    1045
                                        1050
      Thr Thr Glu Arg Ile Ser Asn Asn Glu Tyr Lys Val Asn Glu Arg Glu
                1060
                                    1065
      Asp Glu Arg Thr Leu Thr Lys Glu Tyr Glu Asp Ile Val Leu Lys Ser
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            1075
                                1080
                                                    1085
      His Met Asn Arg Glu Ser Asp Asp Gly Glu Leu Tyr Asp Glu Asn Ser
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                                                1100
      Asp Leu Ser Thr Val Asn Asp Glu Ser Glu Asp Ala Glu Ala Lys Met
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                                             1115
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      Lys Gly Asn Asp Thr Ser Glu Met Ser His Asn Ser Ser Gln His Ile
                    1125
                                        1130
                                                            1135
     Glu Ser Asp Gln Gln Lys Asn Asp Met Lys Thr Val Gly Asp Leu Gly
                                    1145
                                                        1150
      Thr Thr His Val Gln Asn Glu Ile Ser Val Pro Val Thr Gly Glu Ile
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             1155
                                1160
                                                    1165
      Asp Glu Lys Leu Arg Glu Ser Lys Glu Ser Lys Ile His Lys Ala Glu
         1170
                            1175
                                                1180
     Glu Glu Arg Leu Ser His Thr Asp Ile His Lys Ile Asn Pro Glu Asp
                          1190
                                            1195
                                                                 1200
40
      Arg Asn Ser Asn Thr Leu His Leu Lys Asp Ile Arg Asn Glu Glu Asn
                     1205
                                        1210
     Glu Arg His Leu Thr Asn Gln Asn Ile Asn Ile Ser Gln Glu Arg Asp
                 1220
                                    1225
                                                        1230
      Leu Gln Lys His Gly Phe His Thr Met Asn Asn Leu His Gly Asp Gly
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            1235
                                 1240
                                                    1245
     Val Ser Glu Arg Ser Gln Ile Asn His Ser His His Gly Asn Arg Gln
                             1255
                                                1260
     Asp Arg Gly Gly Asn Ser Gly Asn Val Leu Asn Met Arg Ser Asn Asn
                          1270
                                             1275
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     Asn Asn Phe Asn Asn Ile Pro Ser Arg Tyr Asn Leu Tyr Asp Lys Lys
                    1285
                                        1290
                                                            1295
     Leu Asp Leu Asp Leu Tyr Glu Asn Arg Asn Asp Ser Thr Thr Lys Glu
                1300
                                    1305
                                                        1310
     Leu Ile Lys Lys Lou Ala Glu Ile Aen Lys Cys Glu Asn Glu Tle Ser
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            1315
                                1320
                                                    1325
     Val Lys Tyr Cys Asp His Met Ile His Glu Glu Ile Pro Leu Lys Thr
                            1335
        1330
                                                1340
     Cys Thr Lys Glu Lys Thr Arg Asn Leu Cys Cys Ala Val Ser Asp Tyr
                         1350
                                             1355
     Cys Met Ser Tyr Phe Thr Tyr Asp Ser Glu Glu Tyr Tyr Asn Cys Thr
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                                       1370
                    1365
                                                            1375
     Lys Arg Glu Phe Asp Asp Pro Ser Tyr Thr Cys Phe Arg Lys Glu Ala
                1380
                                    1385
                                                        1390
     Phe Ser Ser Met Ile Phe Lys Phe Leu Ile Thr Asn Lys Ile Tyr Tyr
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GCCGCTCT

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1395
                                   1400
      Tyr Phe Tyr Thr Tyr Lys Thr Ala Lys Val Thr Ile Lys Lys Ile Asn
                              1415
                                                    1420
      Phe Ser Leu Ile Phe Phe Phe Phe Ser Phe
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      1425
                          . 1430
     (2) INFORMATION FOR SEQ ID NO:5:
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           (i) SEQUENCE CHARACTERISTICS:
                (A) LENGTH: 2288 base pairs
                (B) TYPE: nucleic acid
                (C) STRANDEDNESS: single
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                (D) TOPOLOGY: linear
          (ii) MOLECULE TYPE: DNA (genomic)
        (iii) HYPOTHETICAL: NO
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         (vi) ORIGINAL SOURCE:
                (A) ORGANISM: Plasmodium falciparum
         (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:
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     CACTTTATGC TTCCGGCTCG TATGTTGTGT GGAATTGTGA GCGGATAACA ATTTCACACA 60
     GGAAACAGCT ATGACCATGA TTACGCCAAG CTCTAATACG ACTCACTATA GGGAAAGCTG 120
     GTACGCCTGC AGGTCCGGTC CGGAATTCAA TAAAATATTT CCAGAAAGGA ATGTGCAAAT 180
     TCACATATCC AATATATTCA AGGAATATAA AGAAAATAAT GTAGATATCA TATTTGGAAC 240
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     GTTGAATTAT GAATATAATA ATTTCTGTAA AGAAAAACCT GAATTAGTAT CTGCTGCCAA 300
     GTATAATCTG AAAGCTCCAA ATGCTAAATC CCCTAGAATA TACAAATCTA AGGAGCATGA 360
     AGAATCAAGT GTGTTTGGTT GCAAAACGAA AATCAGTAAA GTTAAAAAAA AATGGAATTG 420
     TTATAGTAAT AATAAAGTAA CTAAACCTGA AGGTGTATGT GGACCACCAA GAAGGCAACA 480
     ATTATGTCTT GGATATATAT TTTTGATTCG CGACGGTAAC GAGGAAGGAT TAAAAGATCA 540
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     TATTAATAAG GCAGCTAATT ATGAGGCAAT GCATTTAAAA GAGAAATATG AGAATGCTGG 600
     TGGTGATAAA ATTTGCAATG CTATATTGGG AAGTTATGCA GATATTGGAG ATATTGTAAG 660
     AGGTTTGGAT GTTTGGAGGG ATATAAATAC TAATAAATTA TCAGAAAAAT TCCAAAAAAT 720
     TTTTATGGGT GGTGGTAATT CTAGGAAAAA ACAAAACGAT AATAATGAAC GTAATAAATG 780
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     AACATGTAAA CGTCATAATA ATTTTGAGAA AATTCCTCAA TTTTTGAGAT GGTTAAAAGA 900
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     ATGGGGTGAT GAATTTTGTG AGGAAATGGG TACGGAAGTC AAGCAATTAG AGAAAATATG 960
     TGAAAATAAA AATTGTTCGG AAAAAAAATG TAAAAATGCA TGTAGTTCCT ATGAAAAATG 1020
     GATAAAGGAA CGAAAAAATG AATATAATTT GCAATCAAAG AAATTTGATA GTGATAAAAA 1080
     ATTAAATAA AAAAACAATC TTTATAATAA ATTTGAGGAT TCTAAAGCTT ATTTAAGGAG 1140
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     TGAATCAAAA CAGTGCTCAA ATATAGAATT TAATGATGAA ACATTTACAT TTCCTAATAA 1200
     ATATAAAGAG GCTTGTATGG TATGTGAAAA TCCTTCATCT TCGAAAGCTC TTAAACCTAT 1260
     AAAAACGAAT GTGTTTCCTA TAGAGGAATC AAAAAAATCT GAGTTATCAA GTTTAACAGA 1320
     TAAATCTAAG AATACTCCTA ATAGTTCTGG TGGGGGAAAT TATGGAGATA GACAAATATC 1380
     AAAAAGAGAC GATGTTCATC ATGATGGTCC TAAGGAAGTG AAATCCGGAG AAAAAGAGGT 1440
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     ACCAAAAATA GATGCAGCTG TTAAAACAGA AAATGAATTT ACCTCTAATC GAAACGATAT 1500
     TGAAGGAAAG GAAAAAAGTA AAGGTGATCA TTCTTCTCCT GTTCATTCTA AAGATATAAA 1560
     AAATGAGGAA CCACAAAGGG TGGTGTCTGA AAATTTACCT AAAATTGAAG AGAAAATGGA 1620
     ATCTTCTGAT TCTATACCAA TTACTCATAT AGAAGCTGAA AAGGGTCAGT CTTCTAATTC 1680
     TAGCGATAAT GATCCTGCAG TAGTAAGTGG TAGAGAATCT AAAGATGTAA ATCTTCATAC 1740
TTCTGAAAGG ATTAAAGAAA ATGAAGAAGG TGTGATTAAA ACAGATGATA GTTCAAAAAG 1800
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     TATTGAAATT TCTAAAATAC CATCTGACCA AAATAATCAT AGTGATTTAT CACAGAATGC 1860
     AAATGAGGAC TCTAATCAAG GGAATAAGGA AACAATAAAT CCTCCTTCTA CAGAAAAAA 1920
     TCTCAAAGAA ATTCATTATA AAACATCTGA TTCTGATGAT CATGGTTCTA AAATTAAAAG 1980
     TGAAATTGAA CCAAAGGAGT TAACGGAGGA ATCACCTCTT ACTGATAAAA AAACTGAAAG 2040
60
     TGCAGCGATT GGTGATAAAA ATCATGAATC AGTAAAAAGC GCTGATATTT TTCAATCTGA 2100
     GATTCATAAT TCTGATAATA GAGATAGAAT TGTTTCTGAA AGTGTAGTTC AGGATTCTTC 2160
     AGGAAGCTCT ATGAGTACTG AATCTATACG TACTGATAAC AAGGATTTTA AAACAAGTGA 2220
     GGATATTGCA CCTTCTATTA ATGGTCGGAA TTCCCGGGTC GACGAGCTCA CTAGTCGGCG 2280
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(2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 749 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (iii) HYPOTHETICAL: NO
 - (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: Plasmodium falciparum
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:
- - Ser Ser Asn Thr Thr His Tyr Arg Glu Ser Trp Tyr Ala Cys Arg Ser 20 25 30
 - Gly Pro Glu Phe Asn Lys Ile Phe Pro Glu Arg Asn Val Gln Ile His
 35 40 45
- Ile Ser Asn Ile Phe Lys Glu Tyr Lys Glu Asn Asn Val Asp Ile Ile
 50 55 60
 - Phe Gly Thr Leu Asn Tyr Glu Tyr Asn Asn Phe Cys Lys Glu Lys Pro
 65 70 75 80
 Glu Leu Val Ser Ala Ala Lys Tyr Asn Leu Lys Ala Pro Asn Ala Lys
- 85 90 95 30 Ser Pro Arg Ile Tyr Lys Ser Lys Glu His Glu Glu Ser Ser Val Phe
 - 100 105 110

 Gly Cys Lys Thr Lys Ile Ser Lys Val Lys Lys Trp Asn Cys Tyr
 - 115 120 125 Ser Asn Asn Lys Val Thr Lys Pro Glu Gly Val Cys Gly Pro Pro Arg
 - 130 135 140
 Arg Gln Gln Leu Cys Leu Gly Tyr Ile Phe Leu Ile Arg Asp Gly Asn
 - Glu Glu Gly Leu Lys Asp His Ile Asn Lys Ala Ala Asn Tyr Glu Ala 165 170 175
- 40 Met His Leu Lys Glu Lys Tyr Glu Asn Ala Gly Gly Asp Lys Ile Cys
 180 185 190
 - Asn Ala Ile Leu Gly Ser Tyr Ala Asp Ile Gly Asp Ile Val Arg Gly
 195 200 205
- Leu Asp Val Trp Arg Asp Ile Asn Thr Asn Lys Leu Ser Glu Lys Phe
 210 215 220
 - Gln Lys Ile Phe Met Gly Gly Gly Asn Ser Arg Lys Lys Gln Asn Asp 225 230 235 240
 - Asn Asn Glu Arg Asn Lys Trp Trp Glu Lys Gln Arg Asn Leu Ile Trp

 245
 250
 255
 - Ser Ser Met Val Lys His Ile Pro Lys Gly Lys Thr Cys Lys Arg His
 - Asn Asn Phe Glu Lys Ile Pro Gln Phe Leu Arg Trp Leu Lys Glu Trp
 275 280 285
- Gly Asp Glu Phe Cys Glu Glu Met Gly Thr Glu Val Lys Gln Leu Glu
 290 295 300
 - Lys Ile Cys Glu Asn Lys Asn Cys Ser Glu Lys Lys Cys Lys Asn Ala 305 310 315 320

Ser Lys Gln Cys Ser Asn Ile Glu Phe Asn Asp Glu Thr Phe Thr Phe

- Cys Ser Ser Tyr Glu Lys Trp Ile Lys Glu Arg Lys Asn Glu Tyr Asn 325 330 335 Leu Gln Ser Lys Lys Phe Asp Ser Asp Lys Lys Leu Asn Lys Lys Asn
- 340 345 350
 Asn Leu Tyr Asn Lys Phe Glu Asp Ser Lys Ala Tyr Leu Arg Ser Glu
 355 360 365

			370						_								
		Pro			Tvr	Lve	Glu	37 הוג	5	M - 4-	** . *	_	_ 3	80			
		385		_,_	- 7 -	Lys	390	Ald	cys	Met	vai	Cys	Glu	Asn	Pro	Ser	Ser
5		Ser	Lys	Ala	Leu	Lys	Pro	Ile	Lys	Thr	Asn	395 Val	Phe	Pro	Ile	Glu	400 Glu
		Ser	Lys	Lys	Ser 420	Glu	Leu	Ser	Ser	Leu 42	41 Thr	Asp	Lys	Ser	Lys	4 Asn	15 Thr
		Pro	Asn	Ser 435	Ser	Gly	Gly	Gly	Asn 44	Tyr	Gly	Asp	Arg	Gln	4 Ile	30 Ser	Lys
10									Gly	Pro				Lys	45 Ser		
		Lys	Glu	Val	Pro	Lys	Ile	Asp	Ala	Ala	Val	Lvs	Thr	60 Glu	Asn	01	5 1-
15															Lys		
		His	Ser	Ser	Pro 500	Val	His	Ser	Lys	Asp 50	Ile 5	Lys	Asn	Glu	Glu	Pro	Gln
20															5: Met 25		
20														Lys	Gly		
		5er 545	Asn	Ser	Ser	Asp	Asn 550	Asp	Pro	Ala	Val	Val	Ser	Gly	Arg	Glu	Ser
25						Leu	His								Asn		
						Thr				Ser	Lys				Ile		
30																	
		Glu	610 Lvs	Δen	T.e.11	Lvc	Clu	615	шуs 5	GIU	Inr	TTE	Asn 62	Pro	Pro	Ser	Thr
															Ser		
35											Glu	Pro			Leu		
					000					n n .	Glu	Ser			Ile	Gly	
40										Ala	Asp				67 Ser	Glu	
40									Arg	Ile					Val '		
•								Met	Ser					Arg	Thr .		
45		Lys									Ala	Pro	Ser	Ile.		Gly .	
		Asn	Ser	Arg	Val 740	Asp	Glu	Leu	Thi	Ser 745	Arg	Arg	g Pr	o Le	u	73	5
50	(2)	INFOR	MATI	ON I	FOR S	SEQ :	ID N	0:7:						•			
		(i)	(A)	LEN TYE	E CHA NGTH: PE: r	260 ucle	06 b ≥ic	ase acid	pair	s		•					
55			(C)	STF	POLOG	EDNES SY:]	SS: line	sing ar	le								

- (ii) MOLECULE TYPE: DNA (genomic)
- (iii) HYPOTHETICAL: NO

- (vi) ORIGINAL SOURCE:
 (A) ORGANISM: Plasmodium falciparum
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

```
AGCTCTATTA CGACTCACTA TAGGGAAAGC TGGTACGCCT GCAGGTACCG GTCCGGAATT 60
      CCCGGGTCGA CGAGCTCACT AGTCGGCGGC CGCTCTAGAG GATCCAAGCT TAATAGTGTT 120
      TATACGTCTA TTGGCTTATT TTTAAATAGC TTAAAAAGCG GACCATGTAA AAAGGATAAT 180
     GATAATGCAG AGGATAATAT AGATTTTGGT GATGAAGGTA AAACATTTAA AGAGGCAGAT 240
     AATTGTAAAC CATGTTCTCA ATTTACTGTT GATTGTAAAA ATTGTAATGG TGGTGATACA 300
     AAAGGGAAGT GCAATGGCAG CAATGGCAAA AAGAATGGAA ATGATTATAT TACTGCAAGT 360
     GATATTGAAA ATGGAGGGAA TTCTATTGGA AATATAGATA TGGTTGTTAG TGATAAGGAT 420
     GCAAATGGAT TTAATGGTTT AGACGCTTGT GGAAGTGCAA ATATCTTTAA AGGTATTAGA 480
     AAAGAACAAT GGAAATGTGC TAAAGTATGT GGTTTAGATG TATGTGGTCT TAAAAATGGT 540
     AATGGTAGTA TAGATAAAGA TCAAAAACAA ATTATAATTA TTAGAGCATT GCTTAAACGT 600
10
     TGGGTAGAAT ATTTTTTAGA AGATTATAAT AAAATTAATG CCAAAATTTC ACATTGTACG 660
     AAAAAGGATA ATGAATCCAC ATGTACAAAT GATTGTCCAA ATAAATGTAC ATGTGTAGAA 720
     GAGTGGATAA ATCAGAAAAG GACAGAATGG AAAAATATAA AAAAACATTA CAAAACACAA 780
     AATGAAAATG GTGACAATAA CATGAAATCT TTGGTTACAG ATATTTTGGG TGCCTTGCAA 840
     CCCCAAAGTG ATGTTAACAA AGCTATAAAA CCTTGTAGTG GTTTAACTGC GTTCGAGAGT 900
15
     TTTTGTGGTC TTAATGGCGC TGATAACTCA GAAAAAAAG AAGGTGAAGA TTACGATCTT 960
     GTTCTATGTA TGCTTAAAAA TCTTGAAAAA CAAATTCAGG AGTGCAAAAA GAAACATGGC 1020
     GAAACTAGTG TCGAAAATGG TGGCAAATCA TGTACCCCCC TTGACAACAC CACCCTTGAG 1080
     GAGGAACCCA TAGAAGAGGA AAACCAAGTG GAAGCGCCGA ACATTTGTCC AAAACAAACA 1140
     GTGGAAGATA AAAAAAAGA GGAAGAAGAA GAAACTTGTA CACCGGCATC ACCAGTACCA 1200
     GAAAAACCGG TACCTCATGT GGCACGTTGG CGAACATTTA CACCACCTGA GGTATTCAAG 1260
     ATATGGAGGG GAAGGAGAA TAAAACTACG TGCGAAATAG TGGCAGAAAT GCTTAAAGAT 1320
AAGAATGGAA GGACTACAGT AGGTGAATGT TATAGAAAAG AAACTTATTC TGAATGGACG 1380
     TGTGATGAAA GTAAGATTAA AATGGGACAG CATGGAGCAT GTATTCCTCC AAGAAGACAA 1440
     AAATTATGTT TACATTATTT AGAAAAAATA ATGACAAATA CAAATGAATT GAAATACGCA 1500
25
     TTTATTAAAT GTGCTGCAGC AGAAACTTTT TTGTTATGGC AAAACTACAA AAAAGATAAG 1560
AATGGTAATG CAGAAGATCT CGATGAAAAA TTAAAAGGTG GTATTATCCC CGAAGATTTT 1620
     AAACGGCAAA TGTTCTATAC GTTTGCAGAT TATAGAGATA TATGTTTGGG TACGGATATA 1680
     TCATCAAAAA AAGATACAAG TAAAGGTGTA GGTAAAGTAA AATGCAATAT TGATGATGTT 1740
30
     TTTTATAAAA TTAGCAATAG TATTCGTTAC CGTAAAAGTT GGTGGGAAAC AAATGGTCCA 1800
     GTTATATGGG AAGGAATGTT ATGCGCTTTA AGTTATGATA CGAGCCTAAA TAATGTTAAT 1860
     CCGGAAACTC ACAAAAAACT TACCGAAGGC AATAACAACT TTGAGAAAGT CATATTTGGT 1920
     AGTGATAGTA GCACTACTTT GTCCAAATTT TCTGAAAGAC CTCAATTTCT AAGATGGTTG 1980
     ACTGAATGGG GAGAAAATTT CTGCAAAGAA CAAAAAAAGG AGTATAAGGT GTTGTTGGCA 2040
35
     AAATGTAAGG ATTGTGATGT TGATGGTGAT GGTAAATGTA ATGGAAAATG TGTTGCGTGC 2100
     AAAGATCAAT GTAAACAATA TCATAGTTGG ATTGGAATAT GGATAGATAA TTATAAAAAA 2160
     CAAAAAGGAA GATATACTGA GGTTAAAAAA ATACCTCTGT ATAAAGAAGA TAAAGACGTG 2220
     AAAAACTCAG ATGATGCTCG CGATTATTTA AAAACACAAT TACAAAATAT GAAATGTGTA 2280
     AATGGAACTA CTGATGAAAA TTGTGAGTAT AAGTGTATGC ATAAAACCTC ATCCACAAAT 2340
     AGTGATATGC CCGAATCGTT GGACGAAAAG CCGGAAAAGG TCAAAGACAA GTGTAATTGT 2400
40
     GTACCTAATG AATGCAATGC ATTGAGTGTA AGTGGTAGCG GTTTTCCTGA TGGTCAAGCT 2460
     TACGTACGCG TGCATGCGAC GTCATAGCTC TTCTATAGTG TCACCTAAAT TCAATTCACT 2520 GGCCGTCGTT TTACAACGTC GTGACTGGGA AAACCTGGCG TTACCCCAACT TAATCGCCTT 2580
     GCAGCACATC CCCCTTTCGC CAGCTG
45
     (2) INFORMATION FOR SEQ ID NO:8:
```

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 921 amino acids

- (B) TYPE: amino acid(C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
 - (iii) HYPOTHETICAL: NO

50

55

- (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: Plasmodium falciparum
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:
 - Lys Leu Asn Ser Val Tyr Thr Ser Ile Gly Leu Phe Leu Asn Ser Leu 10

				20					Asn 25						^	
	Asp	Phe	Gly 35	Asp	Glu	Gly	Lys	Thr	Phe	Lys	Glu	Ala	Asp	Asn	Cys	Lys
5		20					22	Asp	Cys				Λ			
						70					7/5					Asp 80
10					0.5				Asn	9.0)				0	Asn
				100	,				Asp 10	5				7	3 0	
15			- 110	l.				- 12	Phe 0				7 '	Lys	Glu	
15		100					13.	⊃	Leu			74	Gly	Leu		
	T#2					150			Gln		155					100
20					T 0 2	,			Tyr	17	'n					Lys
				100	,				Thr 18	5				1	മറ	
25			エココ					20					21	<u> </u>		
25		210					21:	5	Asn		-	2:	20			
·	423					230			Met		235					240
30					245	•			Asp	25	0				2	C C
				260	l .				Ser 26	5				2.	70	
35			2/5					28	Glu 0				2.5	2 5		
		290					29	-	Ile			3 () ()			
	202					310			Gly		315					330
40					325				Ile	33	0				2	2 =
				340					Thr 34	5				3 1	50	
45			333					36	Ala 0				3.6	5 5		
		3/0					375	>	Thr			3,5	20			
	303					390			Lys		395					400
50					405				Arg	41	ο,				4	1 5
				420					Thr 42	5				47	٩n	
55			435					44					44	15		_
		450					455	5	Thr			46	. 0			_
	400					470			Glu		475					400
60					485				Ala	49	0				4	0.5
	Lys			500					509	5				51	Λ.	
	LIIE	AIG	515	ıyr	Arg	Asp	тте	Cys 520	Leu	Gly	Thr	Asp	Ile 52		Ser	Lys

	Lys	Asp 530	Thr	Ser	Lys	Gly	Val 53	Gly	Lys	Val	Lys			Ile	Asp	Asp
	Val 545			Lys	Ile	Ser 550	Asn	Ser	Ile	Arg	Tyr	Arg	10 Lys	Ser	Trp	Trp
5		Thr	Asn	Gly	Pro 565	Val	Ile	Trp	Glu	Gly	Met	Leu	Cys	Ala	_	
	Tyr	Asp	Thr	Ser 580	Leu		Asn	Val	Asn 58	57 Pro	Glu	Thr	His	Lys	Lys	75 Leu
. 10	Thr	Glu	Gly 595	Asn		Asn	Phe	Glu 60	Lys	Val	Ile	Phe		Ser	90 Asp	Ser
	Ser	Thr 610			Ser	Lys	Phe 61	Ser	Glu	Arg	Pro		Phe	05 Leu	Arg	Trp
	Leu 625		Glu	Trp	Gly	Glu	Asn	Phe	Cys	Lys	Glu	Gln	Lys	Lys	Glu	Tyr
15		Val	Leu	Leu	Ala 645	630 Lys	Cys	Lys	Asp	Cys	Asp	Val	Asp	Gly	Asp	640 Gly
	Lys	Cys	Asn	Gly 660	Lys		Val	Ala	Cys 66	65 Lys	Asp	Gln	Cys	Lys	Gln	55 Tyr
20	His	Ser	Trp 675	Ile		Ile	Trp	Ile 68	Asp	Asn	Tyr	Lys		Gln 85	70 Lys	Gly
		050					69:	Ile 5	Pro			70	Glu	Asp		
	Val 705	Lys	Asn	Ser	Asp	Asp 710	Ala	Arg	Asp	Tyr	Leu 715	Lys	Thr	Gln	Leu	Gln 720
25					125					73	Glu 0			Glu	7	Lys
	Cys	Met.	His	Lys 740	Thr	Ser	Ser	Thr	Asn 74	Ser	Asp	Met	Pro		Ser 50	Leu
30	Asp	Glu	Lys 755	Pro	Glu	Lys	Val	Lys	Asp	Lys	Cys	Asn		Val 55	Pro	Asn
,	Glu	Cys 770	Asn	Ala	Leu	Ser	Val 775	Ser	Gly	Ser	Gly	Phe 78	Pro	Asp	Gly	Gln
	Ala 785	Phe	Gly	Gly	Gly	Val 790	Leu	Glu	Gly	Thr	Cys 795	Lys	Gly	Leu		
35	Pro	Lys	Lys	Lys	Ile 805	Glu	Pro	Pro	Gln	Tyr 81	Asp	Pro	Thr	Asn	Asp	
	Leu	Lys	Ser	Thr 820			Val		Ile 82	Val	Leu	Ala	Leu	Gly	Ser	15 Ile
40	Ala	Phe	Leu 835		Met	Lys	Val	Ile 840	Tyr	Ile	Tyr	Val		Tyr	Ile	Tyr
٠	Met	Leu 850		Val	Gly	Ala	Leu 859	Asp	Thr	Tyr	Ile		Gly	15 Cys	Ile	Cys
	Ile 865	Cys	Ile	Phe	Ile	Cys 870	Val	Ser	Val	Tyr	Val	86 Cys	Val	Tyr		
45			Leu	Tyr	Met 885	Cys	Val	Phe	Tyr	Ile	875 Tyr	Phe	Ile	Tyr	Ile	
	Val	Phe	Ile	Leu 900		Met	Lys	Lys	Met	89 Lys	Lys	Met	Lys	Lys	Met	95 Lys
50	Lys	Met	Lys 915	Lys	Arg	Lys	Lys	920						91	_0	

(2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2101 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

- 60 (ii) MOLECULE TYPE: DNA (genomic)
 - (iii) HYPOTHETICAL: NO
 - (vi) ORIGINAL SOURCE:

(A) ORGANISM: Plasmodium falciparum

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

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5
     GGAACAGGGT GATAATAAAG TAGGAGCCTG TGCTCCGTAT AGACGATTAC ATTTATGTGA 60
     TTATAATTTG GAATCTATAG ACACAACGTC GACGACGCAT AAGTTGTTGT TAGAGGTGTG 120
     TATGGCAGCA AAATACGAAG GAAACTCAAT AAATACACAT TATACACAAC ATCAACGAAC 180
     TAATGAGGAT TCTGCTTCCC AATTATGTAC TGTATTAGCA CGAAGTTTTG CAGATATAGG 240
     TGATATCGTA AGAGGAAAAG ATCTATATCT CGGTTATGAT AATAAAGAAA AAGAACAAAG 300
     AAAAAATTA GAACAGAAAT TGAAAGATAT TTTCAAGAAA ATACATAAGG ACGTGATGAA 360
10
     GACGAATGGC GCACAAGAAC GCTACATAGA TGATGCCAAA GGAGGAGATT TTTTTCAATT 420
     AAGAGAAGAT TGGTGGACGT CGAATCGAGA AACAGTATGG AAAGCATTAA TATGTCATGC 480
     ACCAAAAGAA GCTAATTATT TTATAAAAAC AGCGTGTAAT GTAGGAAAAG GAACTAATGG 540
     TCAATGCCAT TGCATTGGTG GAGATGTTCC CACATATTTC GATTATGTGC CGCAGTATCT 600
15
     TCGCTGGTTC GAGGAATGGG CAGAAGACTT TTGCAGGAAA AAAAAAAAA AACTAGAAAA 660
     TTTGCAAAAA CAGTGTCGTG ATTACGAACA AAATTTATAT TGTAGTGGTA ATGGCTACGA 720
     TTGCACAAAA ACTATATATA AAAAAGGTAA ACTTGTTATA GGTGAACATT GTACAAACTG 780
     TTCTGTTTGG TGTCGTATGT ATGAAACTTG GATAGATAAC CAGAAAAAG AATTTCTAAA 840
     ACAAAAAAGA AAATACGAAA CAGAAATATC AGGTGGTGGT AGTGGTAAGA GTCCTAAAAG 900
20
     GACAAAACGG GCTGCACGTA GTAGTAGTAG TAGTGATGAT AATGGGTATG AAAGTAAATT 960
     TTATAAAAA CTGAAAGAAG TTGGCTACCA AGATGTCGAT AAATTTTTAA AAATATTAAA 1020
     CAAAGAAGGA ATATGTCAAA AACAACCTCA AGTAGGAAAT GAAAAAGCAG ATAATGTTGA 1080
     TTTTACTAAT GAAAAATATG TAAAAACATT TTCTCGTACA GAAATTTGTG AACCGTGCCC 1140
     ATGGTGTGGA TTGGAAAAAG GTGGTCCACC ATGGAAAGTT AAAGGTGACA AAACCTGCGG 1200
25
     AAGTGCAAAA ACAAAGACAT ACGATCCTAA AAATATTACC GATATACCAG TACTCTACCC 1260
     TGATAAATCA CAGCAAAATA TACTAAAAAA ATATAAAAAT TTTTGTGAAA AAGGTGCACC 1320
     TGGTGGTGGT CAAATTAAAA AATGGCAATG TTATTATGAT GAACATAGGC CTAGTAGTAA 1380
     AAATAATAAT AATTGTGTAG AAGGAACATG GGACAAGTTT ACACAAGGTA AACAAACCGT 1440
     TAAGTCCTAT AATGTTTTTT TTTGGGATTG GGTTCATGAT ATGTTACACG ATTCTGTAGA 1500
30
     GTGGAAGACA GAACTTAGTA AGTGTATAAA TAATAACACT AATGGCAACA CATGTAGAAA 1560
     CAATAATAAA TGTAAAACAG ATTGTGGTTG TTTTCAAAAA TGGGTTGAAA AAAAACAACA 1620
     AGAATGGATG GCAATAAAAG ACCATTTTGG AAAGCAAACA GATATTGTCC AACAAAAAGG 1680
     TCTTATCGTA TTTAGTCCCT ATGGAGTTCT TGACCTTGTT TTGAAGGGCG GTAATCTGTT 1740
GCAAAATATT AAAGATGTTC ATGGAGATAC AGATGACATA AAACACATTA AGAAACTGTT 1800
GGATGAGGAA GACGCAGTAG CAGTTGTTCT TGGTGGCAAG GACAATACCA CAATTGATAA 1860
35
     ATTACTACAA CACGAAAAAG AACAAGCAGA ACAATGCAAA CAAAAGCAGG AAGAATGCGA 1920
     GAAAAAAGCA CAACAAGAAA GTCGTGGTCG CTCCGCCGAA ACCCGCGAAG ACGAAAGGAC 1980
     ACAACAACCT GCTGATAGTG CCGGCGAAGT CGAAGAAGAA GAAGACGACG ACGACTACGA 2040
     CGAAGACGAC GAAGATGACG ACGTAGTCCA GGACGTAGAT GTAAGTGAAA TAAGAGGTCC 2100
40
```

(2) INFORMATION FOR SEQ ID NO:10:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 700 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- 50 (ii) MOLECULE TYPE: protein

- (iii) HYPOTHETICAL: NO
- (vi) ORIGINAL SOURCE:

 (A) ORGANISM: Plasmodium falciparum
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:
- Glu Gln Gly Asp Asn Lys Val Gly Ala Cys Ala Pro Tyr Arg Arg Leu

 1 5 10 15

 His Leu Cys Asp Tyr Asn Leu Glu Ser Ile Asp Thr Thr Ser Thr Thr

 20 25 30

 His Lys Leu Leu Leu Glu Val Cys Met Ala Ala Lys Tyr Glu Gly Asn
 35 40

	Ser	Ile 50	Asn	Thr	His	Tyr	Thr 55	Gln	His	Gln	Arg			Glu	Asp	Ser
	Ala 65	Ser	Gln	Leu	Cys	Thr	Val	Leu	Ala	Arg	Ser 75	Phe	Ala	Asp	Ile	Gly
5				Arg	85					90	Gly				0.5	
				Arg 100					10	5				11	Phe	Lys
10			112					120)				12	5		
		T30		Ala			135	5				14	0		_	_
15	145			Asn		150					155					160
				Ala	165					17	0				7 -	75
				Gly 180					18:	5				19	0	
20			195					200)				20	5		
		210		Arg Tyr			215	;				22	0		_	
25	225			Thr		230					235					240
				Cys	245	•				25	0		•		2 5	: 5
				260 Lys		•			26	5	•			27	0	,
30			275	Gly				280)				2.8	5		
		290		Ser			295	•				30	0			
35	305			Leu	Lys	Glu					315					320
	Lys	Ile	Leu	Asn 340	325 Lys		Gly	Ile	Cys	- 3:30 Gln	Lys Lys	Gln	Pro	Gln	33 Val	Gly
40	Asn	Glu	Lys 355	Ala	Asp	Asn	Val	Asp 360			Asn	Glu		_	0 Val	Lys
	Thr	Phe 370		Arg	Thr	Glu	Ile 375	Cys		Pro	Cys	Pro 38	36 Trp	Cys	Gly	Leu
	Glu 385		Gly	Gly	Pro	Pro 390	Trp	Lys	Val	Lys	Gly 395	Asp	Lys	Thr	Cys	
45		Ala	Lys	Thr	Lys 405	Thr	Tyr	Asp	Pro	Lys 410	Asn	Ile	Thr	Asp	Ile 41	
				Pro 420					425	Asn	Ile			43	Tyr 0	Lys
50			435	Glu				440)		•		44	5		
		450		Tyr			455					460	0			
EE	465			Gly		470					475				•	490
55				Asn	485					490)				49	5
				Glu 500					505	5				51	0	
60			515	Asn				520)				52	5		
		530		Gln			535					540	0	_		
	545	- Lys	vəħ	His	FIIG	550	пÀг	GIU	ıuŗ		11e 555	val	GIN	GIn	ГÀ2	Gly 560

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Leu Ile Val Phe Ser Pro Tyr Gly Val Leu Asp Leu Val Leu Lys Gly
                           565
                                               570
          Gly Asn Leu Leu Gln Asn Ile Lys Asp Val His Gly Asp Thr Asp Asp
                                           585
5
          Ile Lys His Ile Lys Lys Leu Leu Asp Glu Glu Asp Ala Val Ala Val
                                       600
          Val Leu Gly Gly Lys Asp Asn Thr Thr Ile Asp Lys Leu Leu Gln His
                                   615
          Glu Lys Glu Gln Ala Glu Gln Cys Lys Gln Lys Gln Glu Glu Cys Glu
10
                              630
                                                  635
          Lys Lys Ala Gln Gln Glu Ser Arg Gly Arg Ser Ala Glu Thr Arg Glu
                           645
          Asp Glu Arg Thr Gln Gln Pro Ala Asp Ser Ala Gly Glu Val Glu Glu
                                           665
          Glu Glu Asp Asp Asp Asp Tyr Asp Glu Asp Asp Glu Asp Asp Val
15
                                       680
          Val Gln Asp Val Asp Val Ser Glu Ile Arg Gly Pro
                                   695
20
     (2) INFORMATION FOR SEQ ID NO:11:
```

- - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 8220 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
- 30 (iii) HYPOTHETICAL: NO
 - (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: Plasmodium falciparum
- 35 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

AAAAATGGGG CCCAAGGAGG CTGCAGGTGG GGATGATATT GAGGATGAAA GTGCCAAACA 60 TATGTTTGAT AGGATAGGAA AAGATGTGTA CGATAAAGTA AAAGAGGAAG CTAAAGAACG 120 TGGTAAAGGC TTGCAAGGAC GTTTGTCAGA AGCAAAATTT GAGAAAAATG AAAGCGATCC 180 40 ACAAACACCA GAAGATCCAT GCGATCTTGA TCATAAATAT CATACAAATG TAACTACTAA 240 TGTAATTAAT CCGTGCGCTG ATAGATCTGA CGTGCGTTTT TCCGATGAAT ATGGAGGTCA 300 ATGTACACAT AATAGAATAA AAGATAGTCA ACAGGGTGAT AATAAAGGTG CATGTGCTCC 360 ATATAGGCGA TTGCATGTAT GCGATCAAAA TTTAGAACAG ATAGAGCCTA TAAAAATAAC 420 AAATACTCAT AATTTATTGG TAGATGTGTG TATGGCAGCA AAATTTGAAG GACAATCAAT 480 45 AACACAAGAT TATCCAAAAT ATCAAGCAAC ATATGGTGAT TCTCCTTCTC AAATATGTAC 540 TATGCTGGCA CGAAGTTTTG CGGACATAGG GGACATTGTC AGAGGAAGAG ATTTGTATTT 600 AGGTAATCCA CAAGAAATAA AACAAAGACA ACAATTAGAA AATAATTTGA AAACAATTTT 660 CGGGAAAATA TATGAAAAAT TGAATGGCGC AGAAGCACGC TACGGAAATG ATCCGGAATT 720 TTTTAAATTA CGAGAAGATT GGTGGACTGC TAATCGAGAA ACAGTATGGA AAGCCATCAC 780 50 ATGTAACGCT TGGGGTAATA CATATTTTCA TGCAACGTGC AATAGAGGAG AACGAACTAA 840 AGGTTACTGC CGGTGTAACG ACGACCAAGT TCCCACATAT TTTGATTATG TGCCGCAGTA 900 TCTTCGCTGG TTCGAGGAAT GGGCAGAAGA TTTTTGTAGG AAAAAAAAA AAAAAATAAA 960 AGATGTTAAA AGAAATTGTC GTGGAAAAGA TAAAGAGGAT AAGGATCGAT ATTGTAGCCG 1020 TAATGGCTAC GATTGCGAAA AAACTAAACG AGCGATTGGT AAGTTGCGTT ATGGTAAGCA 1080 55 ATGCATTAGC TGTTTGTATG CATGTAATCC TTACGTTGAT TGGATAAATA ACCAAAAAGA 1140 ACAATTTGAC AAACAGAAAA AAAAATATGA TGAAGAAATA AAAAAATATG AAAATGGAGC 1200 ATCAGGTGGT AGTAGGCAAA AACGGGATGC AGGTGGTACA ACTACTACTA ATTATGATGG 1260 ATATGAAAAA AAATTTTATG ACGAACTTAA TAAAAGTGAA TATAGAACCG TTGATAAATT 1320 TTTGGAAAAA TTAAGTAATG AAGAAATATG CACAAAAGTT AAAGACGAAG AAGGAGGAAC 1380 60 AATTGATTTT AAAAACGTTA ATAGTGATAG TACTAGTGGT GCTAGTGGCA CTAATGTTGA 1440 AAGTCAAGGA ACATTTTATC GTTCAAAATA TTGCCAACCC TGCCCTTATT GTGGAGTGAA 1500 AAAGGTAAAT AATGGTGGTA GTAGTAATGA ATGGGAAGAG AAAAATAATG GCAAGTGCAA 1560 GAGTGGAAAA CTTTATGAGC CTAAACCCGA CAAAGAAGGT ACTACTATTA CAATCCTTAA 1620 AAGTGGTAAA GGACATGATG ATATTGAAGA AAAATTAAAC AAATTTTGTG ATGAAAAAA 1680

TGGTGATACA ATAAATAGTG GTGGTAGTGG TACGGGTGGT AGTGGTGGTG GTAACAGTGG 1740 TAGACAGGAA TTGTATGAAG AATGGAAATG TTATAAAGGT GAAGATGTAG TGAAAGTTGG 1800 ACACGATGAG GATGACGAGG AGGATTATGA AAATGTAAAA AATGCAGGCG GATTATGTAT 1860 ATTAAAAAC CAAAAAAAGA ATAAAGAAGA AGGTGGAAAT ACGTCTGAAA AGGAGCCTGA 1920 TGAAATCCAA AAGACATTCA ATCCTTTTT TTACTATTGG GTTGCACATA TGTTAAAAGA 1980 TTCCATACAT TGGAAAAAA AACTTCAGAG ATGTTTACAA AATGGTAACA GAATAAAATG 2040 TGGAAACAAT AAATGTAATA ATGATTGTGA ATGTTTTAAA AGATGGATTA CACAAAAAA 2100 AGACGAATGG GGGAAAATAG TACAACATTT TAAAACGCAA AATATTAAAG GTAGAGGAGG 2160 TAGTGACAAT ACGGCAGAAT TAATCCCATT TGATCACGAT TATGTTCTTC AATACAATTT 2220 10 GCAAGAAGAA TTTTTGAAAG GCGATTCCGA AGACGCTTCC GAAGAAAAAT CCGAAAATAG 2280 TCTGGATGCA GAGGAGCAG AGGAACTAAA ACACCTTCGC GAAATCATTG AAAGTGAAGA 2340 CAATAATCAA GAAGCATCTG TTGGTGGTGG CGTCACTGAA CAAAAAAATA TAATGGATAA 2400 ATTGCTCAAC TACGAAAAAG ACGAAGCCGA TTTATGCCTA GAAATTCACG AAGATGAGGA 2460 AGAGGAAAAA GAAAAAGGAG ACGGAAACGA ATGTATCGAA GAGGGCGAAA ATTTTCGTTA 2520 TAATCCATGT AGTGGCGAAA GTGGTAACAA ACGATACCCC GTTCTTGCGA ACAAAGTAGC 2580 15 GTATCAAATG CATCACAAGG CAAAGACACA ATTGGCTAGT CGTGCTGGTA GAAGTGCGTT 2640 GAGAGGTGAT ATATCCTTAG CGCAATTTAA AAATGGTCGT AACGGAAGTA CATTGAAAGG 2700 ACAAATTTGC AAAATTAACG AAAACTATTC CAATGATAGT CGTGGTAATA GTGGTGGACC 2760 ATGTACAGGC AAAGATGGAG ATCACGGAGG TGTGCGCATG AGAATAGGAA CGGAATGGTC 2820 20 AAATATTGAA GGAAAAAAC AAACGTCATA CAAAAACGTC TTTTTACCTC CCCGACGAGA 2880 ACACATGTGT ACATCCAATT TAGAAAATTT AGATGTTGGT AGTGTCACTA AAAATGATAA 2940 GGCTAGCCAC TCATTATTGG GAGATGTTCA GCTCGCAGCA AAAACTGATG CAGCTGAGAT 3000 AATAAAACGC TATAAAGATC AAAATAATAT ACAACTAACT GATCCAATAC AACAAAAAGA 3060 CCAGGAGGCT ATGTGTCGAG CTGTACGTTA TAGTTTTGCC GATTTAGGAG ACATTATTCG 3120 AGGAAGAGAT ATGTGGGATG AGGATAAGAG CTCAACAGAC ATGGAAACAC GTTTGATAAC 3180 25 CGTATTTAAA AACATTAAAG AAAAACATGA TGGAATCAAA GACAACCCTA AATATACCGG 3240 TGATGAAAGC AAAAAGCCCG CATATAAAAA ATTACGAGCA GATTGGTGGG AAGCAAATAG 3300 ACATCAAGTG TGGAGAGCCA TGAAATGCGC AACAAAAGGC ATCATATGTC CTGGTATGCC 3360 AGTTGACGAT TATATCCCCC AACGTTTACG CTGGATGACT GAATGGCTG AATGGTATTG 3420 TAAAGCGCAA TCACAGGAGT ATGACAAGTT AAAAAAAATC TGTGCAGATT GTATGAGTAA 3480 30 GGGTGATGGA AAATGTACGC AAGGTGATGT CGATTGTGGA AAGTGCAAAG CAGCATGTGA 3540 TAAATATAAA GAGGAAATAG AAAAATGGAA TGAACAATGG AGAAAAATAT CAGATAAATA 3600 CAATCTATTA TACCTACAAG CAAAAACTAC TTCTACTAAT CCTGGCCGTA CTGTTCTTGG 3660 TGATGACGAT CCCGACTATC AACAAATGGT AGATTTTTTG ACCCCAATAC ACAAAGCAAG 3720 TATTGCCGCA CGTGTTCTTG TTAAACGTGC TGCTGGTAGT CCCACTGAGA TCGCCGCCGC 3780 35 CGCCCCGATC ACCCCCTACA GTACTGCTGC CGGATATATA CACCAGGAAA TAGGATATGG 3840 GGGGTGCCAG GAACAACAC AATTTTGTGA AAAAAAACAT GGTGCAACAT CAACTAGTAC 3900 CACGAAAGAA AACAAAGAAT ACACCTTTAA ACAACCTCCG CCGGAGTATG CTACAGCGTG 3960 TGATTGCATA AATAGGTCGC AAACAGAGGA GCCGAAGAAA AAGGAAGAAA ATGTAGAGAG 4020 TGCCTGCAAA ATAGTGGAGA AAATACTTGA GGGTAAGAAT GGAAGGACTA CAGTAGGTGA 4080 40 ATGTAATCCA AAAGAGAGTT ATCCTGATTG GGATTGCAAA AACAATATTG ACATTAGTCA 4140 GAGTCAAACA GAAAATATAA AAACAGACGA TAATTTGAAA GATGCTTTTA TTAAAACTGC 4260 AGCAGCAGAA ACTTTTCTTT CATGGCAATA TTATAAGAGT AAGAATGATA GTGAAGCTAA 4320 45 AATATTAGAT AGAGGCCTTA TTCCATCCCA ATTTTTAAGA TCCATGATGT ACACGTTTGG 4380 AGATTATAGA GATATATGTT TGAACACAGA TATATCTAAA AAACAAAATG ATGTAGCTAA 4440 GGCAAAAGAT AAAATAGGTA AATTTTTCTC AAAAGATGGC AGCAAATCTC CTAGTGGCTT 4500 ATCACGCCAA GAATGGTGGA AAACAAATGG TCCAGAGATT TGGAAAGGAA TGTTATGTGC 4560 CTTAACAAAA TACGTCACAG ATACCGATAA CAAAAGAAAA ATCAAAAACG ACTACTCATA 4620 50 TCAATTTCTA CGTTGGATGA TCGAATGGGG AGAAGAGTTT TGTGCTGAAC GTCAGAAGAA 4740 GGAAAATATC ATAAAAGATG CATGTAATGA AATAAATTCT ACACAACAGT GTAATGATGC 4800 GAAACATCGT TGTAATCAAG CATGTAGAGC ATATCAAGAA TATGTTGAAA ATAAAAAAA 4860 AGAATTTTCG GGACAAACAA ATAACTTTGT TCTAAAGGCA AATGTTCAGC CCCAAGATCC 4920 55 AGAATATAAA GGATATGAAT ATAAAGACGG CGTACAACCG ATACAGGGGA ATGAGTATTT 4980 ACTGCAAAAA TGTGATAATA ATAAATGTTC TTGCATGGAT GGAAATGTAC TTTCCGTCTC 5040 TCCAAAAGAA AAACCTTTTG GAAAATATGC CCATAAATAT CCTGAGAAAT GTGATTGTTA 5100 TCAAGGAAAA CATGTACCTA GCATACCACC TCCCCCCCCA CCTGTACAAC CACAACCGGA 5160 AGCACCAACA GTAACAGTAG ACGTTTGCAG CATAGTAAAA ACACTATTTA AAGACACAAA 5220 60 CAATTTTTCC GACGCTTGTG GTCTAAAATA CGGCAAAACC GCACCATCCA GTTGGAAATG 5280 TATACCAAGT GACACAAAAA GTGGTGCTGG TGCCACCACC GGCAAAAGTG GTAGTGATAG 5340 TGGTAGTATT TGTATCCCAC CCAGGAGGCG ACGATTATAT GTGGGGAAAC TACAGGAGTG 5400 GGCTACCGCG CTCCCACAAG GTGAGGGCGC CGCGCCGTCC CACTCACGCG CCGACGACTT 5460 GCGCAATGCG TTCATCCAAT CTGCTGCAAT AGAGACTTTT TTCTTATGGG ATAGATATAA 5520

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AGAAGAGAAA AAACCACAGG GTGATGGGTC ACAACAAGCA CTATCACAAC TAACCAGTAC 5580
     ATACAGTGAT GACGAGGAGG ACCCCCCGA CAAACTGTTA CAAAATGGTA AGATACCCCC 5640
     CGATTTTTTG AGATTAATGT TCTATACATT AGGAGATTAT AGGGATATTT TAGTACACGG 5700
     TGGTAACACA AGTGACAGTG GTAACACAAA TGGTAGTAAC AACAACAATA TTGTGCTTGA-5760
     AGCGAGTGGT AACAAGGAGG ACATGCAAAA AATACAAGAG AAAATAGAAC AAATTCTCCC 5820
 5
     AAAAAATGGT GGCACACCTC TTGTCCCAAA ATCTAGTGCC CAAACACCTG ATAAATGGTG 5880
     GAATGAACAC GCCGAATCTA TCTGGAAAGG TATGATATGT GCATTGACAT ATACAGAAAA 5940
     GAACCCTGAC ACCAGTGCAA GAGGCGACGA AAACAAAATA GAAAAGGATG ATGAAGTGTA 6000
     CGAGAAATTT TTTGGCAGCA CAGCCGACAA ACATGGCACA GCCTCAACCC CAACCGGCAC 6060
     ATACAAAACC CAATACGACT ACGAAAAAGT CAAACTTGAG GATACAAGTG GTGCCAAAAC 6120
CCCCTCAGCC TCTAGTGATA CACCCCTTCT CTCCGATTTC GTGTTACGCC CCCCCTACTT 6180
10
     CCGTTACCTT GAAGAATGGG GTCAAAATTT TTGTAAAAAA AGAAAGCATA AATTGGCACA 6240
     AATAAAACAT GAGTGTAAAG TAGAAGAAAA TGGTGGTGGT AGTCGTCGTG GTGGTATAAC 6300
     AAGACAATAT AGTGGGGATG GCGAAGCGTG TAATGAGATG CTTCCAAAAA ACGATGGAAC 6360
     TGTTCCGGAT TTAGAAAAGC CGAGTTGTGC CAAACCTTGT AGTTCTTATA GAAAATGGAT 6420
15
     AGAAAGCAAG GGAAAAGAGT TTGAGAAACA AGAAAAGGCA TATGAACAAC AAAAAGACAA 6480
     ATGTGTAAAT GGAAGTAATA AGCATGATAA TGGATTTTGT GAAACACTAA CAACGTCCTC 6540
     TAAAGCTAAA GACTTTTTAA AAACGTTAGG ACCATGTAAA CCTAATAATG TAGAGGGTAA 6600
     AACAATTTTT GATGATGATA AAACCTTTAA ACATACAAAA GATTGTGATC CATGTCTTAA 6660
     ATTTAGTGTT AATTGTAAAA AAGATGAATG TGATAATTCT AAAGGAACCG ATTGCCGAAA 6720
20
     TAAAAATAGT ATTGATGCAA CAGATATTGA AAATGGAGTG GATTCTACTG TACTAGAAAT 6780
     GCGTGTCAGT GCTGATAGTA AAAGTGGATT TAATGGTGAT GGTTTAGAGA ATGCTTGTAG 6840
     AGGTGCTGGT ATCTTTGAAG GTATTAGAAA AGATGAATGG AAATGTCGTA ATGTATGTGG 6900
     TTATGTTGTA TGTAAACCGG AAAACGTTAA TGGGGAAGCA AAGGGAAAAC ACATTATACA 6960
     AATTAGAGCA CTGGTTAAAC GTTGGGTAGA ATATTTTTTT GAAGATTATA ATAAAATAAA 7020
25
     ACATAAAATT TCACATCGCA TAAAAAATGG TGAAATATCT CCATGTATAA AAAATTGTGT 7080
     AGAAAAATGG GTAGATCAGA AAAGAAAAGA ATGGAAGGAA ATTACTGAAC GTTTCAAAGA 7140
     TCAATATAAA AATGACAATT CAGATGATGA CAATGTGAGA AGTTTTTTGG AGACCTTGAT 7200
     ACCTCAAATT ACTGATGCAA ACGCTAAAAA TAAGGTTATA AAATTAAGTA AGTTCGGTAA 7260
     30
     TATAGATTGT ATGCTTAAAA AGCTTAAAGA TAAAATTGGC GAGTGCGAAA AGAAACACCA 7380
     TCAAACTAGT GATACCGAGT GTTCCGACAC ACCACAACCG CAAACCCTTG AAGACGAAAC 7440
     TTTGGATGAT GATATAGAAA CAGAGGAGGC GAAGAAGAAC ATGATGCCGA AAATTTGTGA 7500
     AAATGTGTTA AAAACAGCAC AACAAGAGGA TGAAGGCGGT TGTGTCCCAG CAGAAAATAG 7560
     TGAAGAACCG GCAGCAACAG ATAGTGGTAA GGAAACCCCC GAACAAACCC CCGTTCTCAA 7620
35
     ACCCGAAGAA GAAGCAGTAC CGGAACCACC ACCTCCACCC CCACAGGAAA AAGCCCCGGC 7680
    ACCAATACCC CAACCACAAC CACCAACCC CCCCACACAA CTCTTGGATA ATCCCCACGT 7740
    TCTAACCGCC CTGGTGACCT CCACCCTCGC CTGGAGCGTT GGCATCGGTT TTGCTACATT 7800
     CACTTATTTT TATCTAAAGG TAAATGGAAG TATATATATG GGGATGTGGA TGTATGTGGA 7860
    TGTATGTGAA TGTATGTGGA TGTATGTGGA TGTATGTTAT GGATATGTAT 7920
40
    ATATATAT GTGTATGTAT ATGATTTTCT GTATATGTAT TTGTGGGTTA AGGATATATA 8040
    AAAAGAAATA TAAAAACAAA TTTATTAAAA TGAAAAAAG AAAAATGAAA TATAAAAAAA 8160
45
    AATTTATTAA AATAAAAAA AAAAAAAAA AAAAGGAGAA AAATTTTTTA AAAAATAATA 8220
     (2) INFORMATION FOR SEQ ID NO:12:
          (i) SEQUENCE CHARACTERISTICS:
               (A) LENGTH: 2710 amino acids
50
               (B) TYPE: amino acid(C) STRANDEDNESS: single
```

- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
 - (iii) HYPOTHETICAL: NO

60

- (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: Plasmodium falciparum
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Asn Val Met Val Glu Leu Ala Lys Met Gly Pro Lys Glu Ala Ala Gly

	1				5					10					٦.	
	Gly	Asp	Asp	Ile 20		Asp	Glu	Ser	Ala 25	Lys	His	Met	Phe	Asp 30	_	Ile
5			, 3 5					40		Glu			45	Glu	Arg	_
		50					55			Ala		60				
10	65					70				Cys	75					80
10					85					Asn 90					9.5	
				100					10					11	0	_
15			115					120	0	Lys -			12	5		_
		130					135	5		Leu		14	0			
	145	TTE	Inr	Asn	Thr	150	Asn	Leu	Leu	Val	155	Val	Cys	Met	Ala	Ala 160
20					165	•				Asp	Tyr 0				17	Ala
				180					18					19	0	
25			195					200	0	Gly			20	5		_
		210					215	5		Gln		22	0			-
20	225					230				Leu	235					240
30					245	•				Leu 25	0				25	5
				260					. 26					27	0	_
35			275			•		280)	Arg			28	5		_
		290					295	,		Pro		30	0	_	_	
40	305					310				Trp	315					320
					325					Lys 330)				33	5
				340					34	Ser 5				35	0 _	-
45			355					360)	Leu -			36	5		_
		370					375			Tyr		38	0			
50	385					390				Lys	395					400
50					405					Gly 410)				41	5
				420					425					43	0	
55			435					440)	Arg			44	5 -		
	Glu	Lys 450	Leu	Ser	Asn	Glu	Glu 455	Ile	Cys	Thr	Lys	Val 46		Asp	Glu	Glu
	Gly 465	Gly	Thr	Ile	Asp	Phe 470	Lys	Asn	Val	Asn	Ser 475	Asp	Ser	Thr		Gly 480
60					485					Gly 490	Thr			_	Ser 49	Lys 5
				500					509	Val	Lys			51	Asn 0	Gly
	Gly	Ser	Ser	Asn	Glu	Trp	Glu	Glu	Lys	Asn	Asn	Gly	Lys			Ser

			515					52	0				52	5		
	Gly	Lys 530	Leu	Tyr	Glu	Pro	Lys 539	Pro	Asp	Lys	Glu	Gly 54	Thr	Thr	Ile	Thr
5	Ile 545	Leu	Lys	Ser	Gly	Lys 550	Gly	His	Asp	Asp	Ile 555	Glu	Glu	Lys	Leu	Asn 560
	Lys	Phe	Cys	Asp	Glu 565	Lys	Asn	Gly	Asp	Thr 57	Ile	Asn	Ser	Gly	Gly 57	Ser
	Gly	Thr	Gly	Gly 580	Ser	Gly	Gly	Gly	Asn 58	Ser	Gly	Arg	Gln	Glu	Leu	Tyr
10	Glu	Glu	Trp 595	Lys	Cys	Tyr	Lys	Gly 600	Glu		Val	Val	Lys 60		Gly	His
	Asp	Glu 610	Asp	Asp	Glu	Glu	Asp 615	Tyr		Asn	Val	Lys 62	Asn	Ala	Gly	Gly
15	Leu 625	Cys	Ile	Leu	Lys	Asn 630			Lys	Asn	Lys 635	Glu	Glu	Gly	Gly	Asn 640
	Thr	Ser	Glu	Lys	Glu 645	Pro	Asp	Glu	Ile	Gln 65	Lys	Thr	Phe	Asn	Pro 65	Phe
	Phe	Tyr	Tyr	Trp 660	Val	Ala	His	Met	Leu 66	Lys	Asp	Ser	Ile	His	Trp	Lys
. 20	· Lys	Lys	Leu 675	Gln	Arg	Cys	Leu	Gln 680	Asn	Gly	Asn	Arg	Ile 68	Lys	Cys	Gly
	Asn	Asn 690	Lys	Cys	Asn	Asn	Asp 695		Glu	Cys	Phe	Lys	Arg	Trp	Ile	Thr
25	705			Asp		710					715	His	Phe			720
				Gly	/25)				73	Thr				73	Pro
				Asp 740					74	5				75	Phe	Leu
30	Lys		755					760)				76	Asn 5	Ser	
•		770		Glu			775	5				78	0			
35	785			Asn		790					795					800
				Ile	805	•				81	0				ខា	Ala 5
				Leu 820					82	5				8.3	0	_
40			835					84()				84	5		
		850		Gly			855	;				86	0			
45	003			Tyr		8/0					875					880
				Arg	885)				89	0				89	5
50				Arg 900					90.	5				91	0	
50			915	Tyr				920)				92	5		-
		930		Asp			935	,				94	0		_	
55	945			Asn		950					955					960
				Pro	965					97	0				97	5
	Leu	Asp	val	Gly 980	Ser	Val	Thr	Lys	Asn 98		Lys	Ala	Ser	His 99		Leu
60	Leu	Gly	Asp	Val	Gln	Leu	Ala			Thr	Asp	Ala	Ala	Glu	Ile	Ile
		Arg	995 Tyr	Lys			Asn	100 Asn	00		•		10	05		
		101	υ				101	.5				10				

	Gln Ly 1025				103	()				コハコ	=				2040
	Asp Le			107	:				1 ()	Met	Trp				Lys
5	Ser Se		100	, ,				T O :	Ile 65	Thr			٦ ر	Asn	
	Lys G	10	' ⊃				10	Asp	Asn			10	Thr	Gly	
10		J 9 U				103) 5				11	Asp	Trp		
	Ala As 1105				111	כ				יווו	5				1120
15	Ile Il			112	:5				11	30				7.7	2 E
13	Arg Tr		114	. 0				114	45				11	50	
	Glu Ty	113	, ,				110	50				7 1	65		
20		L / U				117	/5				11	Lys	Cys		
•	Ala Cy 1185				1190	כ				1199	5				1200
ar.	Arg Ly			120	95				12	10				12	15
25	Thr Se		122	0				12:	25				12	Pro	Asp
	Tyr Gl	123	5				124	10				12	45		
30		250				125	> 5				12	60			
	Ala Al 1265				1270)				1275	5			_	1280
	His Gl			128	5				12	90				Phe	Cys
35	Glu Ly	s Lys	His	Gly 0	Ala	Thr	Ser	Thr	Ser	Thr	Thr		Glu 13	Asn	Lys
	Glu Ty	r Thr 131	Phe 5	Lys	Gln	Pro	Pro 132	Pro	Glu	Tyr	Ala	Thr 13	Ala	Cys	Asp
40		330				133	Glu 5	Glu			13	Lys	Glu		
·	Val Gl 1345	u Ser	Ala	Cys	Lys 1350	Ile	Val	Glu	Lys	Ile 1355	Leu	Glu	Gly		
•	Gly Ar	g Thr	Thr	Val 136	Gly		Cys	Asn	Pro	Lys	Glu	Ser	Tyr	Pro	1360 As p 75
45	Trp As	p Cys	Lys 138	Asn 0	Asn	Ile	Asp	Ile 138	Ser	His	Asp	Gly	Ala 13	Cys	Met
	Pro Pr	o Arg 139	Arg 5	Gln	Lys	Leu	Cys 140	Leu	Tyr	Tyr	Ile	Ala 14	His	Glu	Ser
50	~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~ ~												0.5	Dho	Ile
	Gin Tn	r Glu	Asn	Ile	Lys	Thr 141	Asp 5	Asp	Asn	Leu			Ala	Pne	
	Lys Th	:10			Glu	141 Thr	.5			rrp	14: Gln	2.0		Lys	Ser
	Lys Th	r Ala	Ala	Ala Glu	Glu 1430 Ala	141 Thr	5 Phe	Leu	Ser Asp	Trp 1435 Arg	14: Gln	20 Tyr	Tyr	Lys Pro	Ser 1440 Ser
55	Lys Th 1425	n Asp	Ala Ser Arg	Ala Glu 144 Ser	Glu 1430 Ala 5	141 Thr) Lys	5 Phe Ile	Leu Leu Thr	Ser Asp 14! Phe	Trp 1435 Arg 50	Gln Gly	20 Tyr Leu	Tyr Ile Arg	Lys Pro 14 Asp	Ser 1440 Ser
55	Lys Th 1425 Lys As	r Ala n Asp e Leu u Asn	Ala Ser Arg 146 Thr	Ala Glu 144 Ser 0	Glu 1430 Ala 5 Met	141 Thr Lys Met	Phe Ile Tyr Lys	Leu Leu Thr 146 Lys	Ser Asp 14! Phe	Trp 1435 Arg 50 Gly	Gln Gly Asp	Tyr Leu Tyr Val	Tyr Ile Arg 14 Ala	Lys Pro 14 Asp	Ser 1440 Ser 55 Ile
55 60	Lys Th 1425 Lys As Gln Ph Cys Le	r Ala n Asp e Leu u Asn 147 p Lys	Ala Ser Arg 146 Thr	Ala Glu 144 Ser O Asp	Glu 1430 Ala 5 Met Ile	141 Thr Lys Met Ser	Phe Ile Tyr Lys 148 Phe	Leu Leu Thr 146 Lys	Ser Asp 14! Phe 55 Gln	Trp 1435 Arg 50 Gly Asn	Gly Asp Asp Gly	20 Tyr Leu Tyr Val 14 Ser	Tyr Ile Arg 14 Ala	Lys Pro 14 Asp 70 Lys	Ser 1440 Ser 55 Ile Ala
	Lys Th 1425 Lys As Gln Ph Cys Le Lys As 14 Ser Gl	n Asp e Leu u Asn 147 p Lys	Ala Ser Arg 146 Thr 5 Ile	Ala Glu 144 Ser 0 Asp Gly	Glu 1430 Ala 5 Met Ile Lys Gln	141 Thr Lys Met Ser Phe 149 Glu	5 Phe Ile Tyr Lys 148 Phe 5	Leu Leu Thr 146 Lys 0 Ser	Ser Asp 14! Phe 55 Gln Lys	Trp 1435 Arg 50 Gly Asn Asp	Gly Asp Gly Asp Gly Asp Gly Asn	20 Tyr Leu Tyr Val 14 Ser	Tyr Ile Arg 14 Ala 85 Lys	Lys Pro 14 Asp 70 Lys Ser Glu	Ser 1440 Ser 55 Ile Ala Pro
	Lys Th 1425 Lys As Gln Ph Cys Le Lys As	n Asp e Leu u Asn 147 p Lys 90 y Leu	Ala Ser Arg 146 Thr 5 Ile Ser	Ala Glu 144 Ser O Asp Gly Arg	Glu 1430 Ala 5 Met Ile Lys Gln 1510 Cys	141 Thr Lys Met Ser Phe 149 Glu	5 Phe Ile Tyr Lys 148 Phe 5 Trp	Leu Leu Thr 146 Lys 30 Ser	Asp 14! Phe 55 Gln Lys	Trp 1435 Arg 50 Gly Asn Asp Thr 1515	Gly Asp Gly Asp Gly Asp Asp	20 Tyr Leu Tyr Val 14: Ser 00 Gly	Tyr Ile Arg 14 Ala 85 Lys Pro	Lys Pro 14 Asp 70 Lys Ser Glu	Ser 1440 Ser 55 Ile Ala Pro Ile 1520 Asp

	Asn	Lys	Arg	Lys	Ile	Lys	Asn	Asp	Tyr	Ser	Tyr	Asp	Lys	Val	Asn	Gln
																Gln
5				, ,				~								
										Glu						
	Gin 158	Lys 5	Lys	Glu	Asn	Ile 159	lle n	Lys	Asp	Ala	Cys	Asn	Glu	Ile	Asn	Ser
10	Thr	Gln	Gln	Cys	Asn	Asp	Ala	Lys	His	Arg	159 Cys	a Asn	Gln	Ala	Cvs	1600 Ara
10										16 Lys						
									1 h	<i>_</i>						
15								1 1	4()	Val						
15	Tyr	Lys 165	Gly 0	Tyr	Glu	Tyr	Lys 16	Asp	Gly	Val	Gln	Pro	Ile	Gln	Gly	Asn
	Glu	Tyr	Leu	Leu	Gln	Lys	Cys	Asp	Asn	Asn	Lys	Cys	60 Ser	Cys	Met	Asp
										Glu						
20					200	,					un					
				1 /0					1.7	Cys 05						
	Pro	Ser	Ile 171	Pro	Pro	Pro	Pro	Pro	Pro	Val	Gln	Pro	Gln	Pro	Glu	Ala
25	Pro	Thr	Val		Val	Asp	Val	17 Cys	20 Ser	Ile	Val-	Lys	17 Thr	'25 Leu	Phe	Lvs
										Gly						
		-				エ /コ	U				7'/ 6					
30										Ser						
	Gly	Ala	Thr	Thr	Gly 0	Lys	Ser	Gly	Ser	Asp	Ser	Gly	Ser	Ile	Cys	Ile
	Pro	Pro	Arg	Arg			Leu	Tyr	178 Val	Gly	Lys	Ĺeu	Gln	17 Glu	90 Trp	Ala
35				J				ואו) ()	Ala			70	$\Delta \Gamma$		
		101	U				T 2 7	LO				1 2	2 ハ			
	102	,				T83()			Ser	1835	:				1040
40	Phe	Leu	Trp	Asp	Arg 184	Tyr 5	Lys	Glu	Glu	Lys 189	Lys	Pro	Gln	Gly		Gly
•	Ser	Gln	Gln	Ala	Leu	Ser	Gln	Leu	Thr	Ser	Thr	Tyr	Ser	Asp	Asp	55 Glu
			Pro	Pro	Asp.	Lys	Leu	Leu	Gln	ob Asn				A	70	
45				_				124	411				70	0 -		
		~~~	_				T 0 2	70		Gly		79	ለለ .			
	vai 1905	His	GIY	GIY	Asn	Thr 1910	Ser	Asp	Ser	Gly	Asn 1915	Thr	Asn	Gly.		
50	Asn	Asn	Asn	Ile	Val	Leu	Glu	Ala	Ser	Gly	Asn	Lys	Glu	Asp	Met	1920 Gln
	Lys				エフム	_				797	የበ				10	3 E
				194	U				194	15				10	E 0	
EE			133	•				196	o O	Thr			7 9	65		
55	Glu	His 1970	Ala D	Glu	Ser	Ile	Trp 197	Lys 5	Gly	Met	Ile	Cys 198		Leu	Thr	Tyr
	Thr	Glu	Lys	Asn	Pro	Asp	Thr	Ser	Ala	Arg	Gly	Asp	Glu	Asn	Lys	Ile
	1985 Glu					TAAC	)				1995					2000
60					200	⋾				201	. 0				20	1 =
	Lys			2021	J				202	25				20.	2 0	
	Asp	Tyr	Glu 2039	Lys 5	Val	Lys	Leu	Glu 204	Asp	Thr	Ser	Gly		Lys	Thr	Pro
-								~ ~ 7	. •				204	± ⊃		

	Ser	Ala 205	Ser	Ser	Asp	Thr	Pro	Leu	Leu	Ser	Asp			Leu	Arg	Pro
	Pro 2069	Tyr		Arg	Tyr	Leu 207	205 Glu	Glu	Trp	Gly	Gln 2075	Asn	60 Phe	Cys		
5	Arg	Lys			208	Ala 5	Gln			20	Glu 90	Cys			Glu	0.5
				210	U				210	Ile 05	Thr			21	Ser	Gly
10			211	5				212	20		Lys		21	25		
		213	U				213	15			Pro	21	40			
15	214:	2				2150	)				Glu 2155	,			_	2160
15					216	5				21	Gly 70				21	75
				218	O				210	85	Ser			21	90	
20			219	5				220	Pro	Asn	Asn		22	Gly 05	Lys	
		221	0				221	.5			Thr	22	20	_	_	
	2223	>				223	)				Asp 2235	;				2240
25					224	:5				22	Ile 50				Asp	Ile 55
	Glu	Asn	Gly	Val 226	Asp 0	Ser	Thr	Val	Leu 22	Glu	Met	Arg	Val		Ala 70	Asp
30			227	5			•	228	Gly 30	Leu	Glu		22	Cys 85	Arg	_
		229	U				229	95			Glu	23	Lys	Cys		
	2305	5				2310	)				Asn 2315	;		_		2320
35					232	5				23	Leu 30			_	วิจ	Val
				234	0				234	45	Lys			23	Ser	His
40			235	5				236	0		Ile		23	65		
		2370	)				237	'5			Lys	23	80			_
45	200	,				233	,				Asp 2395					2400
45					240	5				24	Thr 10				24	15
				242	0				242	25	Asn			24	Cys	Ser
50	Ala	Ser	Ala 243	Asn 5	Glu	Gln	Asn	Lys 244	Asn 0	Gly	Glu	Tyr	Lys 24	Asp	Ala	Ile
		2450	כ				245	5			Ile	24	Glu 60	Cys		_
	Lys 2465	His	His	Gln	Thr	Ser. 2470	Asp	Thr	Glu		Ser 2475		Thr	Pro	Gln	Pro 2480
55	Gln	Thr	Leu	Glu	Asp 248	Glu 5	Thr	Leu	Asp		Asp		GĪu	Thr	_	Glu 95
	Ala	Lys	Lys	Asn 250	Met		Pro	Lys	11e 250	Cys	Glu	Asn	Val		Lys 10	Thr
60	Ala	Gln	Gln 251	Glu		Glu	Gly	Gly 252	Cys		Pro	Ala	Glu 25	Asn	Ser	Glu
	Glu	Pro 2530	Ala		Thr	Asp	Ser 253	Gly		Glu	Thr	Pro 25	Glu	Gln	Thr	Pro
	Val 2545	Leu		Pro		Glu 2550	Glu		Val		Glu 2555	Pro	Pro	Pro	Pro	Pro 2560

	Pro Gln Glu Lys Ala Pro Ala Pro Ile Pro Gln Pro Gln Pro Pro Thr 2565 2570 2575
	Pro Pro Thr Gln Leu Leu Asp Asn Pro His Val Leu Thr Ala Leu Val 2580 2585 2590 •
5	Thr Ser Thr Leu Ala Trp Ser Val Gly Ile Gly Phe Ala Thr Phe Thr
	Tyr Phe Tyr Leu Lys Val Asn Gly Ser Ile Tyr Met Gly Met Trp Met 2610 2620
10	Tyr Val Asp Val Cys Glu Cys Met Trp Met Tyr Val Asp Val Cys Gly 2625 2630 2635 2640
	Cys Val Leu Trp Ile Cys Ile Cys Asp Tyr Val Trp Ile Tyr Ile Tyr 2645 2650 2655
15	Ile Tyr Ile Cys Leu Cys Ile Cys Val Phe Gly Tyr Ile Tyr Val Tyr  2660 2665 2670
15	Val Tyr Asp Phe Leu Tyr Met Tyr Leu Trp Val Lys Asp Ile Tyr Ile 2675 2680 2685 Trp Met Tyr Leu Tyr Val Phe Tyr Ile
	Trp Met Tyr Leu Tyr Val Phe Tyr Ile Tyr Ile Leu Tyr Ile Cys Ile 2690 2695 2700 Tyr Ile Lys Lys Glu Ile
20	2705 2710
	(2) INFORMATION FOR SEQ ID NO:13:
25	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 19124 base pairs
	(B) TYPE: nucleic acid (C) STRANDEDNESS: single
30	(D) TOPOLOGY: linear
30	(ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO (iv) ANTI-SENSE: NO
35	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:
	ACATTTTTC GTAATATATA TATATATAT TATATATAT TCTCTTTTTC TAATATATAT
	ATCCTTCTAT TTTCGATTTT TTCATTTTTT TCCAGTATTA ATTTATTTAT TTATTTGTGA 120 TATTTTATAA TATATTATTT AAATGTGTAT TTATATATGT GTTTTATTTT TGTTATTAAT 180
<b>40</b>	TTGAATAATC CGAGCGAAAA AAAATATATA ATCTCATATA AAAATTATTT ATAATACAAT 240 ATTATATAGT TTCCTATTAA AATAAATTAA TATAATATAC AATAATATTT CTTGTTATTT 300
	TTATAAATAT AACTAATTTC TTATTTTAT TTAACTTTAT TCCTTTTTAA TTTCTTAATT 360
45 ·	TTATAAATAT AACTAATTTC TTATTTTTAT TTAACTTTAT TCCTTTTTAA TTTCTTAATT 360 CTTTTATGCA AACAAAAAC ATAAAGTAAT TCTACATATC AACAAAAAA AAAAAAAAA 420 AAAAAAAAAA ATTTATTATA ATATAATAAA AAATATAAAG ACATACGTTC ACTTATTATT 480 ATAAATGATT TATTACGATT AAAACATATT GAGATTATAA TAATATAATT TAACATAGAA 540
45 -	TTATAAATAT AACTAATTTC TTATTTTAT TTAACTTTAT TCCTTTTTAA TTTCTTAATT 360 CTTTTATGCA AACAAAAAAC ATAAAGTAAT TCTACATATC AACAAAAAAA AAAAAAAAA 420 AAAAAAAAAA ATTTATTATA ATATAATAAA AAATATAAAG ACATACGTTC ACTTATTATT 480 ATAAATGATT TATTACGATT AAAACATATT GAGATTATAA TAATATAATT TAACATAGAA 540 AGAGTTAAGA ATACATTTTT TTTTTTTTTT TGATATGTAA TTCAACATAT ATATATATAT 600 ATATCTTTTT AATTTAATTA AATAAAATTC CTTATTATTC ATATTGTTTC TTTTATCACA 660
	TTATAAATAT AACTAATTTC TTATTTTAT TTAACTTTAT TCCTTTTTAA TTTCTTAATT 3600 CTTTTATGCA AACAAAAAAC ATAAAGTAAT TCTACATATC AACAAAAAAA AAAAAAAAAA
45 ·	TTATAAATAT AACTAATTTC TTATTTTAT TTAACTTTAT TCCTTTTTAA TTTCTTAATT 3600 CTTTTATGCA AACAAAAAAC ATAAAGTAAT TCTACATATC AACAAAAAAA AAAAAAAAAA
	TTATAAATAT AACTAATTTC TTATTTTAT TTAACTTTAT TCCTTTTTAA TTTCTTAATT 3600 CTTTTATGCA AACAAAAAAC ATAAAGTAAT TCTACATATC AACAAAAAAA AAAAAAAAAA
50	TTATAAATAT AACTAATTTC TTATTTTAT TTAACTTTAT TCCTTTTTAA TTTCTTAATT 3600 CTTTTATGCA AACAAAAAAC ATAAAGTAAT TCTACATATC AACAAAAAAA AAAAAAAAAA
	TTATAAATAT AACTAATTTC TTATTTTAT TTAACTTTAT TCCTTTTTAA TTTCTTAATT 3600 CTTTTATGCA AACAAAAAAC ATAAAGTAAT TCTACATATC AACAAAAAAA AAAAAAAAAA
50	TTATAAATAT AACTAATTTC TTATTTTAT TTAACTTTAT TCCTTTTTAA TTTCTTAATT 3600 CTTTTATGCA AACAAAAAAC ATAAAGTAAT TCTACATATC AACAAAAAAA AAAAAAAAAA
50 55	TTATAAATAT AACTAATTTC TTATTTTAT TTAACTTTAT TCCTTTTTAA TTTCTTAATT 3600 CTTTTATGCA AACAAAAAAC ATAAAGTAAT TCTACATATC AACAAAAAAA AAAAAAAAAA
50	TTATAAATAT AACTAATTTC TTATTTTAT TTAACTTTAT TCCTTTTTAA TTTCTTAATT 3600 CTTTTATGCA AACAAAAAAC ATAAAGTAAT TCTACATATC AACAAAAAAA AAAAAAAAAA
50 55	TTATAAATAT AACTAATTC TTATTTTAT TTAACTTTAT TCCTTTTTAA TTTCTTAATT  CTTTTATGCA AACAAAAAAC ATAAAGTAAT TCTACATATC AACAAAAAA AAAAAAAAA 420 AAAAAAAAAA ATTTATTATA ATATAAATAA AAATATAAAG ACATACGTTC ACTTATTATT  480 ATAAATGATT TATTACGATT AAAACATATT GAGATTATAA TAATATAATT TAACATAGAA 540 AGAGTTAAGA ATACATTTTT TTTTTTTTTT TGATATGTAA TTCAACATAT ATATATATAT 600 ATATCTTTTT AATTTAAATA AATAAAAATC CTTATTATC ATATTGTTC TTTTATCACA 600 ATATCTTTTT AAAAAAATAAT TTTCGATTTT ATCGATATAT TTATGCGTT TATATACTTA 720 TGTGAAATAT TAAAAAAATAT TTTCGATTTT ATCGATATAT TAATGTCGTT TATATACTTA 720 ATTTATAAAT TCATTTATAT ATTTCAAATA TATTTCGATG GTTTATTTTC AAATACAATT 840 AATTAGATTT CTTAAATATT TCTTCATTTA TTCATTTTTA TAGCATATAC ATGCACATTA 900 TAAAATTATTA ATAAAAAAATT TTTATTTTAA TATATAATA
50 55	TTATAAATAT AACTAATTTC TTATTTTAT TTAACTTAT TCCTTTTTAA TTTCTTAATT  CTTTTATGCA AACAAAAAAC ATAAAGTAAT TCTACATATC AACAAAAAAA AAAAAAAAAA
50 55	TTATAAATAT AACTAATTC TTATTTTAT TTAACTTTAT TCCTTTTTAA TTTCTTAATT  CTTTTATGCA AACAAAAAAC ATAAAGTAAT TCTACATATC AACAAAAAA AAAAAAAAA 420 AAAAAAAAAA ATTTATTATA ATATAAATAA AAATATAAAG ACATACGTTC ACTTATTATT  480 ATAAATGATT TATTACGATT AAAACATATT GAGATTATAA TAATATAATT TAACATAGAA 540 AGAGTTAAGA ATACATTTTT TTTTTTTTTT TGATATGTAA TTCAACATAT ATATATATAT 600 ATATCTTTTT AATTTAAATA AATAAAAATC CTTATTATC ATATTGTTC TTTTATCACA 600 ATATCTTTTT AAAAAAATAAT TTTCGATTTT ATCGATATAT TTATGCGTT TATATACTTA 720 TGTGAAATAT TAAAAAAATAT TTTCGATTTT ATCGATATAT TAATGTCGTT TATATACTTA 720 ATTTATAAAT TCATTTATAT ATTTCAAATA TATTTCGATG GTTTATTTTC AAATACAATT 840 AATTAGATTT CTTAAATATT TCTTCATTTA TTCATTTTTA TAGCATATAC ATGCACATTA 900 TAAAATTATTA ATAAAAAAATT TTTATTTTAA TATATAATA

TATATACATG CAATGATATG TTTCTGTTGG AATATGTATT ATATACTTAT ATGTTCTAAT 1740 AAATGTATTG TACACCTTTA GCAACTATTA CTACACACAT TTTTATATAA TTTATAACAG 1800 GAAAATATGT TATATTATTA CAATATCTTA ATGTGTTTTT GCAAAAATAT AAAAAACAAG 1860 AAAATTACAA TTGTAATTAA TCGTATGACA TAAAATTATA TTATATTAGA AATTAAAATT 1920 CAAAATTATA AAAAATATGG AAATGTTTTG TTATATTATT TTTTTAAAAA TTTAATTATT 1980 5 TTATTTTATT ATTTATTTTT TTTTTTTTT GTGTTCTAAA TAAAAAGGCA AATATGATTC 2040 AAGTAAAAA TATATATT TACATAATGG CAAAATAATT GTTTATTATA TTATATGACT 2100 ATAATAATAT TTTAGATTAA ACATATGTAA TTCATTTAAC AGAATAAAAT AAAATATTAT 2160 ATATATAT TAATTATTAA GTTATAGATT TAATAAAAAT ATATTATACA TATGAGATTA 2220 10 AAAATGAAAG TTCACTACAG TAATATATTA TTATATGTCG TCAATTTAAG TATATTCTTA 2280 ATATCACGTA TGCACTAAAT AATGACAATA ATAATATATA TGTAACATTT TATAATTGAT 2340 GTAAATAAAA AAATATACAT ATATACAAAA ACATATATGA TATTTACATT CTTTTTTATA 2400 GATAAATATC CAGAAGAACT ATTACATCAC TTCACTTCAT ATACCAAACA CGAAAAAAAT 2460 ACAACCACTA GGTTATTATG CGAATGTGAC TTATATACGT CCATTTATGA TAATGACCCG 2520 GAAATGATAT TAGTGATGGA AAATTTCAAT AAACAGACAG AAGAAAGGTT TCATGAATAC 2580 15 AATGAACGCA TGCAAGAAAA ACGAAAAATA TGTAAAGAAC AATGCGAAAA GGATATACAA 2640 AAAATTATTT TAAAAGATAA AATCGAAAAG GAATTAACAG AAAAGTTAGA GGCATTGGAA 2700 ACGAATATAA AGACTGAGGA TATACCTACT TGTGTATGCG AAAAATCAGT AGCAGATAAA 2760 GTGGAAAAA CGTGTTTGAA ATGTGGAGGT ATATTGGGTG TTGGTGTGAC TCCATCTTTA 2820 20 GGTTTATTAG GAGAAATAGG TGGACTTGTT ATAAATAATT GGACAAATAC TCCTTTTTAT 2880 AAAGCTTTTC TTACTTTTGC TCAAAAGGAA GGTATAGCTG CCGGTAAAAT TGCTAGTGAT 2940 ACTGCTCGTA TTGATACAGT TATTTAAGGA ATAATATCAA ATTTTGATGT GCACACTATA 3000 AATGGTTCTA CGTTGGGGAA AGTTATTACC GTAGAAGCTC TTAAGGATGA CACTACTCTT 3060 ACTACGGCAC TATATAATGA ATATGTAAGC ATGTGTGTAA ATACGAACCC TGTCGAAGAC 3120 25 AAATTAATTT GTGCTTTTGG GATGAGAGAC GGTCTAGTTG CAGGGCAATA TGCTTCATCG 3180 CGAGACGTTA TAGGATCAAG TGTAAAAGGA ATTATTAGAA AAGCTGCAAA CGCTGCTTCA 3240 CAAGCTGCTG AGACAGCTGC TAACGAAACT ACTTCCGGAA TGATCGAAGC CGAGTTAAGT 3300 AAAATAACAT CTGCAGGTGC TAATTTACAC AGTGCAATTA CTTACTCAGT AACTGCGATA 3360 TTGGTTATAG TTTTGGTTAT GGTAATTATT TATTTAATAT TACGTTATCG TAGAAAAAAA 3420 AAAATGAAGA AAAAATTGCA ATATATAAAA TTATTAAAGG AATAGATATA CGATGTCGAG 3480 CTATTAGCGG TAATTTAAAG TATTGTGAAT TTTTCATTTA ATATGCTATG ATCATTTGAT 3540 AATTAATTTT TTTTTATAAT ATTATATTTT TTTATACCTT GGATTCTTAC ATTGTTTTAT 3600 30 TATTATATGA TTATTTAATT ATTATACTTA TATATATATA TATTTTTACA TTAAGATATT 3660 35 TTATTATTAT TAGATGCATA TTAGTGATGA TTATAATAAT AACCTATTGA AGAGAATAGA 3780 ACATAATAAT ATATTAAATT AATAGAACTT CATTTTTATT GTTATATGTA TATAAAAATA 3840 AGAAATTTGA AAAAGTAATT TACACATGAT AATGTATTTT ATTTTATTTG TGTTGTTTTA 3900 TATTTATTTA TAAAAATTGT TTAATATAAG TTGTTATTAT AATTTTTTAA TATGGCACCA 3960 TTAGCTTTCC ATTATACAAA TATATATTTC CTCATTAGAA TCTGAATATT TATTGTATTA 4020 40 TAAAAAAGT ATAATATAAT AAAATATCTA AGATTTTTTC TAATTTGTTT AATTTATAAT 4080 AAATTTTAAT TTTATACGAT AGAATAAATT ATAATCAACA TATATATATG TATTCATCTT 4140 AAGAACCTAT TACAATATAG TAACAACTGG TTCCTTTTTA TTATAAATAA CATAAGAATG 4200 TGTAAAAGGA TAGTTGTTAA AGGCTTTTTT AATATTGATT ATAAATGTTT GTAAGATATA 4260 TATAATAGAT ATCTTAACAT ACAACTTTGC ATAATTGTAA TTAAAAAAAT ATATAATA 4320 AGAAATATTA TAAAAAAAT TAAGCATAAA TGTCACAATA AATTTTTTT 4380 TATTAATTTA ATTTTTTTT ATTGTTCTAA AATATATTGA TTATGAGAAT ATTATTTGTG 4440 TCTAATATAA TTAAGATATT TCTAATATTA ATTTATATAT ATATATTTAA AAGTATTTTA 4500 AGAATAATTT TTTACTTATT TATTATAATA TGAAATATGC ATGGAGTATA TATAAATATT 4560 GATGACAAAA AAAAAACTTT TAAAATGGAA AATATGCATA TAATAAAATA CTATATAGTA 4620 50 TAATTGGTGA AATAGTTGTA ACTTATACAA ACATGTTGCA TTCATAATTT AGAGATTATG 4680 TAATATTGTT TATGTATCGT AATATATAT AATATAATTG TTTTTTTAGT ATGTATGGTA 4740 TTCTAATAAT ATATTCATAT GTAGTCATAG TGTCAATGAA TATAAAATAT GGTATATTTA 4800 TATTATTGTA TATATTAAAT AAGTAACACA GAACATTATA TATAGTAATA AATAGAAGAA 4860 ATAATATAT TTTATGTTAT ATATTATTAG TTATTATAAA CGGGAAAATT CATAATATTT 4920 ATGGAAAGCA TAAAAAATGT TACTGTAATA GGATAAAATA TATTATATAA AATGTTTATT 5040 TTATCTTAAA AAGGTTCCTA TTATAACATT AAAAAAAATT TGTCCCATTT TATAAATAAT 5100 TAACTACATT TACATAATGA AATTTCGATT TTGTGTTTTT TTGATGAATA TTATGGACTA 5160 ATTATTATA TGTGAATGCG TTCTATATAA TAATAATAAT TTTATTTAAA AAAATGAAAA 5220 60 ATAAGAAATA AATATCCTGA TTTTGTAGTT CCAATAGCTT AATATAATTA TGGACTCATA 5280 TATATATTAT ATATATCTTT ACAACAAGTA ATAAGTAAAT ATTATTTAA TCTTAATAAG 5340 GAAAATAAAA ATAATAAAAT AAGAATACTG AATAATAAGT CATATTATAC ATTTTTTAAA 5400 AATGTAACAT AATTACAAAT ACGTAACATG TATTATAGAA ATAATAAGAA TTTAATATTA 5460 

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	ATATTAACAT	GATTAGTTTT	TTGAAAAATA	TTTAAATATC	ATATAATAAT	AATAAATTAG	5580
	TTAAAATAAT	AGTATTTCAT	ACAAAATACT	AACTTATAAG	ΤΔΤΔΤΓΔΤΛΤ	እ እ ጥ እ ጥጥ እ ጥ እ ጥ	5640
	WINIWINI	TIMIGIGITI	TIGATIGGGT	GTATATAAGG	ርጥልጥል ልርጥአጥ	እጥእጥ <b>ር ርርጥጥር</b>	F700
	IICAITATAT	ATTTATATGT	GAATAGATAC	ATATAAGTTA	<b>ስጥልጥልጥጥ</b> ላጥ ለ	TTCTCTTATATA	
5	GTCTGTGTTA	AGATAGATAT	GCATTACAGT	TAAGGGTTAT	<b>አርጥጥጥጥጥጥ</b>	TTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTT	5760
	GTACATATAT	ATAAAAAATA	GATAACTAAC	AATATCCATA	TTACAACAAC		5820
	ТААААТАТАТ	ATATATAT	אראדאדנדאר	ACATTRICCALA	TIACAAGAAT	AATATTTGTA	5880
	ערער ערער ערערער ערערער ערערער		TATALAMA NA	ACATTAAAAC	TATACTAATA	GGTAATTAGT	5940
		CATCCTTTTA	ITATIATAAT	TTTTTTTGTT	TTACTTCTTG	TCGTTCTTTT	6000
10	TIGITATIAL	AATATAACAA	ATATAAAACA	ATATCAGTAT	TTGGAATATA	AATAAATTTA	6060
10	TICIACATAT	ATGCATATAT	ATATATAT	ATATATATAT	<b>ስጥልጥልጥልጥልጥ</b>	ስጥለጥ <b>ለጥለጥ</b> ለጥ	C122
	AIAIGIAIGA	TTTTATACTA	TTTTTATACA	TGCATTTTTA	$T\Delta T\Delta TTTTTT\Delta$	ጥ እጥ እጥ እ ረጥጥጥ	6100
	AAAGATATTA	TTAATATTTA	TATAGTAGCA	TATATGTATT	ΤΔΤΔͲΤΔͲλλ	CV V V T V T T T T T T T T T T T T T T T	C240
	CATTATATA	AATATATAGA	ACATGAACAT	TTTATTAATA	<b>Δርፕር</b> ΔͲΔͲͲͲ	$C \lambda \lambda T \lambda T \lambda T \lambda T \lambda T \lambda T$	6300
	ATTTATAATG	TGTATTTTTA	CTTATTTTTT	TATATTATAC	ΔΔΥΔΔΔΛΥΥΥ	TCNNNTTCNT	6300
15	AAAATGCATG	AAATACATAA	AAAAATACAA	СРУРОСУРУА	CATAAAAAIII	TUTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTT	6360
	TATAATATAA	TATAATATAA	ΤΔΔΤΔΤΔΤΤΤ	TTCCTCTT	TT ATAMAMCA	TTTTTTATTAA	6420
	GATGCTATAT	<b>ΔΤΔΤΤΛΤΤΛΤ</b>	חיית א איי א איי א	ATAATATATA	ITATTTATCA	TTTTTTTT	6480
	TAATATACTA	ATATTATTAT	VIUWIUWII	MIAAIAIAIA	ACAACAAAAA	TTAATAATAA	6540
	TATIAIACIA	CTTTTAATAT	AATACAACAA	TACAAAGAAT	ATGTATCTAT	ATCAATTATA	6600
20	CECECEEEE	TATATAAATA	TGATAGATAA	TATAGATAGA	GAGAAACGAA	GAACATATTT	6660
20	GICICITITG	TTATCTCTAA	TATATATATA	TATATAATAA	ATTAAAATAA	AGTCAAAAAA	6720
	AATATACATA	TATTAATGTT	AATAATTAAA	TATATAAACA	CGTTGCATAT	$V \oplus V \oplus U \oplus U \oplus U \oplus U$	C700
	ATATGTTTGT	ATTTTCGTAT	TTTTTTTC	TCATTTATAA	מ מדידים מידידידים	TAAATAAAA	6040
	AIAAAAAAA	TAATATAT	ATAATTAAAT	AGATAAATAA	AGGAATACAT	<b>העעבעעעעעעעע</b>	6000
	ATTTCTGATT	ATATTTTTTT	TTTGTTAGAA	TATTTAAATT	TATTATAAAT	ጥጥልጥጥልልጥእጥ	6060
25	ATATATATAT	TTTTTTTAAA	AATATATAAA	ACTAATAATT	ΔΤΤΔΤΤΔΤΤΔΤ	1 TAT 1 TAT 1 A 1	7020
	TATTATTTT	TTAACATATA	CATATATTGT	ΔΑΤΑΤΤΑΤΑΛ	TACTACAACT	ACAIAIIAAA	7020
	ATATATATAT	ATATACAATA	ΤΤΤΑΤΑΤΑΤΑ	TTCTN NTNCN	TAGIACAACI	ATTAATATAT	7080
	ТАТАТАСАТТ	CACAAAAGTG	עריי עריי עריי עריי עריי עריי עריי עריי	TATTCTACA	TAAATTATAC	CTTACATATA	7140
	ΑΤΑΓΑΤΑΤΑΤ	ACATACCCCC	ACCURACCURAC	CARACTACCA	TATTATAATA	CTACTGTAAT	7200
30	TATCTATCCC	ACATACCCCC	CCACCOTAC	GAAACACCAC	CAAACCATGT	ATCACGTATG	7260
00	AAAAATCCCC	ACGATATAAA	CCACGTACCA	CGTATGACAT	AATGTAATGG	TGGAGTTAGC	7320
	TAMMAN I GGGG	CCCAAGGAGG	CTGCAGGTGG	GGATGATATT	GAGGATGAAA	GTGCCAAACA	7380
	TAIGITIGAT	AGGATAGGAA	AAGATGTGTA	CGATAAAGTA	AAAGAGGAAG	CTAAAGAACG	7440
	TGGTAAAGGC	TTGCAAGGAC	GTTTGTCAGA	AGCAAAATTT	GAGAAAATG	AAACCCATCC	7500
0.5	ACAAACACCA	GAAGATCCAT	GCGATCTTGA	TCATAAATAT	CATACAAATG	<u>ምልልርጥልርጥል</u> ል	7560
35	IGIAATTAAT	CCGTGCGCTG	ATAGATCTGA	CGTGCGTTTT	TCCGATGAAT	ATCCACCTCA	7620
	ATGTACACAT	AATAGAATAA	AAGATAGTCA	ACAGGGTGAT	AATAAAGGTG	CATCTCCTCC	7600
	ATATAGGCGA	TTGCATGTAT	GCGATCAAAA	TTTAGAACAG	ATAGAGCCTA	<b>ፕ</b> ለአአአካጥአአር	7740
	AAATACTCAT	AATTTATTGG	TAGATGTGTG	TATGGCAGCA	ΔΔΔΥΥΥΘΑΛΟ	CACAAMCAAM	7740
	AACACAAGAT	TATCCAAAAT	ATCAAGCAAC	ATATGGTGAT	TORAG	A A TATIONA O	7800
40	TATGCTGGCA	CGAAGTTTTG	CCCACATACC	CCACATTCTC	ACACCARACIC	AAATATGTAC	7860
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	ATTEMANTIA	CGAGAAGATT	GGTGGACTGC	TAATCGAGAA	ACAGTATGGA	AAGCCATCAC	8100
A.E.	ATGTAACGCT	TGGGGTAATA	CATATTTTCA	TGCAACGTGC	AATAGAGGAG	AACGAACTAA	8160
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	ACACGATGAG	GATGACGAGG	AGGATTATGA	AAATGTAAAA	AATGCAGGCG	Carragiles	0100
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	TGAAATCCAA	AAGACATTCA	Δη/ Δη/ Δη/ Δη/ Δη/ Δη/ Δη/ Δη/	TAMADD 1004441	CTTCCACATA	TOTAL TEACH	9240
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		- ooi maaaaa	CIICAGAG	VIGITIACAY	MAIGGIAACA	GAATAAAATG	9360

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- (2) INFORMATION FOR SEQ ID NO:14:
- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 3060 amino acids

- (B) TYPE: amino acid(C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

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50				Asp 20					25		-			3.0	Gly	-
			35	Asp				40					45	Gly	_	-
5̄5		50		Arg			55					60				-
	65			Pro		70					75			_		RΛ
	Asn	Val	Thr	Thr	Asn 85	Val	Ile	Asn	Pro	Cys 90	Ala	Asp	Arg	Ser	Asp	Val
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65	Leu	His 130	Val	Cys	Asp	Gln	Asn 135	Leu	Glu	Gln	Ile	Glu 140		Ile	Lys	Ile

	Thr	Δen	Thr	Wie	Nan	T 011	7	17m 1	<b>&gt;</b>		_					
	743					Leu 150					155					
_					T02					- 170	Lys				376	• -
5				700		Gln			185	Met	Leu			100	Phe	Ala
			TSS			Val		200					つんに	Gly	Asn	
10		210				Arg	215					220	Leu	Lys		
	223					230					235	Glu	Ala			Gly 240
15					245	Phe				250					255	Asn
15				260		Lys			265					270		
			2/5			Cys		280					285			_
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	505					Glu 310					315					220
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				340		Tyr Gly			345				•	350		_
			355			Asn		360					365			
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	202					390 Ser					395					400
35					400	Asn				410					<i>4</i> 7 E	
			Asn	420		Glu			425					430		_
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	#02				Glu	470 Ser					475					400
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					645	Glu				650	Phe				655	Tyr
65	Tyr	Trp	Val	Ala	His	Met	Leu	Lys	Asp	Ser	Ile	His	Trp	Lys	Lys	Lys

					660					C C F							
-				0/3	Cys	Leu		Asn	680		Arg			625		Asn	
5			090					Glu 695	Cys	Phe			700	Ile	Thr		
		/05					710	Ile				715	Lys	Thr			720
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10					740			Tyr		745					750	Lys	Gly
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15			//0					Lys 775					780				
		785					790	Ser				795					900
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20					820			Asp		825					830		
				835				Glu	840					845			
25	•		850					Lys 855 Lys					860				
		000					870	Gly				875					000
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			•		900			Arg		905				-	910		
	•			312				Gly	920					925			_
35			930					935 Lys					940				-
		945					950	Met				955			-	-	960
40	,					965		Asn			970					975	_
				Gln	980			Lys		985					990		_
45			Lys	Asp				Ile	1000 Gln	)				100	5		_
40		Asp	Gln	י			Cys	1015 Arg	5			Tyr	1020 Ser	2			_
		1025 Gly		Ile	Ile	Arg	1030 Gly	Arg.	Asp	Met	Trp	1035 Asp	Glu	Asp	Lys	Ser	1040 Ser
50		Thr	Asp	Met	Glu 1060	1045 Thr		Leu	Ile			Phe	Lys	Asn		-	5 Glu
		Lys	His	Asp	Gly		Lys	Asp	Asn 1080	1069 Pro		Tyr	Thr			) Glu	Ser
55		Lys	Lys	Pro		Tyr	Lys	Lys	Leu		Ala	Asp			Glu	Ala	Asn
		Arg 1105	His		Val	Trp	Arg	Ala		Lys	Cys	Ala 1115		Lys	Gly	Ile	
				Gly	Met	Pro 1125	Val	Asp	Asp	Tyr	Ile 1130	Pro	Gln	Arg	Leu		
60		Met	Thr	Glu	Trp	Ala		Trp	Tyr	Cys 1145	Lys		Gln	Ser	Gln 1150		Tyr
		Asp	Lys	Leu 1155	Lys		Ile	Cys	Ala 1160	Asp	Cys	Met	Ser	Lys 1165	Gly	Asp	Gly
65		Lys	Cys 1170	Thr		Gly	Asp	Val 1175	Asp		Gly	Lys	Cys 1180	Lys	Ala	Ala	Cys

	Asp 1			•		1190	)				1195	5				1200
	Ile	Ser	Asp	Lys	Tyr 120	Asn	Leu	Leu	Tyr	Leu 121	Gln	Ala	Lys	Thr	Thr 121	Ser
5	Thr I			1221	,				122	Asp 5	Asp			123	Tyr	Gln
	Gln 1		1235	>				124	0				124	Ile 5	Ala	
10		T520	)				125	5				126	n			
	Ala 2 1265					1270	)				1275	;				1290
45	Glu :				128	•				1290	0				129	ξ
15	Lys I			1300	)				130	5				121	Λ	
	Thr 1		TRI	•				1320	0				132	5		
20		1330	)				1339	5				134	0			
	Ser 1					1350	)				1355	5				1360
20	Thr :				1365	•				1370	n .				127	ξ
25	Cys I			<b>138</b> (	)				1389	5				139	n	
•	Arg A		1395	•				140	0				140	5		
30		1410	)				1419	5				142	0		_	
	Ala 1 1425					1430	)				1435	i .			•	1440
0.5	Asp S				144	)				1450	)				145	ξ
35	Leu A			1460	)				1469	5				147	n	
	Asn 1		1475	•				1480	)				148	ς		
40	_	1490	,				1495	>				150	0			
•	Leu S 1505					1510	)				1515	,				1520
45	Gly N				1525	5				1530	)				1530	ξ -
45	Arg I			1540	)				1549	5				155	n	
	Asn (		1555	5				1560	)				1569	5		
50		1570	)	_			1575	5				1580	)	_		-
	Lys (					1590	)				1595	,				1600
Fr	Gln (				1605	5				1610	)			_	1619	5 -
55	Gln (			1620	)				1625	5				1636	0	
	Asn I		1635	,				1640	)				1649	5	_	-
60		1650	)				1655	5				1660	)			-
	Leu I 1665					1670	)				1675				=	1680
	Val I				1685	5				1690	)				1699	His
65	Lys 1	ſyr	Pro	Glu	Lys	Cys	Asp	Cys	Tyr			Lys	His	Val	Pro	Ser

				170					170	5				171	0	
			Pro 171	5				172	Gln 0	Pro			172	Ala	Pro	
5		1/3					173	5				174	Phe	Lys		
	1/4	<b>-</b>	Phe			175	U				175	Gly	Lys			1760
10			Trp		1/6	5				177	0				177	Ala
10			Gly	178	U				178	5				170	Λ	
			Arg 179	>				180	0				180	5		
15		TRT					181	5				182	0			_
	T87	5	Asn			183	0				183	5				1940
20			Arg		184	>				185	0				1 2 5	5
20			Leu	T86	כ				186	5				187	Λ	=
			Asp 1879	>				188	0				188	5		
25		TRA					189	5				190	0			
	TAO	5	Asn			191	0				1915	5				Asn 1920 Ile
30			Lys		1929	õ				193	0				102	<b>.</b>
			Lys	194(	J				194	5				195	Λ	
			1959 Ser	>			,	1960	0			•	196	5		
35		19/	0 Pro				197.	5				1986	n			
	190	5	Glu			1990	)				1999	5	-			2000
40			Ala		2005	5				201	0			•	201	5
			Val	2020	)				202	5				203	n	
45			2035 Asp	•				2040	)				204	5		
45	Phe	Arg	0 Tyr				205	5				2060	כ			-
	206	<b>&gt;</b>	Leu		Gln	Ile	)				2075	;		•		2080
50	Gly	Gly	Ser	Arg	2085 Arg		Gly	Ile	Thr	2090 Arg	Gln Gln	Tyr	Ser	Gly	2099 Asp	Gly
	Glu	Ala	Cys	2100 Asn		Met	Leu	Pro	2109 Lys		Asp	Gly			) Pro	Asp
55	Leu	Glu	2115 Lys		Ser	Cys	Ala	2120 Lys -		Cys	Ser	Ser	212! Tyr	Arg	Lys	Trp
	Ile 2149	Glu	Ser	Lys	Gly	Lys 2150		Phe	Glu	Lys				Ala	Tyr	
			Lys	Asp	Lys 2165	Cys							His	Asp		_
60	Phe	Cys	Glu	Thr 2180	Leu		Thr	Ser	Ser 2189		Ala	Lys	Asp			Lys
	Thr	Leu	Gly 2195	Pro		Lys	Pro	Asn 2200	Asn		Glu	Gly	Lys 2209		Ile	Phe
65	Asp	Asp 2210	Asp		Thr	Phe	Lys 221	His		Lys	Asp	Cys 2220	Asp	Pro	Cys	Leu
																•

	Lys Phe 2225	Ser Val	Asn Cys 223	Lys Ly	s Asp	Glu Cys 223	Asp Asn	Ser Lys	
	Thr Asp	Cys Arg			r Ile	Asp Ala 2250	Thr Asp	Ile Glu	
5	Gly Val	Asp Ser 226	Thr Val	Leu Gl	u Met 2265	Arg Val	Ser Ala	225 Asp Ser 2270	Lys
		2215		22	u Glu 80	Asn Ala	228	Gly Ala	
10 -	229	U		2295			Cys Arg	Asn Val	
	2305		231	0		231	5	Ala Lys	2320
15			2325			2330		Val Glu 233	5
15		234	U		2345	5		His Arg 2350	
		2355		23	60	•	236	Glu Lys	_
20	237	0		2375			2380	Arg Phe	_
	2385		239	0		2399	5	Arg Ser	2400
25			2405			2410		Lys Asn 241	5
25		242	O		2425	5		Ser Ala 2430	
•		2435 .		24	40		244	Ile Asp	
30	245	0		2455			2460	Lys Lys	
	2465		247	0		247	5	Pro Gln	2480
			2485			2490		Glu Ala 249	5
35		250	0		2505	5		Thr Ala 2510	
		2515		25	20		252	Glu Glu 5	
40	253	0		2535			2540	Pro Val	
	2545		255	0		255	5	Pro Pro	2560
45			2565			2570		Thr Pro 257	5
45		258	0		2585	5		Val Thr 2590	
		2595		26	00		260	Thr Tyr	
50	261	0		2615			2620	Phe Gln	
	2625		263	0		2639	5	Leu Ser	2640
			2645			2650		Lys Arg 265	5
55		266	0		2665	5		Thr Asp 2670	
		2675		26	80		268	Met Asp	
60	269	0		2695			2700	Leu Ile	
	2705		271	0		2719	5	Gly Asn	2720
			2725			2730	_	Asn Thr 273	Pro 5
65	Ser Asp	Thr Gln	Asn Asp	Ile Gl	n Asn	Asp Gly	Ile Pro	Ser Ser	Lys

				274	^					_						
		Thr	275	>	Glu			276	0	Lys			276	ς.	Ser	
5	Tyr	Leu 277	Gln 0	Ser	Glu	Pro	Asn 277	Thr 5	Glu	Pro	Asn	Met 278	Leu	Gly	Tyr	Asn
	Val 278	Asp 5	Asn	Asn	Thr	His 279	Pro	Thr	Thr	Ser	His 2799	His	Asn	Val	Glu	
	Lys	Pro	Phe	Ile	Met 280	Ser		His	Asp	Arg 281	Asn	Leu	Phe	Ser	Gly 281	
10		Tyr		2820	)				282	Gly	Asn			283	Asn	Ile
		Asp	283	>				284	Ser	Leu			284	Asn 5	His	
15		Tyr 285	U				285	5				286	Asp 0	Leu		
	Asp 286	Ala 5	Leu	Ser	Gly	Asn 2870	His	Ile	Asp	Ile	Tyr 2875	Asp	Glu	Met	Leu	Lys 2880
	Arg	Lys	Glu	Asn	Glu 2885	Leu 5	Phe	Gly	Thr	Lys 289	His	His	Thr	Lys	His 289	Thr
20	Asn	Thr	Tyr	Asn 290	Val	Ala	Lys	Pro	Ala 290	Arg	Asp	Asp	Pro	Ile 291	Thr	Asn
	Gln	Ile	Asn 2915	Leu	Phe	His	Lys	Trp 292	Leu		Arg	His	Arg 292	Asp	Met	Cys
25	Glu	Lys 293	Trp	Lys	Asn	Asn	His 293	Glu 5	Arg	Leu	Pro	Lys 294	Leu	Lys	Glu	Leu
	294	-				2950	)				2955	Ser	Gly			2960
		Asn			296					2970	Ile	Gln			297	Asp
30	Asn	Pro	Lys	Thr 2980	Lys )	Asn	Glu	Ile	Thr 2989	Asn	Met	Asp		Asn 299	Pro	Asp
		Ser	2995	j .				3000	Asp	Asp			Lys	Tyr.	Asn	
35		Tyr 301	)				301	5				3020	Tyr	His	_	
	302					3030	)				3035	Val	Asp			3040
		Thr			Asn 3045	Met 5	Asp	Val	Pro	Thr 3050	Lys O	Met	His	Ile	Glu 3059	Met
40	Asn	Ile	Val	Asn 3060	)											
	(2) INFO	RMAT:	ION F	OR S	SEQ I	D NO	0:15	:								
45	(i) SE	QUENC	CE CH	IARAC	TER	STIC	CS:									
	(B) '	LENG:	nuc	cleid	aci	iď	irs					÷				
F0	(C) : (D) '	STRAI TOPOI	NDEDN LOGY:	ESS:	sir near	ngle										
50	(ii) MO															
	(iii) (iv) AN				.: NC	)										
55	(xi) SE	<u> </u>	E DE	SCRI	PTIC	DN: S	SEQ :	ID N	0:15:							
	TCCAAGCT	GT TI	TTTT	TTCI	TTT	TCTA	GTT.	TTTC	CATT	GT A	TATT	CGTC.	A AA	TACG'	TACA	60
en	CATATATATATATATATATATATATATATATATATATAT	TT TI	TCCC	CAGA	TCA	CATA	TAG	TACG	ACTA	AG A	AACA	AAAT	A AC	ATCA	CAAC	120 180
60	AAACATAG' TTGCATGT'	ra gi	GATA	ACTA	CTA	TATC	ATA	TACA	CCAC	TA C'	TAAC'	TATC	A CT	ACAT	AGTA	240 300
	ACAGTAGTA ATTGTTTG	TA TI	ACAT	ACAC	TAT	TAAT	ATG	TATT	TATG	TT A'	TAAT	GTA	G AC	ТАТС	ממדיו	420
65	CATTTATG	AC AI	CACC	TAGT	CGG	ATTA GAAG	CAT .	acag Acaa	ACGC.	GG A	AAAA GCCA	CAGT(	G TA	ratg: gcgg:	rgtg rggt	480 540

	ССТССТАСТС	CCCCTACTAC	ma cmccma a a				
	CTCACCCATC	CTARCATECT	TAGTGGTAAA	GGGAAGAAGG	ATACATCTGA	GTATATTTAT	600
	AAAAATCCTC	ATCCTAAAAAA	ATATATTATT	GTTGGAGAAA	AAGTGTACGA	AGAAAAAGTG	<b>6</b> 60
	CCTCCTACTT	CCCNANCACC	TACCACTATE	GCGTTGAAAG	GAAATTTGAA	CACAGCAAAT	720
5	GAGCGTGTTA	ATGGTGATGG	TABCAGIATI	GAAACGTGCA	CCCTTGTAAA	AGAATATTAT	~ <b>7</b> 80
•	GTAAACCGTT	TTTCCCATAC	) CTTCCTCCC	CCGTGCAGAA	AAGACGCAAA	AAATGAAGAT AAAAGATAGT	840
	CAACAGGGTG	ATAATAAACT	ACTIGGIGGE	CCTCCCTATA	ACAATAGGAT	AAAAGATAGT TTTATGTGAT	900
	TATAATTTGG	ΔΑΤΟΤΑΤΑΘΑ	CACAACCTCC	ACCACCCAMA	GACGATTACA	AGAGGTGTGT	960
	ATGGCAGCAA	AATACGAAGG	AAACTCAATA	ACGACGCATA	AGTTGTTGTT	TCAACGAACT	1020
10	AATGAGGATT	CTGCTTCCCA	ATTATCTACT	CTATTACCACALL	CAACACAACA	AGATATAGGT	1080
	GATATCGTAA	GAGGAAAAGA	TCTATATCTC	CCTTATCATA	ATARACARA	AGATATAGGT	1140
	AAAAAATTAG	AACAGAAATT	GAAAGATATT	TTCAACAAAA	TACATAACCA	CGTGATGAAG	1200
	ACGAATGGCG	CACAAGAACG	CTACATAGAT	GATGCCAAAG	CACCACATTT	TTTTCAATTA	1260
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15	CCAAAAGAAG	CTAATTATTT	TATAAAAACA	GCGTGTAATG	TAGGAAAAGG	AACTAATCCT	1440
	CAATGCCATT	GCATTGGTGG	AGATGTTCCC	ACATATTTCG	ATTATGTGCC	CCACTATCTT	1500
	CGCTGGTTCG	AGGAATGGGC	AGAAGACTTT	TGCAGGAAAA	ΔΑΑΑΑΑΑΑ	ΔΟΤΔGΔΔΔΔΤ	1560
	TTGCAAAAAC	AGTGTCGTGA	TTACGAACAA	AATTTATATT	GTAGTGGTAA	TCCCTACCAT	1620
	TGCACAAAAA	CTATATATAA	AAAAGGTAAA	CTTGTTATAG	GTGAACATTG	TACAAACTCT	1600
20	TCTGTTTGGT	GTCGTATGTA	TGAAACTTGG	ATAGATAACC	AGAAAAAAGA	ልጥጥጥር ተልልልል ል	1740
	CAAAAAAGAA	AATACGAAAC	AGAAATATCA	GGTGGTGGTA	GTGGTAAGAG	TCCTAAAACC	1000
	ACAAAACGGG	CTGCACGTAG	TAGTAGTAGT	AGTGATGATA	ATGGGTATGA	ΔΔCTΔΔΔTTT	1860
	TATAAAAAAC	TGAAAGAAGT	TGGCTACCAA	GATGTCGATA	ΔΑΤΤΤΤΤΔΔΔ	<b>እልጥልጥጥልአል</b> ሮ	1020
25	AAAGAAGGAA	TATGTCAAAA	ACAACCTCAA	GTAGGAAATG	AAAAAGCAGA	TAATGTTGAT	1980
23	TCCTCTCCTCT	AAAAATATGT	AAAAACATTT	TCTCGTACAG	AAATTTGTGA	ACCGTGCCCA	2040
	ACTCCAAAAA	TGGAAAAAGG	TGGTCCACCA	TGGAAAGTTA	AAGGTGACAA	AACCTGCGGA	2100
	CATAAATCAC	CAAAGACATA	CGATCCTAAA	AATATTACCG	ATATACCAGT	ACTCTACCCT	2160
	GGTGGTGGTC	AGCARARIAI AAATTAAAA	ACTAAAAAAA	TATAAAAATT	TTTGTGAAAA	AGGTGCACCT TAGTAGTAAA	2220
30	ATAATAATA	ATTGTGTAGA	ACGAACATGO	CACAACTTTA	AACATAGGCC	TAGTAGTAAA ACAAACCGTT	2280
	AAGTCCTATA	ATGTTTTTTT	TTCCCATTCC	GTTCATCATA	TCTTT CACCA	TTCTGTAGAG	2340
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	GAATGGATGG	CAATAAAAGA	CCATTTTGGA	AAGCAAACAG	ATATTGTCCA	ACAAAAAGGT	2520
35	CTTATCGTAT	TTAGTCCCTA	TGGAGTTCTT	GACCTTGTTT	TGAAGGGCGG	TAATCTGTTG	2640
	CAAAATATTA	AAGATGTTCA	TGGAGATACA	GATGACATAA	AACACATTAA	GAAACTGTTG	2700
	GATGAGGAAG	ACGCAGTAGC	AGTTGTTCTT	GGTGGCAAGG	ACAATACCAC	AATTGATAAA	2760
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40	AAAAAAGCAC	AACAAGAAAG	TCGTGGTCGC	TCCGCCGAAA	CCCGCGAAGA	CGAAAGGACA	2880
40	CAACAACCTG	CTGATAGTGC	CGGCGAAGTC	GAAGAAGAAG	AAGACGACGA	CGACTACGAC	2940
	GAAGACGACG	AAGATGACGA	CGTAGTCCAG	GAGGAGGAAG	AGGGAAAGGA	GGAAGGAACG	3000
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45	CCAAGTGGTG	TCACTACTCC	AGGTGGAAAA	GAAAAATTCC	CCAATTGGAA	GTGTGTCACA	3180
10	ACACCACCAT	TATACCTACC	TCCTTTTATCA	AAAGACGGCG	CTATATGTGT	GCCACCCAGG TGACGAGACC	3240
	ACGGAGGTGT	CCACTCAACC	CACTTCCCCC	CCCTCACACT	GTCGTGGTGG	TGACGAGACC AAAACTACGT	3300
	ACTGCGTTTA	TTGAGTCCGC	TCCDDTDCDC	A COMMUNICATION	TCTCCCATA	GTATAAAGAA	3360
	GAGAAAAAC	CACCAGCAAC	ACAAGATGGA	GCGGGACTTG	CACTATCACT	CCCAGAACCG	3420
50	TCACCACCGG	GAGAGGACCC	CCAAACACAA	TTACAACAAA	CTGGTGTTAT	ACCCCCGAT	3480
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<b>5</b> 5	AATAGTGTCA	AAACCCCCCA	ACAAACCTGG	TGGGAAAACA	TCGCGAAGGA	TATCTGGAAT	3840
	GCTATGGTAT	GTGCACTAAC	ATATAAAGAA	AATGACGCCA	GAGGCACAAG	TGCCAAAATA	3900
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	GAGAAATACC	AATACACAAA	TGTCAAACTC	GAAGATGAAA	GTGGTGCCAA	AAGCAACGAC	4020
CO .	ACCATCCAAC	CCCCCACGTT	AAAAAATTTT	GTGGAAATAC	CTACATTTTT	TCGTTGGTTA	4080
60	CATGAGTGGG	GAAACAGTTT	TTGTTTTGAG	AGAGCAAAAC	GATTGGCACA	AATAAAACAT	4140
	GAGTGTATGG	ATGAGGATGG	TGAAAAACAA	TATAGTGGGG	ATGGGGAATA	TTGTGAAGAA	4200
	ATTTTTAGTA	AGCAATATAA	TGTTCTCCAG	GATTTAAGTT	CCAGTTGCGC	TAAACCTTGT	4260
	MATCARCAR	AAACGTGGAT	AGAAAAAAA	AAAACAGAAT	ATGAGAAACA	ACAAAAGGCA	4320
<b>6</b> 5	THIGHACAAC	AAAAAAGTAA	TTACGAAAAT	GAACAAAAAG	ACAAATGCCA	AACACAAAGT	4380
	THAT TAN I AM I G	CIAAIGAAIT	TICIAGAACA	CIAGGAGCGT	CCCCTACAGC	TGCAGAATTT	4440

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TTACAAAAGT TAGGATCATG TAAAAATGAT AATGGATATG AGAATGGAGA GGATAATAAA 4500
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     GAAATGGTTG TCAGTGATGA CAGTACAAAT ACATTTGAAC ATTTAGGCGA TTGTAAAAGC 4740
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     GATATATGTA CTCTGGAAAA AAAAATTAAG AATGGGCAAG AAGGTGATAA AAAATATATC 4860
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     GATGATTATG AGGAACAAAA CCCAGAAAAC AAAGTGGAAC AACCTAAATT TTGTCCAGAT 5400
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20
     GTCATACCCG CCCTCATGTC TTCTACCATC ATGTGGAGTA TTGGCATCGG TTTTGCTGCG 5700
     TTCACTTATT TTTATCTAAA GAAAAAACC AAATCATCTG TTGGAAATTT ATTCCAAATA 5760
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     GGAGATGAAA AATATGCATT TATGTCTGAT ACTACTGATA TAACTTCATC CGAAAGTGAG 5940
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30
     TTACATGATA ATGTGGATAA TAATACCCAT CCTACCATGT CACGTCATAA TATGGACCAA 6240
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35
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    TGGTTAĞATA GGCATAGAGA TATGTGCGAA AAGTGGAAAA ATAATCACGA ACGGTTACCC 6660
     AAATTGAAAG AATTGTGGGA AAATGAGACA CATAGTGGTG ACATAAATAG TGGTATACCT 6720
     AGTGGTAACC ATGTGTTGAA TACTGATGTT TCTATTCAAA TAGATATGGA TAATCCGAAA 6780
40
    ACAATGAATG AATTTACTAA TATGGATACA AACCCCGACA AATCTACTAT GGATACTATA 6840
    TTGGATGATC TAGAAAAATA TAACGAACCC TACTACTATG ATTTTTATAA ACATGATATC 6900
    TATTATGATG TAAATGATGA TAAAGCATCT GAGGATCATA TAAATATGGA TCATAATAAG 6960
     ATGGATAATA ATAATTCGGA TGTCCCCACT AACGTACAAA TTGAAATGAA TGTCATTAAT 7020
     AATCAGGAGT TACTACAAAA TGAATATCCT ATATCGCATA TGTAGGGAAT ATGAAAATAA 7080
45
     TAGATGTATA TATGTTTTTT TCTTTTTTTG TGTGTGTGCA GTTTATATTT TTTATTTGTA 7140
     TATATTTTT TTTTTGTGCA TTTGTCTATT TTTTATTTGT GCTTTATATA TATATATATT 7260
     TTATTCAGCT TGGACTTAAC CAGGCTGAAC TTGCT
```

- 50 (2) INFORMATION FOR SEQ ID NO:16:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 2182 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: protein
- 60 (iii) HYPOTHETICAL: NO
  - (iv) ANTI-SENSE: NO
    - (v) FRAGMENT TYPE: N-terminal

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

•	(1111)	2021	· ·			ON.	SEQ	TD M	0:16	:						
_				Gly	5					10					7 5	
5				Lys 20					25					3 0		
			33	Leu				40			•		45			
10		50		Asp			55					60				
	60			Asn		70					75					0.0
15				Val	85					90					95	
15				Cys 100					105					110		
			TTD	Leu				120					125			
20		130		Asp			135				•	140				
	145			Asp		150					155					160
25				Leu	162					170					175	
23				Thr 180					185					190		
			T 3 2	Leu				200					205			
30		210		Arg			215					220				
	4,45			Arg		230					235					240
35				Lys	245					250					255	_
				Ala 260					265					270		_
			2/5	Asn				280					285			
40		290		Ala			295					300				
	305			Gly Val		310					315					220
45				Arg	325					330					225	
-				340 Tyr					345					350		
			355	Thr				360					365			_
50		370		Cys			375					380				
	385			Lys		390					395					400
55				Gly	405		-			410					415	
,				420 Ser					425					430		
			435	Leu				440					445			
60		450		Asn			455					460				
	465			Ala		470					475					480
65					485					490					495	_
	1111	* *1C	261	Arg	TITE	Giu	116	Cys	GIU	PLO	cys	Pro	irp	cys	GIA	Leu

				500												
	Glu	Lys	Gly 515		Pro	Pro	Trp	Lys 520	505 Val	Lys	Gly	Asp	Lys 525		Cys	Gly
5		Ala 530					535	Asp	Pro			540	Thr	Asp		
	242					550					555	Leu				E 6 0
10		Phe			565					570					575	Trp
10		Cys		580					585					590		
		Val	595					600					605			
15		Ser 610 Ser					615					620				
	625	Asn				630					635					640
20		Cys			645					650					655	
		Lys	Asp	660					665					670		
25	Leu	Ile 690	675 Val	Phe	Ser	Pro	Tyr	680 Gly	Val	Leu	Asp		685 Val	Leu	Lys	Gly
20	Gly 705	Asn	Leu	Leu	Gln	Asn 710	695 Ile	Lys	Asp	Val	His 715	700 Gly	Asp	Thr		
•	Ile	Lys			725	Lys				730	Glu				Ala	
30		Leu		740					745	Ile				750	Gln	His
•		Lys	/55					760					765			
35		Lys 770					775					780				
	785	Glu Glu				790					795			_		800
40		Gln			805					810					815	
·		Glu		820					825					830		
45		Lys	835					840					845			_
45	Leu	850 Lys				Gly	855					860				
	865 Phe	Pro	Asn	Trp	Lys	870 Cys	Val	Thr	Pro	Ser	875 Gly	Val	Ser	Thr		880 Thr
50	Ser	Gly	Lys	Asp	885 Gly	Ala	Ile	Cys	Val 905	890 Pro	Pro	Arg	Arg		895 Arg	Leu
•	Tyr	Val	Gly 915		Leu	Ser	Gln	Trp 920		Ser	Arg	Gly	Gly 925	910 Asp	Glu	Thr
55		Glu 930					935					940	Gln			
	945	Lys -				950					955					960
60		Leu			965					970					975	
		Gly Asp		980					985					990		_
		Leu	995					1000	)				1005	5		
65		1010	)	- <b></b>			1015	5				1020		nap	*16	<b>n</b> eu

	Tyr 1025					1030	)				1035	5				1040
_	Ser				104	5				105	0	-			105	<b>5</b> .
5	Ser	Thr	Glu	Gln 1060	Glu O	Lys	Glu	Lys	Met 1069	Lys 5	Gln	Ile	Gln	Ala 107	Lys	Ile
	Lys		1075	•				1080	0				108	Val	Thr	=
10		TOAL	,				109	5				110	Asn 0	Ile		
	Asp 1105					1110	)				1115	5				1120
•	Ala .				112	5				1130	0				113	5
15	Ala			1140	)				114	5				115	n -	
	Tyr		1155	5				1160	0				116	5		_
20		1170	)				117	5				118	0			
	Phe .					1190	)				1195	5		•		1200
05	Lys .				1209	5				1210	0				121	5
25	Lys			1220	)				122	5				123	n	_
	Gln '		1235	•				1240	)				124	5		
30		1250	)				125	5				1260	0	_		-
	Gln 1265					1270	)				1275	5				1280
	Lys 2				1289	5				1290	)				129	5
35	Arg '			1300	)				1309	5				131	Lys	Leu
	Gly :		1315	5				1320	)				1325	5		-
40		1330	)				133	5				1340	0			_
	Asp :					1350	)				1355	;	_			1360
45	Gly :				1365	5				1370	)				137	5
45	Asp :			1380	)				1385	5				139	n	
	Ser i		1395	5				1400	)				140	5		
50		1410	) .				1419	5				1420	0			
	Val (					1430	)				1435	,				1440
	Gln (				1445	5				1450	)				145	Arg 5
55	Trp :			1460	)				1465	5				147	0 _	
	Lys 1		1475	•				1480	)				148	5		
60		1490	)				1499	5				1500	Gln O	Lys		
	Asp '					1510	) .				1515	•				1520
	Asn :				1525	5				1530	)				153	Ile 5
65	Lys 1	Pro	Cys	Asp			Asp	Gln	Phe			Ser	Cys	Gly	Leu	Asn

				154					154	5				155	0	
· •		~ -	700	Э.		_		156	Asn 0	Asn			156	Val	Leu	
5		13/	U	ьуѕ	Leu	GIN	ьуs 157	Lys 5	Ile	Ser	Glu	Cys	Lys	Glu	Gln	
	400	_				TOD	U				159	5				Lys 1600
10					700	5				Asp 161	O.				161	Pro
10				T02	U				162	5				162	Δ.	Pro
			T03:	9				164	U	Thr			764	5		
15		T 6 2	U				165	5		Glu Leu		166	0			
	TOO	<b>&gt;</b>				1670	כ			Pro	167	5				1600
20					TPR:	5				169 His	0				169	5
				T.70	U				170	5 Ile				171	^	
			1/13	•				1/2	U	Thr			172	5		
25	Leu	Phe	U				173	5				174	٥			Thr
	1/4	<b>5</b>			Asn	Arg	)				175	5				1760 Lys
30	Gly	Lys	Thr	Tyr	1769   Ile		Met	Glu	Gly	1770 Asp	Ser	Ser	Gly		Glu	5 Lys
	Tyr	Ala	Phe 1795	Met	Ser	Asp	Thr	Thr 180	178 Asp	Ile	Thr	Ser	Ser	179 Glu	0 Ser	Glu
35	Tyr	Glu 181	Glu		Asp	Ile	Asn 181	Asp	Ile	Tyr	Val	Pro 182	180 Gly	Ser	Pro	Lys
	Tyr	Lys	Thr	Leu	Ile	Glu	Val	Val	Leu	Glu	Pro	Car	Tara	Arg	Asp	Thr
	102	<b>&gt;</b>				Asn	) '			Ser	1835 Asp	5			Ser	1840 Asp
40	Thr	Pro	Pro	Pro 1860	Ile		Asp	Asp	Glu 186	1850 Trp	Asn	Gln	Leu			5 <b>As</b> p
	Phe	Ile	Ser 1875	Asn		Leu	Gln	Asn 1880	Thr	Gln	Asn	Thr	Glu 188		o Asn	Ile
45	Leu	His 1890	Asp	Asn	Val	Asp	Asn 189	Asn	Thr	His	Pro	Thr 190	Met	Ser	Arg	His
	190	>				1910	Phe	Ile			1915	His	Asp			Leu 1920
F0					1925	5				Met 1930	)				193	Asn 5
50				1940	)				1945		-			1950	0	
			1955	•				1960	)	Asn			1969	5	_	
55		1970		•			197	ร		Asn -		198	0		_	
	1985	5				1990	)			Leu	1995	i				2000
60					2005	5				Val 2010 Phe	)				201	5
				2020	)				2025	5				2030	<u> </u>	_
						1 - 1 11	1.370	7.77	1.770	000	/\ C \	H 3 C	G 1 11	Δνα	T.OIL	Pro
			Asp 2035	,				2040	)	Thr			2045	5		

	Ser Gly Ile Pro Ser Gly Asn His Val Leu Asn Thr Asp Val Ser Il 2065 2070 2075 20	
_	Gln Ile Asp Met Asp Asn Pro Lys Thr Met Asn Glu Phe Thr Asn Me	t
5	Asp Thr Asn Pro Asp Lys Ser Thr Met Asp Thr Ile Leu Asp Asp Le	u
	Glu Lys Tyr Asn Glu Pro Tyr Tyr Tyr Asp Phe Tyr Lys His Asp Il 2115 2120 2125	
10	Tyr Tyr Asp Val Asn Asp Asp Lys Ala Ser Glu Asp His Ile Asn Me 2130 2135 2140	
	Asp His Asn Lys Met Asp Asn Asn Asn Ser Asp Val Pro Thr Asn Va 2145 2150 2155 21	60
15	Gin lie Glu Met Asn Val Ile Asn Asn Gln Glu Leu Leu Gln Asn Gl 2165 2170 2175	u
13	Tyr Pro Ile Ser His Met 2180	
	(2) INFORMATION FOR SEQ ID NO:17:	
20	<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 30 base pairs</li></ul>	
	(B) TYPE: nucleic acid (C) STRANDEDNESS: single	
25	(D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: CDNA (iii) HYPOTHETICAL: NO	
	(iv) ANTISENSE: NO (v) FRAGMENT TYPE:	
30	(vi) ORIGINAL SOURCE:	
•	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:	
<b>3</b> 5	ATCGATCAGC TGGGAAGAAA TACTTCATCT	30
	(2) INFORMATION FOR SEQ ID NO:18:	
	<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 30 base pairs</li></ul>	
40	(B) TYPE: nucleic acid	
	<pre>(C) STRANDEDNESS: single (D) TOPOLOGY: linear</pre>	
45	(ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO	
	(iv) ANTISENSE: NO	
	(v) FRAGMENT TYPE: (vi) ORIGINAL SOURCE:	
50	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:	
	ATCGATGGGC CCCGAAGTTT GTTCATTATT	3 0
55	(2) INFORMATION FOR SEQ ID NO:19:	
•	(i) SEQUENCE CHARACTERISTICS:	
	(A) LENGTH: 30 base pairs (B) TYPE: nucleic acid	
	(C) STRANDEDNESS: single	
60 -	(D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO	
	(iv) ANTISENSE: NO	
6E .	(1.) EDACMENT TUDO	

	(VI) ORIGINAL SOURCE:	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:	
5	TCTCGTCAGC TGACGATCTC TAGTGCTATT	• · 30
	(2) INFORMATION FOR SEQ ID NO:20:	
10	<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 30 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: linear</li> </ul>	
15	<pre>(ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO (iv) ANTISENSE: NO (v) FRAGMENT TYPE: (vi) ORIGINAL SOURCE:</pre>	
20	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:	
	ACGAGTGGGC CCTGTCACAA CTTCCTGAGT	
25		30
23	(2) INFORMATION FOR SEQ ID NO:21:	
30	<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 17 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: linear</li> </ul>	
35	<pre>(ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO (iv) ANTISENSE: NO (v) FRAGMENT TYPE: (vi) ORIGINAL SOURCE:</pre>	
40	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:	·
40 .	AGACCTCAAT TTCTAAG	17
	(2) INFORMATION FOR SEQ ID NO:22:	
45	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 18 base pairs  (B) TYPE: nucleic acid	
50	<pre>(C) STRANDEDNESS: single (D) TOPOLOGY: linear</pre>	
50	<pre>(ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO (iv) ANTISENSE: NO (v) FRAGMENT TYPE:</pre>	
<b>5</b> 5	(vi) ORIGINAL SOURCE:	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:	
60	AATCGCGAGC ATCATCTG	18
_	(2) INFORMATION FOR SEQ ID NO:23:	
65	<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 20 base pairs</li><li>(B) TYPE: nucleic acid</li></ul>	

```
(C) STRANDEDNESS: single
            (D) TOPOLOGY: linear
           (ii) MOLECULE TYPE: cDNA
5
           (iii) HYPOTHETICAL: NO
           (iv) ANTISENSE: NO
           (v) FRAGMENT TYPE:
           (vi) ORIGINAL SOURCE:
10
           (xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:
     CCRAGRAGRC AARAAYTATG
                                                              20
             (2) INFORMATION FOR SEQ ID NO:24:
15
           (i) SEQUENCE CHARACTERISTICS:
            (A) LENGTH: 18 base pairs
            (B) TYPE: nucleic acid
            (C) STRANDEDNESS: single
20
            (D) TOPOLOGY: linear
           (ii) MOLECULE TYPE: cDNA
          (iii) HYPOTHETICAL: NO
           (iv) ANTISENSE: NO
25
           (v) FRAGMENT TYPE:
          (vi) ORIGINAL SOURCE:
          (xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:
30
     CCAWCKKARR AATTGWGG
                                                              18
             (2) INFORMATION FOR SEQ ID NO:25:
          (i) SEQUENCE CHARACTERISTICS:
35
            (A) LENGTH: 291 amino acids
            (B) TYPE: amino acid
            (C) STRANDEDNESS: single
            (D) TOPOLOGY: linear
40
          (ii) MOLECULE TYPE: peptide
          (iii) HYPOTHETICAL: NO
          (iv) ANTISENSE: NO
          (v) FRAGMENT TYPE: internal
          (vi) ORIGINAL SOURCE:
45
          (xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:
     10
50
     Xaa Xaa Xaa Val Cys Ile Pro Asp Arg Tyr Gln Leu Cys Met Lys
     40
     55
                          55
                                           60
     70
                                       75
     Xaa Asp Phe Cys Lys Asp Ile Arg Trp Ser Leu Gly Asp Phe Gly Asp
                   85
                                    90
                                                     95
60
     Ile Ile Met Gly Thr Asp Met Glu Gly Ile Gly Tyr Ser Lys Xaa Xaa
                                105
                                                 110
     Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Thr Asp Glu Lys Ala Gln Gln
           115
                             120
                                              125
     Arg Arg Lys Gln Trp Trp Asn Glu Ser Lys Ala Gln Ile Trp Thr Ala
65
```

```
150
                              155
   Cys Xaa Xaa Xaa Xaa Xaa Xaa Xaa Glu Pro Gln Ile Tyr Arg Trp
              165
                          170
                                        175
5
   Ile Arg Glu Trp Gly Arg Asp Tyr Val Ser Glu Leu Pro Thr Glu Val
           180
                        185
                                     190
   Gln Lys Leu Lys Glu Lys Cys Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
                      200
   Xaa Xaa Cys Xaa Val Pro Pro Cys Gln Asn Ala Cys Lys Ser Tyr Asp
10
                   215
                                220
   Gln Trp Ile Thr Arg Lys Lys Asn Xaa Xaa Xaa Xaa Xaa Xaa Xaa
                 230
                              235
   245
                           250
15
   260
                        265
                                     270
   275
                      280
   Cys Xaa Cys
20
      290
         (2) INFORMATION FOR SEQ ID NO:26:
        (i) SEQUENCE CHARACTERISTICS:
25
```

- - (A) LENGTH: 271 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
  - (iii) HYPOTHETICAL: NO
  - (iv) ANTISENSE: NO
  - (v) FRAGMENT TYPE: internal
- (vi) ORIGINAL SOURCE: 35

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

	1				5					10					15	Xaa
40				20	Xaa				25					30		_
			35		Xaa			40					45			
45	Xaa	Xaa 50	Xaa	Xaa	Xaa	Xaa	Xaa 55	Xaa	Xaa	Xaa	Xaa	Xaa 60	Xaa	Xaa	Xaa	Xaa
	65				Xaa	70					75				-	80
	Ser	Phe	Leu	Asp	Tyr 85	Gly	His	Leu,	Ala	Met 90	Gly	Asn	Asp	Met	Asp 95	Phe
50	Gly	Gly	Tyr	Ser 100	Thr	Xaa	Xaa	Xaa	Xaa 105	Xaa	Xaa	Хаа	Xaa	Xaa 110	Xaa	Xaa
	Xaa	Xaa	Xaa 115	Xaa	Xaa	Xaa	Ser	Glu 120	His	Lys	Ile	Lys	Asn 125	Phe	Arg	Lys
วิ <b>ว</b> ิ	Glu	Trp 130	Trp	Asn	Glu	Phe	Arg	Glu	Lys	Leu	Trp	Glu 140	Ala	Met	Leu	Ser
	Glu 145	His	Xaa	Xaa	Xaa	Xaa 150	Xaa	Xaa	Cys	Xaa	Xaa 155	Xaa	Xaa	Xaa	Xaa	Glu 160
	Leu	Gln	Ile	Thr	Gln 165	Trp	Ile	Lys	Glu	Trp 170	His	Gly	Glu	Phe	Leu 175	
60	Glu	Arg	Asp	Asn 180	Arg	Ser	Lys	Leu	Pro 185	Lys	Ser	Lys	Cys	Xaa 190		Xaa
	Xaa	Xaa	Xaa 195	Xaa	Xaa	Cys	Xaa	Glu 200	Lys	Glu	Cys	Ile	Asp 205	Pro	Cys	Met
65	Lys	Tyr 210	Arg	Asp	Trp	Ile	Ile 215		Ser	Lys	Phe	Xaa 220		Xaa	Xaa	Xaa

20

60

```
230
                235
 245
              250
                      255
 Xaa Xaa Xaa Cys Xaa Xaa Xaa Xaa Xaa Cys Xaa Cys
5
      260
             265
```

#### (2) INFORMATION FOR SEQ ID NO:27:

```
10
             (i) SEQUENCE CHARACTERISTICS:
```

- (A) LENGTH: 277 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide (iii) HYPOTHETICAL: NO

- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE: internal
- (vi) ORIGINAL SOURCE:

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

25	<u> </u>				5					10					1 =	Xaa
٠				20	Xaa				25					3 0	Arg	_
20			35		Leu			40					45			
30		50			Xaa		55					60				
	05				Xaa	70					75					0.0
35					Thr 85					90					95	Thr
				T00	Asp				105					110	Xaa	
40			TT2		Xaa			120					125			
40		130			Trp		135					140				
	145				Xaa	150					155					160
45					Phe 165					170					175	Cys
				180	Lys				185					190	Xaa	
			195		Asp			200					205	Ser		_
50		210			Lys		215					220	Xaa			
٠	225				Xaa	230					235					240
55	Xaa	Xaa	Xaa	Xaa	Xaa 245	Xaa	Xaa	Xaa	Xaa	Xaa 250	Xaa	Xaa	Cys	Xaa	Xaa 255	Xaa
	Xaa	Xaa	Xaa	Xaa 260	Xaa	Xaa	Xaa	Xaa	Xaa 265		Xaa	Xaa	Xaa	Xaa 270	Xaa	Xaa
	Xaa	Cys	Xaa 275	Xaa	Cys									J. <b>J</b>		

(2) INFORMATION FOR SEQ ID NO:28:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 282 amino acids
- 65 (B) TYPE: amino acid

```
(C) STRANDEDNESS: single
           (D) TOPOLOGY: linear
          (ii) MOLECULE TYPE: peptide
5
          (iii) HYPOTHETICAL: NO
          (iv) ANTISENSE: NO
         (v) FRAGMENT TYPE: internal
         (vi) ORIGINAL SOURCE:
10
         (xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:
     Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Val Cys Gly Pro Pro Arg Arg
15
              20
                              25
    Gln Gln Leu Cys Leu Gly Tyr Ile Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
                           40
    55
    20
                                    75
    Ala Ile Leu Gly Ser Tyr Ala Asp Ile Gly Asp Ile Val Arg Gly Leu
                                 90
    Asp Val Trp Arg Asp Ile Asn Thr Asn Xaa Xaa Xaa Xaa Xaa Xaa
25
                              105
    Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Lys Lys Gln Asn Asp Asn
                          120
    Asn Glu Arg Asn Lys Trp Trp Glu Lys Gln Arg Asn Leu Ile Trp Ser
                       135
30
    Ser Met Val Lys His Ile Xaa Xaa Xaa Xaa Cys Xaa Xaa Xaa Xaa
                    150
                                    155
    Xaa Xaa Xaa Ile Pro Gln Phe Leu Arg Trp Leu Lys Glu Trp Gly
                                 170
    Asp Glu Phe Cys Glu Glu Met Gly Thr Glu Val Lys Gln Leu Glu Lys
35
              180
                              185
    Ile Cys Xaa Xaa Xaa Cys Xaa Glu Lys Lys Cys Lys Asn Ala Cys
                          200
    Ser Ser Tyr Glu Lys Trp Ile Lys Glu Arg Lys Asn Xaa Xaa Xaa Xaa
                       215
                                       220
40
    230
    250
    45
                              265
    Xaa Xaa Xaa Xaa Xaa Cys Xaa Xaa Cys
            (2) INFORMATION FOR SEQ ID NO:29:
50
         (i) SEQUENCE CHARACTERISTICS:
           (A) LENGTH: 324 amino acids
           (B) TYPE: amino acid
           (C) STRANDEDNESS: single
55
           (D) TOPOLOGY: linear
         (ii) MOLECULE TYPE: peptide
         (iii) HYPOTHETICAL: NO
         (iv) ANTISENSE: NO
60
         (v) FRAGMENT TYPE: internal
         (vi) ORIGINAL SOURCE:
         (xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:
```

	1				5					10						
	_	Xaa	Xaa	Xaa 20		Xaa	Xaa	Ala	Cys	10 Ile	Pro	Pro	Arg		15 Gln	Lys
5	Leu	Cys	Leu 35	His	Tyr	Leu	Xaa	Xaa 40	25 Xaa	Xaa	Xaa	Xaa		30 Xaa	Xaa	Xaa•
	Xaa	Xaa 50		Xaa	Xaa	Xaa	Xaa		Xaa	Xaa	Xaa		45 Xaa	Xaa	Xaa	Xaa
	Xaa 65	Xaa	Xaa	Xaa	Xaa	Xaa	55 Xaa	Xaa	Xaa	Xaa		60 Xaa	Xaa	Xaa	Xaa	Xaa
10		Xaa	Xaa	Xaa	Xaa	70 Xaa	Xaa	Xaa	Xaa	Asp	75 Phe	Lys	Arg	Gln	Met	80 Phe
	Tyr	Thr	Phe	Ala 100	85 Asp	Tyr	Arg	Asp	Ile	90 Cys	Leu	Gly	Thr		95 Ile	Ser
15	Ser	Lys	Lys 115		Thr	Ser	Xaa	Xaa 120	105 Xaa	Xaa	Xaa	Xaa		110 Xaa	Xaa	Xaa
	Xaa	Xaa 130	Xaa	Xaa	Xaa	Lys	Ile 135	Ser	Asn	Ser	Ile	Arg	Tyr	Arg	Lys	Ser
	Trp 145	Trp	Glu	Thr	Asn	Gly 150	Pro	Val	Ile	Trp	Glu 155	Gly	Met	Leu	Cys	
20					165		Xaa			170	Xaa				175	
				TRO			Xaa		185	Xaa		•		100	Xaa	
25			エココ				Xaa	200					205	Gln		
		210					Gly 215					220				
20	223					230	Ala				235					240
30					245		Xaa			250					255	Cys
				200			Ile		265					270		
35			2/5				Xaa 	280					285			
		290					Xaa 295					300				
40	305	Xaa			лаа	310	Cys	хаа	Xaa	Xaa	Xaa 315	Xaa	Xaa	Xaa	Xaa	Cys 320
		naa	naa	Cys												
			(2)	INF	ORMA	MOITA	FOF	SEÇ	] ID	NO:3	0:					
45		(i	(A) (B) (C)	LENG TYPE STRA	TH: : an NDED	362 nino NESS	ACTER amin acid S: si	o ac l .ngle	ids							
50		. , ,	(D)	TOPC	LOGY	: li	.near	•								

(ii) MOLECULE TYPE: peptide (iii) HYPOTHETICAL: NO

- (iv) ANTISENSE: NO
- (v) FRAGMENT TYPE: internal
- (vi) ORIGINAL SOURCE:

55

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

Ala Cys Ala Pro Tyr Arg Arg Leu His Leu Cys Asp Tyr Asn Leu Xaa 60 10 25 40 65 Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Gln Leu Cys Thr Val Leu

```
50
                  55
    Ala Arg Ser Phe Ala Asp Ile Gly Asp Ile Val Arg Gly Lys Asp Leu
   65 - - - - 70 - - - 80
    Tyr Leu Gly Tyr Asp Asn Lys Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
5
             85
                         90
   100
                       105
   120
                                125
   Phe Phe Gln Leu Arg Glu Asp Trp Trp Thr Ser Asn Arg Glu Thr Val
10
                  135
   Trp Lys Ala Leu Ile Cys His Ala Xaa Xaa Xaa Xaa Xaa Xaa Xaa
               150
                           155
   15
             165
                         170
   Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Val Pro Gln Tyr Leu
                      185
   Arg Trp Phe Glu Glu Trp Ala Glu Asp Phe Cys Arg Lys Lys Lys
        195
                    200
                                205
   Lys Leu Glu Asn Leu Gln Lys Gln Cys Xaa Xaa Xaa Xaa Xaa Cys
20
                 215
                              220
   230
                           235
   Thr Asn Cys Ser Val Trp Cys Arg Met Tyr Glu Thr Trp Ile Asp Asn
25
             245
                         250
   265
                                  270
   275
                    280
                                285
30
   295
                              300
   310
                           315
   Xaa Xaa Xaa Xaa Xaa Xaa Cys Xaa Xaa Xaa Xaa Xaa Xaa Xaa
35
             325
                         330
   340
                      345
   Xaa Xaa Xaa Xaa Xaa Cys Xaa Xaa Cys
40
         (2) INFORMATION FOR SEQ ID NO:31:
       (i) SEQUENCE CHARACTERISTICS:
        (A) LENGTH: 411 amino acids
45
        (B) TYPE: amino acid
        (C) STRANDEDNESS: single
        (D) TOPOLOGY: linear
       (ii) MOLECULE TYPE: peptide
50
       (iii) HYPOTHETICAL: NO
       (iv) ANTISENSE: NO
       (v) FRAGMENT TYPE: internal
       (vi) ORIGINAL SOURCE:
55
       (xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:
   60
                      25
   Ala Cys Ala Pro Tyr Arg Arg Leu His Val Cys Asp Gln Asn Leu Xaa
                    40
```

```
65
                                        80
   85
                         90
   Met Leu Ala Arg Ser Phe Ala Asp Ile Gly Asp Ile Val Arg Gly Arg.
5
           100
                       105
   Asp Leu Tyr Leu Gly Asn Pro Gln Glu Xaa Xaa Xaa Xaa Xaa Xaa
        115
                    120
                                125
   135
                              140
   Xaa Xaa Xaa Xaa Xaa Xaa Asn Asp Pro Glu Phe Phe Lys Leu Arg
10
               150
                           155
   Glu Asp Trp Trp Thr Ala Asn Arg Glu Thr Val Trp Lys Ala Ile Thr
                         170
   Cys Asn Ala Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Cys Xaa Xaa Xaa
15
                       185
   200
   Xaa Xaa Xaa Val Pro Gln Tyr Leu Arg Trp Phe Glu Glu Trp Ala
                  215
                              220
20
   Glu Asp Phe Cys Arg Lys Lys Asn Lys Lys Ile Lys Asp Val Lys Arg
               230
                           235
   250
   25
          260
                       265
                                  270
   Xaa Xaa Xaa Xaa Cys Ile Ser Cys Leu Tyr Ala Cys Asn Pro Tyr
                    280
   Val Asp Trp Ile Asn Asn Gln Lys Glu Xaa Xaa Xaa Xaa Xaa Xaa Xaa
                  295
                              300
   30 -
               310
                           315
   330
   35
           340
                       345
   360
                                365
   375
                              380
40
   390
                           395
   Xaa Xaa Xaa Xaa Xaa Xaa Cys Xaa Xaa Cys
             405
45
         (2) INFORMATION FOR SEQ ID NO: 32:
       (i) SEQUENCE CHARACTERISTICS:
        (A) LENGTH: 411 amino acids
        (B) TYPE: amino acid
50
        (C) STRANDEDNESS: single
        (D) TOPOLOGY: linear
       (ii) MOLECULE TYPE: peptide
       (iii) HYPOTHETICAL: NO
55
       (iv) ANTISENSE: NO
       (v) FRAGMENT TYPE: internal
       (vi) ORIGINAL SOURCE:
       (xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:
60
```

```
35
                     40
    Xaa Xaa Val Phe Leu Pro Pro Arg Arg Glu His Met Cys Thr Ser Asn
                 55
                               6.0
    5
    90
    105
    Xaa Xaa Xaa Xaa Xaa Xaa Xaa Ala Met Cys Arg Ala Val Arg Tyr
10
                     120
    Ser Phe Ala Asp Leu Gly Asp Ile Ile Arg Gly Arg Asp Met Trp Asp
                   135
                               140
   15
                150
    170
   Xaa Xaa Xaa Xaa Lys Lys Pro Ala Tyr Lys Lys Leu Arg Ala Asp
                       185
   Trp Trp Glu Ala Asn Arg His Gln Val Trp Arg Ala Met Lys Cys Ala
20
                     200
    Thr Xaa Xaa Xaa Xaa Cys Xaa Xaa Xaa Xaa Xaa Xaa Xaa Ile Pro
                  215
                               220
   Gln Arg Leu Arg Trp Met Thr Glu Trp Ala Glu Trp Tyr Cys Lys Ala
25
                230
                            235
   Gln Ser Gln Glu Tyr Asp Lys Leu Lys Lys Ile Cys Xaa Xaa Xaa
                          250
   265
30
   Lys Cys Lys Ala Ala Cys Asp Lys Tyr Lys Glu Glu Ile Glu Lys Trp
                     280
   Asn Glu Gln Trp Arg Lys Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
                  295
                               300
   35
                310
                            315
   330
   345
   40
                     360
   Xaa Xaa Xaa Xaa Xaa Xaa Cys Xaa Xaa Xaa Xaa Xaa Xaa Xaa
                  375
                               380
   45
                390
                            395
   Xaa Xaa Xaa Xaa Xaa Xaa Cys Xaa Xaa Cys
             405
         (2) INFORMATION FOR SEQ ID NO:33:
50
       (i) SEQUENCE CHARACTERISTICS:
        (A) LENGTH: 311 amino acids
        (B) TYPE: amino acid
        (C) STRANDEDNESS: single
55
        (D) TOPOLOGY: linear
       (ii) MOLECULE TYPE: peptide
       (iii) HYPOTHETICAL: NO
       (iv) ANTISENSE: NO
60
       (v) FRAGMENT TYPE: internal
       (vi) ORIGINAL SOURCE:
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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:

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10
    Xaa Xaa Xaa Xaa Xaa Ala Cys Met Pro Pro Arg Arg Gln Lys Leu
            20
    5
                       40
    60
    10
    Xaa Xaa Xaa Xaa Xaa Xaa Xaa Gln Phe Leu Arg Ser Met Met
                            90
    Tyr Thr Phe Gly Asp Tyr Arg Asp Ile Cys Leu Asn Thr Asp Ile Ser
            100
                         105
                                       110
    Lys Lys Gln Asn Asp Val Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa Xaa
15
                      120
    Xaa Xaa Xaa Xaa Ser Lys Ser Pro Ser Gly Leu Ser Arg Gln Glu
                    135
    Trp Trp Lys Thr Asn Gly Pro Glu Ile Trp Lys Gly Met Leu Cys Ala
                 150
                               155
20
    165
                            170
                                         175
    180
                         185
                                       190
    Xaa Xaa Xaa Xaa Xaa Lys Pro Gln Phe Leu Arg Trp Met Ile Glu
25
         195
                      200
                                    205
    Trp Gly Glu Glu Phe Cys Ala Glu Arg Gln Lys Lys Glu Asn Ile Ile
      210
                    215
                                 220
    230
                               235
30
    Lys His Arg Cys Asn Gln Ala Cys Arg Ala Tyr Gln Glu Tyr Val Glu
              245
                            250
                                         255
    260
                         265
                                       270
    35
                      280
    295
    Xaa Xaa Xaa Cys Xaa Cys
                 310
40
          (2) INFORMATION FOR SEQ ID NO:34:
        (i) SEQUENCE CHARACTERISTICS:
         (A) LENGTH: 7 amino acids
45
         (B) TYPE: amino acid
         (C) STRANDEDNESS: single
         (D) TOPOLOGY: linear
        (ii) MOLECULE TYPE: peptide
50
        (iii) HYPOTHETICAL: NO
        (iv) ANTISENSE: NO
        (v) FRAGMENT TYPE: N-terminal
        (vi) ORIGINAL SOURCE:
55
        (xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:
    Pro Arg Arg Gln Xaa Leu Cys
60
          (2) INFORMATION FOR SEQ ID NO:35:
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(i) SEQUENCE CHARACTERISTICS:(A) LENGTH: 20 base pairs(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

```
(D) TOPOLOGY: linear
             (ii) MOLECULE TYPE: cDNA
             (iii) HYPOTHETICAL: NO
 5
             (iv) ANTISENSE: NO
             (v) FRAGMENT TYPE:
             (vi) ORIGINAL SOURCE:
             (xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:
10
      CCRAGRAGRC AARAAYTATG
                                                                           20
                (2) INFORMATION FOR SEQ ID NO:36:
15
             (i) SEQUENCE CHARACTERISTICS:
               (A) LENGTH: 20 base pairs
               (B) TYPE: nucleic acid
               (C) STRANDEDNESS: single
               (D) TOPOLOGY: linear
20
             (ii) MOLECULE TYPE: cDNA
             (iii) HYPOTHETICAL: NO
             (iv) ANTISENSE: NO
             (v) FRAGMENT TYPE:
25
             (vi) ORIGINAL SOURCE:
             (xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:
      CCSMGSMGSC AGCAGYTSTG
                                                                         . 20
30
                (2) INFORMATION FOR SEQ ID NO:37:
             (i) SEQUENCE CHARACTERISTICS:
               (A) LENGTH: 7 amino acids
35
               (B) TYPE: amino acid
               (C) STRANDEDNESS: single
               (D) TOPOLOGY: linear
             (ii) MOLECULE TYPE: peptide
40
             (iii) HYPOTHETICAL: NO
             (iv) ANTISENSE: NO
             (v) FRAGMENT TYPE: N-terminal
             (vi) ORIGINAL SOURCE:
45
            (xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:
      Phe Ala Asp Xaa Xaa Asp Ile
50
                (2) INFORMATION FOR SEQ ID NO:38:
             (i) SEQUENCE CHARACTERISTICS:
               (A) LENGTH: 20 base pairs
               (B) TYPE: nucleic acid
55
               (C) STRANDEDNESS: single
               (D) TOPOLOGY: linear
            (ii) MOLECULE TYPE: cDNA
            (iii) HYPOTHETICAL: NO
60
            (iv) ANTISENSE: NO
            (v) FRAGMENT TYPE:
            (vi) ORIGINAL SOURCE:
            (xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:
```

	TTTGCWGATW WWSGWGATAT	20
	(2) INFORMATION FOR SEQ ID NO:39:	
5	<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 20 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: linear</li> </ul>	• •
10	(ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO	
15	(iv) ANTISENSE: NO (v) FRAGMENT TYPE: (vi) ORIGINAL SOURCE:	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:39:	
20	TTCGCSGATW WCSGSGACAT	20
•	(2) INFORMATION FOR SEQ ID NO:40:	
25	<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 6 amino acids</li> <li>(B) TYPE: amino acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: linear</li> </ul>	
30	<pre>(ii) MOLECULE TYPE: peptide (iii) HYPOTHETICAL: NO (iv) ANTISENSE: NO (v) FRAGMENT TYPE: N-terminal (vi) ORIGINAL SOURCE:</pre>	
35	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:40:	
	Pro Gln Phe Xaa Arg Trp 1 5	
40	(2) INFORMATION FOR SEQ ID NO:41:	
45	<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 18 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
50	(ii) MOLECULE TYPE: cDNA (iii) HYPOTHETICAL: NO (iv) ANTISENSE: NO (v) FRAGMENT TYPE: (vi) ORIGINAL SOURCE:	
55	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:41:	
<b></b>	CCAWCKKARR AATTGWGG	18
	(2) INFORMATION FOR SEQ ID NO:42:	
60	<ul><li>(i) SEQUENCE CHARACTERISTICS:</li><li>(A) LENGTH: 18 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li></ul>	
65	(D) TOPOLOGY: linear	

```
(ii) MOLECULE TYPE: cDNA
             (iii) HYPOTHETICAL: NO
             (iv) ANTISENSE: NO
             (v) FRAGMENT TYPE:
 5
             (vi) ORIGINAL SOURCE:
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      CCASCKGWAG AWCTGSGG
                                                                             18
10
                (2) INFORMATION FOR SEQ ID NO:43:
             (i) SEQUENCE CHARACTERISTICS:
               (A) LENGTH: 7 amino acids
15
               (B) TYPE: amino acid
               (C) STRANDEDNESS: single
               (D) TOPOLOGY: linear
             (ii) MOLECULE TYPE: peptide
20
             (iii) HYPOTHETICAL: NO
             (iv) ANTISENSE: NO
             (v) FRAGMENT TYPE: N-terminal
             (vi) ORIGINAL SOURCE:
25
             (xi) SEQUENCE DESCRIPTION: SEQ ID NO:43:
      Glu Trp Gly Xaa Xaa Xaa Cys
                         5 '
30
                (2) INFORMATION FOR SEQ ID NO:44:
             (i) SEQUENCE CHARACTERISTICS:
               (A) LENGTH: 20 base pairs
               (B) TYPE: nucleic acid
35
               (C) STRANDEDNESS: single
               (D) TOPOLOGY: linear
             (ii) MOLECULE TYPE: cDNA
             (iii) HYPOTHETICAL: NO
40
             (iv) ANTISENSE: NO
             (v) FRAGMENT TYPE:
             (vi) ORIGINAL SOURCE:
             (xi) SEQUENCE DESCRIPTION: SEQ ID NO:44:
45
      CAAWAWTCWT CWCCCCATTC
                                                                             20
                (2) INFORMATION FOR SEQ ID NO:45:
50
             (i) SEQUENCE CHARACTERISTICS:
               (A) LENGTH: 20 base pairs
               (B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
55
             (ii) MOLECULE TYPE: cDNA
             (iii) HYPOTHETICAL: NO
             (iv) ANTISENSE: NO
             (v) FRAGMENT TYPE:
60
             (vi) ORIGINAL SOURCE:
             (xi) SEQUENCE DESCRIPTION: SEQ ID NO:45:
      CAGWASTCST CSCCCCACTC
                                                                             20
65
```

#### WE CLAIM:

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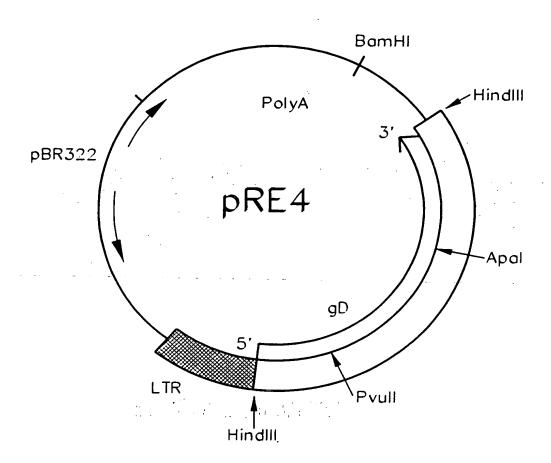
- 1. A composition comprising a nucleotide sequence of the *DBL* gene family, wherein said nucleotide sequence is selected from the group consisting of the *var-1*, *var-2*, *var-3* and *var-7* genes.
- 2. The composition of Claim 1, wherein the nucleotide sequence of the var-1, var-2, var-3 or var-7 gene encodes a cysteine-rich domain homologous to a cysteine-rich domain of a Duffy Antigen Binding Protein (DABP) derived from *Plasmodium vivax* and a Sialic Acid Binding Protein (SABP) derived from *Plasmodium falciparum*.
- 3. The composition of Claim 1, wherein the nucleotide sequence of the *var-1*, *var-2*, *var-3* or *var-7* gene encodes a cysteine-rich interdomain region between a first domain and a second domain.
- 4. The composition of Claim 1, wherein the nucleotide sequence is derived from a coding region of SEQ ID NO:13 or SEQ ID NO:15.
  - 5. A composition comprising a polypeptide encoded by a nucleotide sequence of the *DBL* gene family, wherein said polypeptide is encoded by a *var-1*, *var-2*, *var-3* or *var-7* gene.
  - 6. The composition of claim 5, wherein the polypeptide comprises a sequence of amino acid residues homologous to cysteine-rich domains of a Duffy Antigen Binding Protein (DABP) derived from *Plasmodium vivax* and a Sialic Acid Binding Protein (SABP) derived from *Plasmodium falciparum*.
  - 7. The composition of claim 5, wherein the polypeptide comprises a sequence of about 300 to 400 amino acid residues occuring in the cysteine-rich interdomain region between a first domain and a second domain of a polypeptide encoded by the *var-1*, *var-2*, *var-3* or *var-7* gene.
- 8. The composition of claim 5, wherein the polypeptide comprises a sequence of amino acid residues of SEQ ID NO:14 or SEQ ID NO:16.
  - 9. The composition of claim 5, wherein the polypeptide comprises a sequence of about 50 to about 325 amino acid residues of SEQ ID NO:14 or SEQ ID NO:16.
  - 10. The composition of claim 5, wherein the polypeptide comprises a sequence of about 75 to about 300 amino acid residues of SEQ ID NO:14 or SEQ ID NO:16.
  - 11. The composition of claim 5, wherein the polypeptide comprises a sequence of about 100 to about 250 amino acid residues of SEQ ID NO:14 or SEQ ID NO:16.
  - 12. The composition of claim 5, further comprising a pharmaceutically acceptable carrier and an isolated Duffy Antigen Binding Protein (DABP) binding domain polypeptide, a Sialic Acid Binding Protein (SABP) binding domain polypeptide, or a combination thereof, in an amount sufficient to induce a protective immune response to *Plasmodium* merozoites in a mammal.
  - . 13. The composition of any of the preceding claims for use in inducing a protective immune response to *Plasmodium* merozoites in a mammal.
  - 14. Use of the composition of any one of claims 1-12 in the preparation of a medicament for inducing a protective immune response to *Plasmodium* merozoites in a mammal.
  - 15. A method of inducing a protective immune response to *Plasmodium* merozoites in a mammal, comprising administering to a mammal an immunologically effective amount of a pharmaceutical composition

comprising a pharmaceutically acceptable carrier and an isolated cysteine-rich polypeptide encoded by a *var* gene selected from the group of genes consisting of *var-1*, *var-2*, *var-3* and *var-7* genes.

16. The method of claim 15, further comprising administering to said mammal an immunologically effective amount of a Duffy Antigen Binding Protein (DABP) binding domain polypeptide, a Sialic Acid Binding Protein (SABP) binding domain polypeptide, or a combination thereof.

K11- K17- 615- [17-	137- 130- 132- 15-	LKEKC $x_{11}$ C- $x_{1}$ PKSKC $x_{8}$ C- $x_{0}$ LKVEC $x_{4}$ C- $x_{1}$ LEKIC $x_{4}$ C- $x_{1}$	LQKQCX ₆ C-X ₁ VKRNCX ₁₂ C-X ₂ LKKICX ₁₁ C-X ₆ IKDACX ₈ C-X ₆ LLAKCX ₁₁ C-X ₆		<del>-</del> ·
DFCKDIRWSLGDFGDIIMGTDMEGIGYSK-X ₁₁ -KFCNDLKNSELDYGHLAMGNDMDFGGYST-X ₁₇ -EVCKIINKTEADIRDIIGGTDYWNDLSNR-X ₁₅ -KICNAILGSYADIGDIVRGLDVWRDINTN-X ₁₇ -	DIVRGKDLYLGYDNK-X DIVRGRDLYLGNPQE-X DIIRGRDMWDEDKSS-X DICLNTDISKKQNDV-X DICLGTDISSKKDTS-X	REMGRDXVSELPTEVQK KEMHGEELLERDNRSKL SEMGDDXCQDKTKMIET KEMGDEECEEMGTEVKQ	EEWAEDECRKKKKLENI EEWAEDECRKKNKKIKDI TEWAEWYCKAQSQEYDKI IEWGEBECABROKKENII	FIG. 1	
$ \begin{array}{llllllllllllllllllllllllllllllllllll$	C-X ₁₅ -C-X ₁₅ -ACAPYRRLHLCDYNL-X ₄₃ -QLCTVLARSEADIGDIVRGKDLYLGYDNK-X ₃₇ -C-X ₁₅ -ACAPYRRLHVCDQNL-X ₄₅ -QICTMLARSEADIGDIVRGRDLYLGNPQR-X ₃₀ -C-X ₁₁ -VFLPPRRHMCTSNL-X ₅₅ -AMCRAVRYSEADLGDIIRGRDMWDEDKSS-X ₃₂ -C-X ₁₀ -C-X ₁₀ -ACMPPRRQKLCLYYI-X ₅₂ -QFLRSMMYTEGDYRDICLNTDISKKQNDV-X ₁₅ -C-X ₁₀ -C-X ₁₁ -ACIPPRRQKLCLYYI-X ₅₁ -DFKRQMFYTEADYRDICLGTDISSKKDTS-X ₁₅ -C-X ₁₀ -C-X ₁₁ -ACIPPRRQKLCLHYL-X ₅₁ -DFKRQMFYTEADYRDICLGTDISSKKDTS-X ₁₅ -	TDRKAQQRRKQMMNBSKAQIMTAMMYSV- $x_{11}$ -C- $x_{8}$ ePQIYRMIREMGRDXVSELPTEVQKLKEKC $x_{11}$ C- $x_{1}$ Sehkiknfrkemmnsprekengerlysel- $x_{6}$ C- $x_{6}$ C- $x_{0}$ Nkkndklfrdemmsprekengerlysel- $x_{6}$ C- $x_{0}$ Nkkndklfrdemmyvikkdvmnvishve- $x_{5}$ C- $x_{7}$ IPQFFRMFSEMGDDXCQDKTKMIETLKVEC $x_{4}$ C- $x_{1}$ Kkqndnnernkmmekqrnlimssmvkhi- $x_{5}$ C- $x_{6}$ IPQFLRMLKEMGDECEEMGTEVKQLEKIC $x_{4}$ C- $x_{1}$	KGGDFFQLREDMMTSNRETVWKALICHA-X ₁₁ -C-X ₂₃ -VPQYLRWFEEWAEDECRKKKKKLENLQKQCX ₆ C-X ₁₅ - NDPRFFKLREDMMTANRETVWKAITCNA-X ₉ C-X ₂₃ -VPQYLRWFEEWAEDECRKKNKKIKDVKRNCX ₁₂ C-X ₂₂ - KKPAYKKLRADMMEANRHQVWRAMKCAT-X ₄ C-X ₈ IPQRLRWMTEWAEWYCKAQSQEYDKLKKICX ₁₁ C-X ₆ SKSPSGLSRQEMWKTNGPBIHKGMLCAL-X ₃ KPQFLRWMIEWGEECABRQKKENIIKDACX ₈ C-X ₃ KISNSIRYRKSWWETNGPVIHEGMLCAL-X ₄ 2RPQFLRWLTEWGENECKEQKKEYKVLLAKGX ₁₁ C-X ₃	HITRKKN-X56CXC HIIRSKF-X41-C-X7CXC HISKKKK-X36-C-X20CXX-C HIKERKN-X38-C-X19CXX-C	CTNCSVWCRMXET HIDNQKK- $X_{6}$ $G$
C-X ₁₂ -C-X ₅ VCIPDRRYQLCMKEL-X ₄₇ -C-X ₁₀ -C-X ₉ VCIPDRRIQLCIVNL-X ₃₆ -C-X ₁₃ -C-X ₁₀ -VCVPPRRQELCLGNI-X ₃₆ -C-X ₁₂ -C-X ₁₁ -VCGPPRRQQLCLGYI-X ₃₆ -	C-X ₁₅ -C-X ₁₅ -ACAPYRRLI C-X ₁₇ -C-X ₃₁ -VFLPPRREI C-X ₁₀ -C-X ₁₀ -ACMPPRRQI C-X ₁₀ -C-X ₁₁ -ACIPPRRQI	TDEKAQORRKOMENESKAQIE SEHKIKNFRKEMENEFREKLE NKKNDKLFRDEMEKVIKKDVE KKONDNNERNKEMEKORNLIH	KGGDFFQLREDHMTSNRETVH NDPEFFKLREDHMTANRETVH KKPAYKKLRADHHEANRHQVH SKSPSGLSRQEHMKTNGPBIH KISNSIRYRKSHHETNGPVIH	VPPCQNACKGYDQ HITRI EKECIDPCMKYRD HIIRS DDNCKSKCNSYKE HISKI EKKCKNACSSYEK MIKER	CTNCSVWCRMYET MIDNG CISCLYACNPYVD MINNG CGKCKAACDKYKERIBKHNEGM KHRCNQACRAYQE YVENK CVACKDQCKQYHS MIGIW
DABP SABP F1 SABP F2 EBL-e1	EBL-e2 Proj3 F1 Proj3 F2 Proj3 F3 E31a	DABP SABP F1 SABP F2 EBL-e1	EBL-e2 Proj3 R1 Proj3 F2 Proj3 F3 E31a	DABP SABP F1 SABP F2 EBL-e1	EBL-e2 Proj3 F1 Proj3 F2 Proj3 F3
Family 1	Pamily 2	Family 1 Cont'd	Family 2 Cont'd	Family 1 Cont'd	Family 2 Cont'd
	CIIDC	THITE CHEET	(2C 3 HIS)		

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F/G. 2

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# FIG. 3

Concensus amino acid sequences and the synthetic oligonucleotide primers designed from them.

UNIEBP5 and 5A: PRRQ K/E L C

UNIEBP5, for A+T biased codon usage: CC(A/G)-AG(G/A)-AG(G/A)-CAA-(G/A)AA-(C/T)TA-TG

UNIEBP5A, for G+C biased codon usage: CC(C/G)-(C/A)G(C/G)-(C/A)G(C/G)-CAG-CAG-(C/T)T(C/G)-TG

UNIEBP5 B and C: F A D I/Y G/R D I

UNIEBP5B, for A+T biased codon usage: TTT-GC(A/T)-GAT-(A/T)(A/T)-(G/C)G(A/T)-GAT-AT

UNIEBP5C, for G+C biased codon usage: TTC-GC(G/C)-GAT-(A/T)(A/T)C-(G/C)G(G/C)-GAC-AT

UNIEBP3 and 3A: P Q F L/F R W

UNIEBP3, for A+T biased codon usage: CCA-(A/T)C(T/G)-(T/G)A(A/G)-(A/G)AA-TTG-(A/T)GG

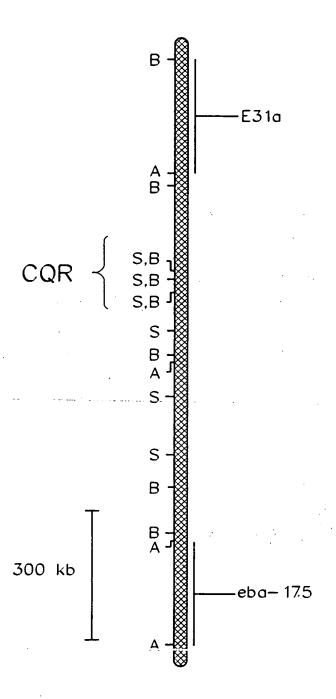
UNIEBP3A, for G+C biased codon usage: CCA-(C/G)C(G/T)-G(A/T)A-GA(A/T)-CTG-(C/G)GG

UNIEBP3 B and C: E W G D/E D/E Y/F C

UNIEBP3B, for A+T biased codon usage: CA-A(A/T)A-(A/T)TC-(A/T)TC-(A/T)CC-CCA-TTC

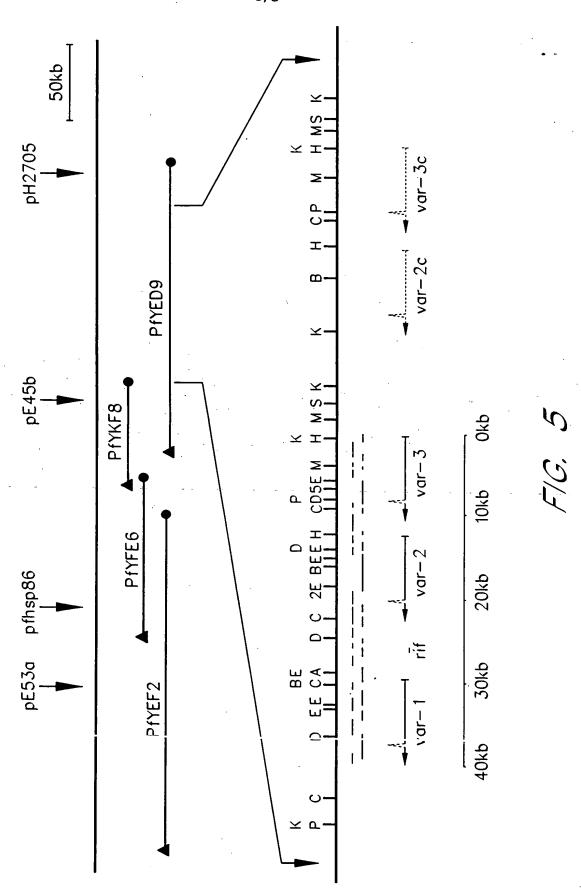
UNIEBP3C, for G+C biased codon usage: CA-G(A/T)A-(G/C)TC-(G/C)TC-(G/C)CC-CCA-CTC G+C Biased

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F/G, 4

# SUBSTITUTE SHEET (RULE 26)



**SUBSTITUTE SHEET (RULE 26)**